

Morphometric Study and Immunolocalization of Androgen Receptors in Epididymis During Postnatal Development in D'Man Lamb Reared Under Arid Environment in Algeria

Nouria BOUKENAOUİ-FERROUK^{1,2} Elara MOUDİLOU³ Zaina AMİRAT²
Jean-Marie EXBRAYAT³ Farida KHAMMAR²

¹ University Blida 1, Institute of Veterinary Sciences, BP 270 Road of Soumaa Blida, ALGERIA

² University of Science and Technology Houari Boumediene, Faculty of Biological Sciences, Arid Zone Research Laboratory, BP 44, Alger Gare, 16000, Algiers, ALGERIA

³ University of Lyon, UMRS 449, General Biology, Reproduction and Comparative Development Catholic University of Lyon, EPHE/PSL, 10 Place of Archives, F-69288 Lyon Cedex 02, FRANCE

Article Code: KVFD-2016-17255 Received: 16.12.2016 Accepted: 27.05.2017 Published Online: 29.05.2017

Citation of This Article

Boukenaoui-Ferrouk N, Moudilou E, Amirat Z, Exbrayat JM, Khammar F: Morphometric study and immunolocalization of androgen receptors in epididymis during postnatal development in D'Man lamb reared under arid environment in Algeria. *Kafkas Univ Vet Fak Derg*, 23 (5): 683-689, 2017. DOI: 10.9775/kvfd.2016.17255

Abstract

The aim of this study was to examine the morphometry and the immunolocalization of androgen receptors in the epididymis of D'Man lamb during postnatal development. The epididymis was collected at the slaughtering lamb, aged of 2 to 5 month. The weight of the epididymis increased with a significant difference at 3 months. The tubular diameter of the corpus and the cauda epididymal tubules increased respectively from 2 till 4 months, and from 2 till 5 months. The luminal diameters of the caput epididymal tubules increased significantly at 3 months. The luminal diameters of the corpus and cauda epididymal tubules increased significantly from 3 and 4 months respectively. The epithelial height of the caput and cauda epididymal tubules increased significantly at 5 months. The epithelial height of the corpus epididymal tubules increased significantly at 3 months. The androgen receptor immunostaining was localized in nuclei and cytoplasm of epithelial cells, smooth muscle cells and in the cytoplasm of interstitial cells of the epididymis at each age. In conclusion, both the morphometric changes and androgen receptors immunolocalization during the postnatal development of epididymis indicated the necessity of androgens for postnatal differentiation and maintaining the structure of the epididymis.

Keywords: Epididymis, Morphometry, Androgen receptor, Immunohistochemistry, Postnatal development, D'Man lamb

Cezayir'in Kurak İkliminde Yetiştirilen D'Man Kuzularında Postnatal Gelişim Süresince Epididimiste Androjen Reseptörlerinin İmmunolokalizasyonu ve Morfometrik Bir Çalışma

Özet

Bu çalışmanın amacı D'Man kuzularında postnatal gelişim süresince epididimiste androjen reseptörlerinin immunolokalizasyonunu ve morfometrisini araştırmaktır. 2 ile 5 ay arasında yaşları değişen kuzulardan kesim sonrasında epididimisler toplandı. Epididymis ağırlığı 3 aylık olanlarda anlamlı oranda farklı bulundu. Corpus ve cauda epididimal tüplerin tubular çapları sırasıyla 2'den 4 aylığa ve 2'den 5 aylığa kadar olanlarda artma gösterdi. Caput epididimal tüplerin luminal çapları 3 aylıklarda anlamlı oranda arttı. Corpus ve cauda epididimal tüplerin luminal çapları sırasıyla 3 ve 4 aylıklarda anlamlı oranda arttı. Caput ve cauda epididimal tüplerin epitel yükseklikleri 5 aylıklarda anlamlı oranda artış gösterdi. Corpus epididimal tüplerin epitel yükseklikleri 3 aylıklarda anlamlı oranda artış gösterdi. Androjen reseptör immunboyanması tüm yaş gruplarında epididimisin epitel hücrelerinin çekirdek ve sitoplazmalarında, düz kas hücrelerinde ve intersitysel hücrelerinin sitoplazmasında lokalize oldu. Sonuç olarak; morfometrik değişimler ve androjen reseptör immunolokalizasyonu epididimisin postnatal gelişimi ve oluşumu süresince androjenlerin gerekli olduğunu göstermiştir.

Anahtar sözcükler: Epididimis, Morfometri, Androjen reseptör, Immunohistokimya, Postnatal gelişim, D'Man kuzu



İletişim (Correspondence)



+213 55 7954637



nouria09@yahoo.fr

INTRODUCTION

Sheep is an important part of the Algerian agricultural economy. D'Man and other second breeds (Hamra, Barbarine, Sidahou, Tazegzawt) represent less than 1% of the Algerian sheep population. D'Man breed acquires its importance from its exceptional reproductive performances and its high adaptation to the oasian environment ^[1]. The productivity was consistently higher due to generally high fertility and prolificacy ^[2] with an early onset of puberty at 3 months ^[3].

The morphohistological change is essential to gain a comprehensive knowledge on the reproductive physiology of epididymis ^[4,5]. Indeed, epididymal functions can be divided into several general categories: concentration of sperm; functional maturation; storage in a quiescent state until ejaculation, removal of degenerating sperm, provision of appropriate conditions for survival, transport by the myoid cells, protection and maintenance of the blood epididymal barrier ^[6]. In most species, the epididymis is divided into the caput, corpus and cauda regions ^[4,5,7]. The activity of these regions is regulated by endocrine, lumicrine, and paracrine factors, the relative importance of which remaining a topic of investigation ^[8]. The presence of androgen receptors (ARs) during spermatogenesis was investigated in rodent models in which testosterone levels were chemically deleted, or in models with transgenic disruption of ARs ^[9]. The use of these models made it possible to identify the steps of spermatogenesis requiring ARs, specifically the maintenance of spermatogonia number, integrity of blood-testis barrier, completion of meiosis, adhesion of spermatids and spermiation. Together these studies detailed the essential nature of androgens in the promotion of male fertility ^[10]. Luminal factors from the testis, in addition to androgens, are important for both the epididymal development ^[11] and maintenance of adult tissues ^[12]. The presence of steroids and their receptors, specifically ARs, which are responsible for maintaining epididymal structure and functions throughout the postnatal development ^[13] has not yet been shown in the epididymis of D'Man lambs. In the prepubertal period several factors can cause epididymal obstruction, such as iatrogenicities due to inguinal herniotomies, inflammatory, tumoral, cystic and similar causes ^[14]. The effect of these conditions on the testes and epididymal ARs distribution are of major importance. In this study, the objective was to characterize the morphological normal changes and immunolocalization of ARs of the epididymis in 2 to 5 months old D'Man lamb.

MATERIAL and METHODS

Twelve lambs aged from 2 to 5 months reared at El Meniaa experimental station in Algeria (30° 34' N., 02° 52' E.) have been used for this study. For each month, three lambs were

weighed and immediately slaughtered; the epididymis was separated from testis and weighed. From each regions of epididymis (caput, corpus and cauda), a sample was fixed in 10% formaldehyde in phosphate buffered saline, dehydrated in a graded series of ethanol, clarified in xylene and embedded in paraffin. The sections were hydrated and stained with Masson's trichroma in order to study general histology. The diameters of tubules and lumen tubules of the epididymis were measured on 10 cross-sections per animal. The height of epithelial cells was measured from the basement to the apical membrane in cross-sections of 10 tubules using a computer program of light microscope Nikon Eclipse E 400 connected to a Nikon DXM 1200 digital camera.

Androgen Receptor Immunohistochemistry

The immunohistochemical studies of ARs were performed using the avidin-biotin complex method (ABC), with Vectastain Elite ABC kit (Vector Laboratories, Burlingame, CA, USA). Paraffin sections (3 µm thick) were deparaffinized, hydrated through a graded ethanol series (100%, 95% and 70%), and washed in PBS. Immunohistochemistry was performed on deparaffinized adjacent sections with heat-induced antigen retrieval in citrate buffer (pH 6.0) using *water bath* set at 94°C, as described in the prospectus for the kit "Vector Antigen Unmasking Solutions" (Vector Laboratories, CA, H3300). This step was followed with endogenous peroxidase blocking 3% H₂O₂ in PBS for 5 min at room temperature. All washes between antibody or reagent incubations were rinsed 5 min 2 times at room temperature in PBS, and all the incubations were carried out in a wet chamber. Tissue sections were first submitted to the appropriate serum in order to block the non-specific binding sites. After that, sections processed for ARs labeling were incubated with normal horse serum at room temperature for 5 min, and then with both avidin and biotin sites subsequently blocked (Vector Laboratories, CA, SP-2001). All sections were incubated overnight at 4°C with the primary antibody: a rabbit polyclonal antibody (C-19) raised against a peptide within the C-terminal domain of the human AR (sc-815, Santa Cruz Biotechnology, Santa Cruz, CA, USA), which was diluted at 1 : 200 in PBS. This antibody being currently used to detect the presence of androgen receptors in several mammals, it was used to detect androgen receptors in D'man lamb epididymis. Bound antibodies were visualized by incubating the sections with biotinylated secondary antibody (Vectastain Elite ABC kit-Vector Laboratories, CA, #PK-6200) for 30 min. Labeling of ARs was performed with 3,3'-diaminobenzidine-tetra-hydrochloride chromogenic substrate (SK-4100, DAB substrate kit for peroxidase; Vector Laboratories) and monitored microscopically. Sections were counterstained with hematoxylin (Hematoxylin QS, H-3404; Vector Laboratories, Burlingame, CA, USA). Those sections were dehydrated and mounted. Sections incubated with normal horse serum instead of primary

antibody were used as negative controls. Images were captured using a light microscope (Nikon Eclipse E 400 connected to a Nikon DXM 1200 digital camera).

The results of immunohistochemical staining, evaluated by semiquantitative methods, were given for the epithelial cells (principal, basal and apical cells), the interstitial stromal cells and the peritubular smooth muscle cells of each epididymis compartment. The staining intensity was evaluated at four different levels: ++ / strong, + / moderate, - / negative, +/- / variable.

Statistical Analysis

Results were expressed as Mean \pm SEM after verification of their homogeneity. The CV observed was below 20%. Analysis of variance was performed using the One-way ANOVA. Each parameter was analyzed by pairwise comparison using the Tukey HSD test. All analysis was performed using XLSTAT version 2016. The correlation between the average epididymis weight and age was analyzed using a Pearson's Linear Correlation test. $P < 0.05$ was considered as significant.

RESULTS

Epididymis Weight

The weight of the paired epididymis related to the age presented in Fig 1, increased ($P < 0.001$) from 2 to 5 months. At 2 month, the epididymis weight was 2.8 ± 0.3 g, and increased quickly to 10.7 ± 1.4 g at 5 month with a significant increase ($P < 0.05$) observed at 3 months. The average epididymal weight was significantly affected by the age ($P < 0.001$) and a positive correlation ($r = 0.76$, $P < 0.05$) was observed between the average epididymis weight and age (Fig. 1).

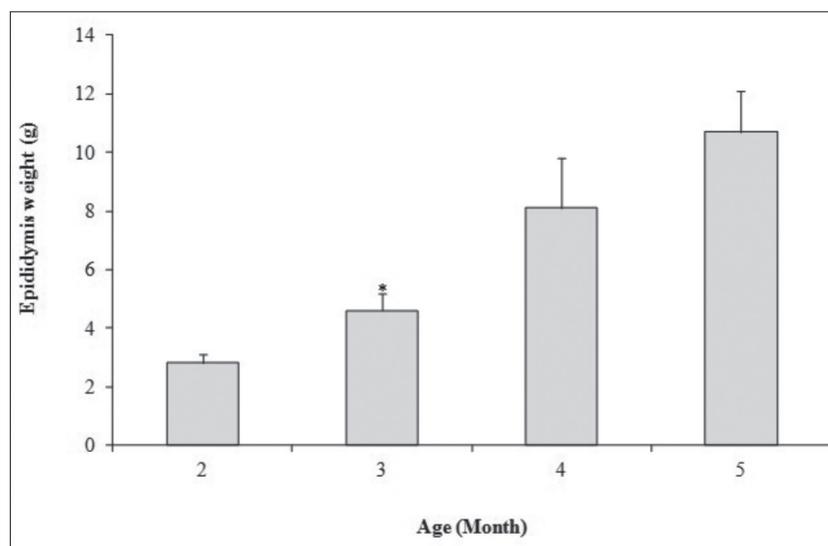


Fig 1. Age-related changes in the epididymis weight (g) in D'Man lamb

Morphometry Development of the Epididymis

The evaluation of the effects of the age on the tubular diameter, luminal diameter, and epithelial height of the epididymis is shown in Fig. 2. The measures of tubular and luminal diameters, and the epithelial height showed some regional differences in the three regions of the epididymis (Fig. 2).

The tubular diameter of the caput epididymal tubules presented a high increase ($P < 0.001$) from 2 (389 ± 9.7 μ m) to 3 (542 ± 11.3 μ m) months, then it stabilized at 5 months (550 ± 13.7 μ m). In 2 months-old animals, the epididymal tubular diameter was 279 ± 9.9 and 294 ± 9.9 μ m, respectively for the corpus and cauda epididymis, increased to 490 ± 7.5 and 467 ± 11.8 μ m respectively in 5 months-old animals. The tubular diameter of the corpus and the cauda epididymal tubules increased ($P < 0.001$) gradually, respectively between 2 to 4, and between 2 to 5 months (Fig. 2a). During the growth period, there were age effects ($P < 0.001$) on the tubular and luminal diameters of the epididymal tubules (Fig. 2a).

The luminal diameters of the caput epididymal tubules increased continuously with the age, with a significant ($P < 0.001$) increasing in 3 months-old animals (412 ± 12.8 μ m) (Fig. 2b). The epididymal luminal diameters measuring 180 ± 3.7 and 221 ± 7.9 μ m, respectively for the corpus and cauda epididymis in 2 months-old animals, increased to 366 ± 8.3 μ m and 373 ± 12.8 μ m in 5 months-old animals. The luminal diameters of the corpus and caudal epididymal tubules increased significantly ($P < 0.001$) from 3 and 4 months respectively (Fig. 2b). During the postnatal growth, there were age effects ($P < 0.001$) on luminal diameters of the epididymal tubules (Fig. 2b).

The epithelial height of the caput and caudal epididymal tubules remained unchanged between 2 till 4 months, then it increased significantly ($P < 0.001$) in 5 months-old animals (Fig. 2c). The epithelial height of the corpus epididymal tubules increased significantly ($P < 0.001$) in 3 months-old animals (Fig. 2c). During the postnatal growth, age effected ($P < 0.001$) the epithelial height of epididymal tubules (Fig. 2c).

Immunolocalization of Androgen Receptor During Epididymis Development

ARs immunostaining was observed in all the segments of lamb epididymis (Fig. 3). ARs were localized in the principal cells of the caput, corpus and cauda. The cytoplasm was slightly positive by comparison with nuclei in all the regions. In the epithelium, the ARs immun-expression was observed in the basal and

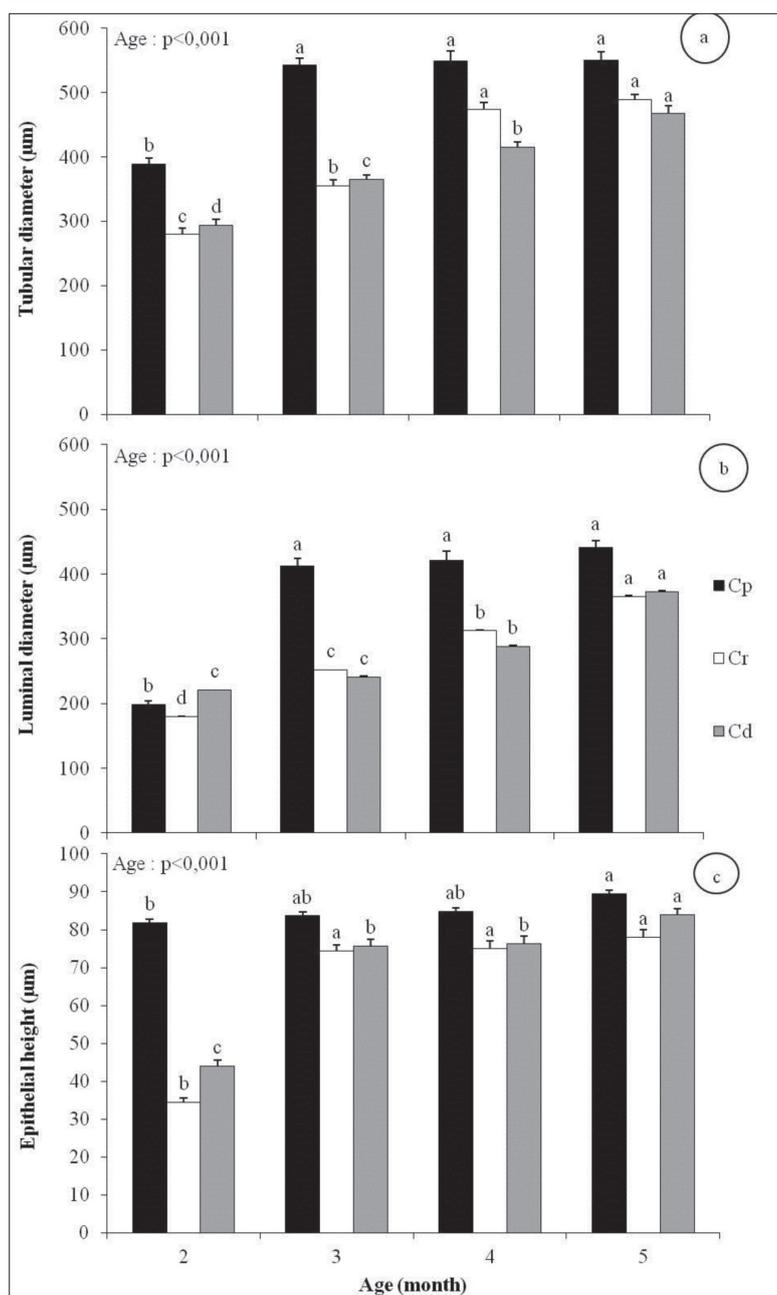


Fig 2. Effect of age on the tubular diameter (a), luminal diameter (b), and epithelium height (c) in epididymis. Bars represent means + SEM (n = 3). Cp: caput, Cr: corpus, Cd: cauda

apical cytoplasm of non-ciliated cells and in apical cytoplasm of ciliated cells.

In the corpus and cauda epididymis, the same pattern of ARs repartition was observed. However, a decreasing intensity of immun-expression was observed, with the lowest intensity in principal cells of corpus, and cauda epididymis (Table 1). The peritubular connective tissue was negative. Intermittent immunostainings for ARs were also seen in smooth muscle and connective tissue. Sperm in the lumen appeared positive for ARs immunostaining. Nuclei of cells belonging to connective tissue and smooth muscle cells were also positive. Additionally, nuclei of interstitial cell and sperm were positive for antibodies directed against ARs. Luminal sperm in corpus and cauda epididymis showed positive immunostaining for ARs in the cytoplasmic droplet.

No immunoreaction was observed in the caput, the corpus and the cauda epididymis incubated without any primary antibody. ARs staining in the epithelial cells appeared to be stronger than in the peritubular smooth muscle cells. In the epithelial cells, staining intensity was stronger in the principal cells than in basal and apical cells. The staining intensity of AR positive cells changed depending on the age of animal. The AR immunostaining intensity increased between 2 till 5 months for all the different regions of epididymis.

DISCUSSION

After this study, the epididymis weight increased quickly during the period comprised between 2 and 5 months. The same increase pattern in testis weight was observed in lambs in postnatal development [3]. Several authors reported epididymis weight was increasing with age [15,16].

Table 1. Average staining intensity of ARs in the different region of epididymis (caput, corpus and cauda) at 2 and 5 months of age during postnatal development in D'Man lamb

Epithelial Cells	2 Months			5 Months		
	Caput	Corpus	Cauda	Caput	Corpus	Cauda
Principal cells	+	+	+	++	++	++
Basal cells	-	-	-	+/-	+	+/-
Apical cells	+	+	+	++	++	++
Interstitial stromal cells	-	+/-	+	+	+/-	+/-
Peritubular smooth muscle cells	-	-	+/-	+/-	+	-

* Symbols are as follows: ++ strong, + moderate, - negative, +/- variable

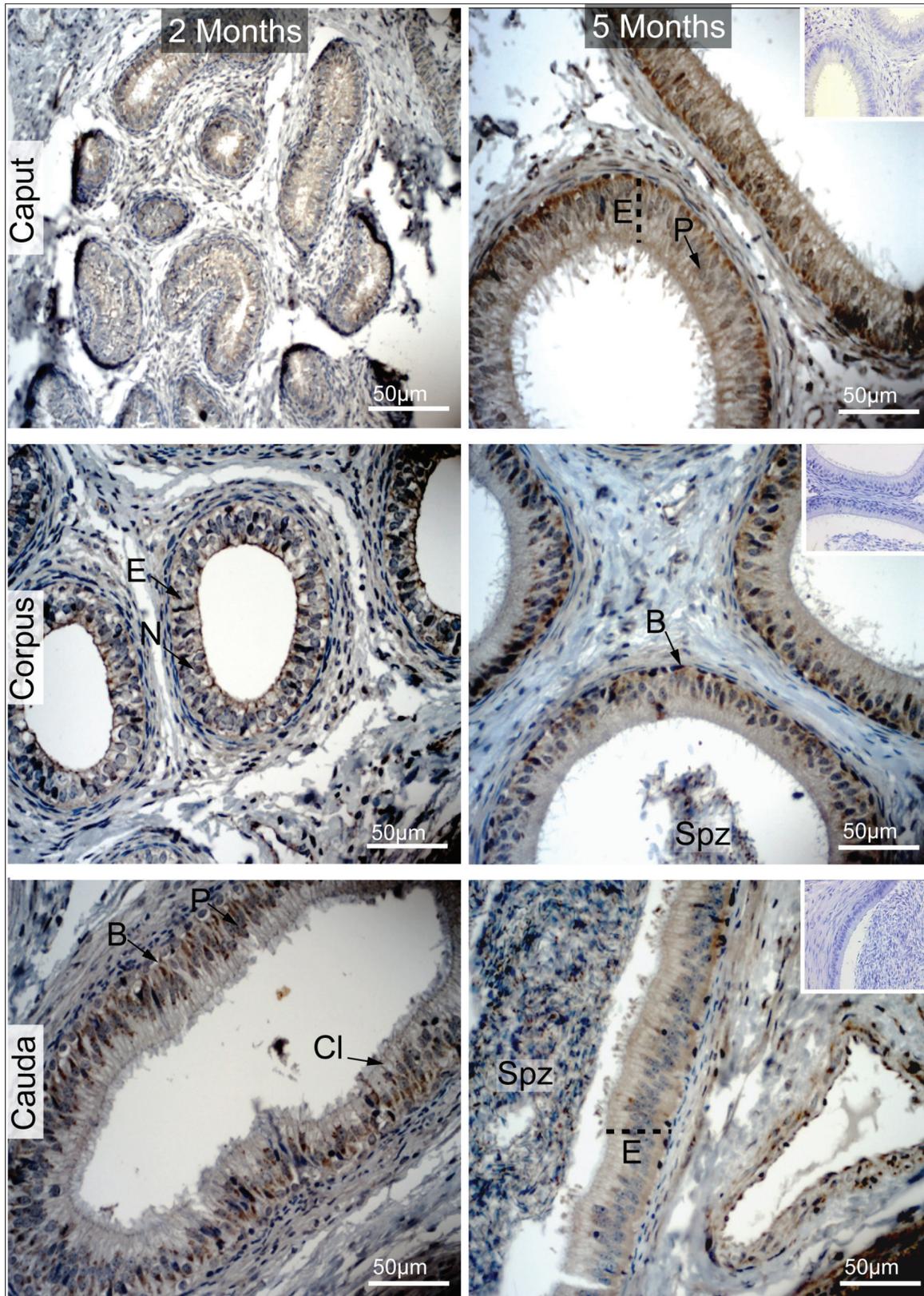


Fig 3. Immunolocalization of androgen receptor in the epididymis of D'Man lamb

AR immunostaining is observed in the nucleus and cytoplasm of epididymal epithelial cells in the caput, corpus and cauda in 2 and 5 months-old D'Man lambs. The images shown here are representative of the results of immunostaining observed in three animals for each age group. No immunostaining was observed in negative controls (inset). E: epithelial cell; P: principal cell; B: basal cell; N: narrow cell; Cl: clear cell; Spz: spermatozoa

At each age, the tubular diameter decreased from the caput to cauda in D'Man lamb. Noviana et al.^[17] reported in Kacang goats and local sheep that the diameter in the corpus was smaller than the caput and cauda regions, due to the narrowed and elongated anatomical structure of the corpus epididymis. The tubular diameter increased significantly with the age in the cauda, because spermatozoa were stocked in this epididymis area.

The luminal diameter of the caput epididymal tubules presented a high significant increase in 3 months-old animals. The epididymal luminal diameters decreased from caput then increased in cauda. For the domesticated adult African great cane rat (*Thryonomys swinderianus*)^[18] and in age-related study in the rat^[19], the epididymal luminal diameters increased progressively from caput to cauda. While the cauda epididymis acted as a sperm reservoir, both the caput and corpus were responsible for sperm maturation^[20].

The height of the epithelium of each region increased, as the age increased. The epithelial height of caput was more developed than in corpus and cauda epididymis. In One-Humped Camel (*Camelus dromedaries*)^[21], in rat^[19], the highest epithelium was seen in the caput and decreased gradually toward the cauda; wards the epididymal duct might mechanically facilitate the passage of the sperms toward the terminal segment^[21].

The development of a fully differentiated epithelium is dependent on androgens and also requires the influence of luminal factors from the testis^[22]. The epithelial cells of epididymis are able to synthesize some steroid hormones because the cytoplasm has accumulated lipid droplets and contains the active enzymes of steroidogenesis, capable to moderate the *in vitro* synthesis of androgens^[23]. Androgens play a crucial role in the proliferation, differentiation and function of the epididymis^[24]. Immunocytochemistry identified the epithelium of the epididymis such as a site of ARs expression, while the connective tissue stroma and the blood vessels lacked specific signals throughout the organ. Androgens are also implicated in the regulation of epididymal blood flow^[25]. The ARs immunostaining in D'Man lamb epididymis was observed in both the nuclei and cytoplasm of ciliated and non-ciliated epithelial cells, in addition to the peritubular and some stroma cells. The epididymal localization of the ARs was reported for various species^[26,27]. However, the presence of ARs epididymis is well documented in adult ram^[26-28] than during postnatal development of lambs. Additionally, sperm in the lumen appeared positive for ARs. The heterogeneous signal distribution for ARs expression along the ram epididymis did not change depending on age as reported in the rat^[29]. During postnatal development, the luminal secretion of androgens is essential for the maintenance of epithelial cell identity^[30], and for the normal development and function of the stromal cells^[31]. Gur and Timurkaan^[14] reported the progressive degenerative alterations occurred in

the seminiferous tubules after prepubertal epididymal ligation. These degenerative changes included increase at the seminiferous tubule diameter and basal membrane thickness, decrease at the germinal epithelium thickness, depletion of spermatids and presence of multinucleated spermatids^[32]. Both the regionalized differentiation of the epididymis and the variation in the luminal fluid composition take place under the control of androgens^[33].

The morphometric changes and immunolocalization of androgen receptors during the postnatal development of epididymis indicated the necessity of androgens for postnatal differentiation and maintaining the structure of the epididymis.

ACKNOWLEDGEMENTS

We would like to thank Pr. Djazouli Z. (Statistical traitement) and Dr. Djazouli Alim Z. (Image Analysis), University of Blida 1 and Ph.D. Mormede P., UMR 1388 INRA-INPT GenPhySE (France), for their helpful and Hadjadj H. and Hakoum B. for their assistance in this work at the Research Laboratory on Arid Zones (LRZA) in El-Menia station.

REFERENCES

- Darfaoui EM:** Workshop on developing breeding strategies for lower input animal production environments. In, Galal S, Boyazoglu J, Hammond K (Eds): D'man sheep Breeding Programme in Morocco. 319-329. ICAR, Villa del Ragnò, Via Nomentana 134, 00162 Rome, Italy, Bella, Italy, 1999.
- Lahlou-Kassi A, Berger YM, Bradford GE, Boukhliq R, Tibary A, Derqaoui L, Boujenane I:** Performance of D'Man and Sardi sheep on accelerated lambing I. Fertility, litter size, postpartum anoestrus and puberty. *Small Ruminant Res*, 2 (3): 225-239, 1989. DOI: 10.1016/0921-4488(89)90003-5
- Boukenaoui N, Moudilou E, Chevalier C, Amirat Z, Exbrayat JM, Khammar F:** Postnatal changes in testicular development and androgen receptors immunolocalization in D'Man ram lambs. *Folia Histochem Cytobiol*, 50, 38-45, 2012. DOI: 10.5603/FHC.2012.0005
- Kishore PVS, Ramesh G, Basha SH:** Postnatal differentiation and regional histological variations in the ductus epididymidis of rams. *Tamilnadu J Vet Anim Sci*, 8, 145-151, 2012.
- Elzoghby IMA, Sosa GA, Mona NAH, Manshawy AA:** Postnatal development of the epididymis in the sheep. *BVMJ*, 26 (1): 67-74, 2014.
- Marengo SR:** Maturing the sperm: Unique mechanisms for modifying integral proteins in the sperm plasma membrane. *Anim Reprod Sci*, 105, 52-63, 2008. DOI: 10.1016/j.anireprosci.2007.11.018
- Goyal HO:** Morphology of the bovine epididymis. *Am J Anat*, 172, 155-172, 1985. DOI: 10.1002/aja.1001720205
- Turner TT, Johnston DS, Finger JN, Jelinsky SA:** Differential gene expression among the proximal segments of the rat epididymis is lost after efferent duct ligation. *Biol Reprod*, 77, 165-171, 2007. DOI: 10.1095/biolreprod.106.059493
- Timurkaan S, Gur FM, Karan M:** Immunohistochemical distribution of androgen receptor in rat testis during postnatal development. *Revue Méd Vét*, 163 (3): 112-115, 2012.
- O'Hara L, Smith LB:** Androgen receptor roles in spermatogenesis and infertility. *Best Pract Res Clin Endocrinol Metab*, 29, 595-605, 2015. DOI: 10.1016/j.beem.2015.04.006
- Abe K, Takano H, Ito T:** Response of the epididymal duct in the corpus epididymidis to efferent or epididymal duct ligation in the mouse. *J Reprod Fertil*, 64 (1): 69-72, 1982.

- 12. Turner TT, Miller DW, Avery EA:** Protein synthesis and secretion by the rat caput epididymidis *in vivo*: Influence of the luminal microenvironment. *Biol Reprod*, 52 (5): 1012-1019, 1995.
- 13. Robaire B, Hamzeh M:** Androgen action in the epididymis. *J Androl*, 32, 592-599, 2011. DOI: 10.2164/jandrol.111.014266
- 14. Gur FM, Timurkaan S:** Effects of prepubertal epididymal ligation on the androgen receptor distribution of the rat testis. *Anal Quant Cytopathol Histopathol*, 34 (6): 317-324, 2012.
- 15. Rathje TA, Johnson RK, Lunstra DD:** Sperm production in boars after nine generations of selection for increased weight of testis. *J Anim Sci*, 73 (8): 2177-2185, 1995.
- 16. Pearl CA, Berger T, Roser JF:** Estrogen and androgen receptor expression in relation to steroid concentrations in the adult boar epididymis. *Domest Anim Endocrinol*, 33, 451-459, 2007. DOI: 10.1016/j.domaniend.2006.09.003
- 17. Noviana C, Boediono A, Wresdiyati T:** Morphology and histomorphometry of testis and epididymis of Kacang goat (*Capra* sp.) and local sheep (*Ovis* sp.). *Media Veteriner*, 7 (2): 12-16, 2000.
- 18. Olukole SG, Obayemi TE:** Histomorphometry of the testes and epididymis in the domesticated adult African great cane rat (*Thryonomys swinderianus*). *Int J Morphol*, 28 (4): 1251-1254, 2010.
- 19. Markey CM, Meyer GT:** A quantitative description of the epididymis and its microvasculature: An age-related study in the rat. *J Anat*, 180 (Pt 2): 255-262, 1992.
- 20. Sullivan R, Saez F, Girouard J, Frenette G:** Role of exosomes in sperm maturation during the transit along the male reproductive tract. *Blood Cells Mol Dis*, 35, 1-10, 2005. DOI: 10.1016/j.bcmd.2005.03.005
- 21. Zayed AE, Aly K, Ibrahim IA, Abd El-Maksoud FM:** Morphological studies on the epididymal duct of the one-humped camel (*Camelus dromedaries*). *Open J Vet Med*, 2, 245-254, 2012. DOI: 10.4236/ojvm.2012.24040
- 22. Rodríguez C M, Kirby JL, Hinton BT:** The Development of the Epididymis. In, Robaire B, Hinton BT (Eds): *The Epididymis: From Molecules to Clinical Practice A Comprehensive Survey of the Efferent Ducts, the Epididymis and the Vas Deferens*. Springer US ed., 251-267, Kluwer Academic/Plenum Publishers, New York, 2002.
- 23. Wiszniewska B:** Steroidogenic characteristics of *in vitro* cultured epididymal epithelial cells of the rat. *Reprod Biol*, 1 (1): 60-66, 2001.
- 24. Hess RA, Bunick D, Lee KH, Bahr J, Taylor JA, Korach KS, Lubahn DB:** A role of oestrogen in the male reproductive system. *Nature*, 390: 509-512, 1997. DOI: 10.1038/37352
- 25. Brown PD, Waites GM:** Regional blood flow in the epididymis of the rat and rabbit: effect of efferent duct ligation and orchidectomy. *J Reprod Fertil*, 28, 221-233, 1972. DOI: 10.1530/jrf.0.0280221
- 26. Carreau S, Drosdowsky MA, Courot M:** Androgen-binding proteins in sheep epididymis: Characterization of a cytoplasmic androgen receptor in the ram epididymis. *J Endocrinol*, 103, 273-279, 1984. DOI: 10.1677/joe.0.1030273
- 27. Tekpetey FR, Veeramachaneni DN, Amann RP:** Localization of androgen receptors in ram epididymal principal cells. *J Reprod Fertil*, 87, 311-319, 1989. DOI: 10.1530/jrf.0.0870311
- 28. Jegou B, Dacheux JL, Terqui M:** Demonstration of a specific androgen binding protein (ABP) in the seminal plasma of the ram. *C R Acad Sci Hebd Seances Acad Sci D*, 286 (4): 347-350, 1978.
- 29. Gür FM, Timurkaan S:** Immunohistochemical localization of androgen receptor in rat caput epididymis during postnatal development. *J Clin Exp Invest*, 2011, 260-266, 2011. DOI: 10.5799/ahinjs.01.2011.03.0051
- 30. O'Hara L, Welsh M, Saunders PT, Smith LB:** Androgen receptor expression in the caput epididymal epithelium is essential for development of the initial segment and epididymal spermatozoa transit. *Endocrinol*, 152, 718-729, 2011. DOI: 10.1210/jcem.96.2.zeg555a
- 31. Hess RA, Zhou Q, Nie R, Oliveira C, Cho H, Nakaia M, Carnes K:** Estrogens and epididymal function. *Reprod Fertil Dev*, 13, 273-283, 2001. DOI: 10.1071/RD00100
- 32. Gur FM, Timurkaan S:** The effects of prepubertal epididymal ligation upon the rat testis. *Iran J Reprod Med*, 12 (10): 673-680, 2014.
- 33. Toshimori K:** Biology of spermatozoa maturation: An overview with an introduction to this issue. *Microsc Res Tech*, 61, 1-6, 2003. DOI: 10.1002/jemt.10311