

## Effect of Fish Oil Addition to the Skim Milk-egg Yolk Extender on the Quality of Frozen-thawed Bali Bull Spermatozoa <sup>[1]</sup>

Abdul MALIK <sup>1</sup>  Muhammad SYARIFDJAYA <sup>1</sup> Aam GUNAWAN <sup>1</sup>  
Siti ERLINA <sup>1</sup> Achmad JAELANI <sup>1</sup> Dadang Budi WIBOWO <sup>2</sup>

<sup>[1]</sup> This study was supported by funding routine from Islamic University of Kalimantan

<sup>1</sup> Department of Animal Science, Faculty of Agriculture, Islamic University of Kalimantan, Banjarmasin, South Kalimantan, INDONESIA

<sup>2</sup> Officer of Animal husbandry, District of Balangan, Province South Kalimantan, INDONESIA

Article Code: KVFD-2017-17556 Received: 14.02.2017 Accepted: 14.04.2017 Published Online: 19.04.2017

### Citation of This Article

Malik A, Syarifdjaya M, Gunawan A, Erlina S, Jaelani A, Wibowo DB: Effect of fish oil addition to the skim milk-egg yolk extender on the quality of frozen-thawed Bali bull spermatozoa. *Kafkas Univ Vet Fak Derg*, 23 (4): 651-654, 2017. DOI: 10.9775/kvfd.2017.17556

### Abstract

This study was conducted to investigate the effect of fish oil addition to the semen extender on post-thaw quality of Bali bull spermatozoa. Fish oil was added at the doses of 0 (control), 50, 100, 150 and 200 mg/100 mL to the skim milk-egg yolk extender. Supplementation of fish oil at the dose of 100 mg/mL significantly ( $P<0.05$ ) increased both live sperm rate and motility, but its dose of 200 mg/mL caused a significant ( $P<0.05$ ) decrease in the live sperm rate when compared with the control group. A significant ( $P<0.05$ ) increment was observed in abnormality rate of frozen-thawed spermatozoa in comparison with the control group when all doses of fish oil except 50 mg/100 mL were added. In conclusion, addition of 100 mg fish oil to 100 mL of skim milk-egg yolk extender could be beneficial for the improvement of the quality of Bali bull spermatozoa after freeze-thawing.

**Keywords:** Bull semen, Fish oil, Cryopreservation, Live sperm, Motility

## Süt Tozu-Yumurta Sarısı Sulandırıcısına Balık Yağı İlavesinin Dondurulmuş-Çözdürülmüş Bali Boğa Spermatozoonlarının Kalitesi Üzerine Etkisi

### Özet

Bu çalışma sperma sulandırıcısına katılan balık yağının Bali boğa spermatozoonlarının çözme sonrası kalitesi üzerine etkisini araştırmak amacıyla yapıldı. Süt tozu-yumurta sarısı sulandırıcısına 0 (kontrol), 50, 100, 150 ve 200 mg/100 mL dozunda balık yağı ilave edildi. 100 mg/100 mL balık yağı ilavesi kontrol grubu ile karşılaştırıldığında hem canlı sperm oranını hem de motiliteyi önemli derecede ( $P<0.05$ ) artırdı, ancak 200 mg/100 mL'lik doz canlı sperm oranında önemli ( $P<0.05$ ) bir azalmaya neden oldu. 50 mg/100 mL'lik doz hariç balık yağının tüm dozları ilave edildiğinde kontrol grubuna kıyasla dondurulmuş-çözdürülmüş spermatozoonların anormallik oranında önemli ( $P<0.05$ ) bir artış gözlemlendi. Sonuç olarak, süt tozu-yumurta sarısı sulandırıcısının 100 mL'sine 100 mg balık yağı ilavesi Bali boğa spermatozoonlarının dondurma-çözme sonrası kalitesinin artırılması için faydalı olabilir.

**Anahtar sözcükler:** Boğa sperması, Balık yağı, Kriyoprezervasyon, Canlı sperm, Motilite

### INTRODUCTION

Artificial insemination (AI) is an important tool for genetic improvement and has effectively been used in several livestock species including cattle, sheep and goats. One of the factors affecting the success of AI is the semen cryopreservation. The good post-thawed semen quality depends on the composition of the cryopreservation media. The process of semen cryopreservation is a stage of procedure that causes several forms of spermatozoa

damages. One of the most significant components in semen processing is the ingredients of extender that provide protection to the sperm against cold shock <sup>[1]</sup>. The injuries in post-thawed spermatozoa are generally associated with osmotic changes, membrane alteration, inter-and intra-cellular ice crystal formation during cryopreservation process <sup>[2,3]</sup>. Therefore, cryoprotectants are supplemented to the semen extender to reduce damaging effects of cryopreservation processes <sup>[4]</sup>. Considerable evidences suggest that lipidic composition of the sperm membrane is



İletişim (Correspondence)



+62 813 35213112



sidol\_99@yahoo.com

responsible for the detrimental effect of cryopreservation [5]. Before semen freezing, percent docosahexaenoic acid was greater in fatty acid treatment than that in the group without fatty acid and reduced significantly in both groups after thawing [6].

Fish oil contains omega 3 fatty acid and is a major source of eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) [7]. However, it has been reported [8] that fish oil contains 1970 mg saturated fatty acid (SFA), 35.5% polyunsaturated fatty acid (PUFA), 180 mg eicosapentaenoic acid (EPA) and 120 mg docosahexaenoic acid (DHA). Many researchers have assessed the effect of fish oil addition on semen quality parameters in domestic animals. They have reported that fish oil caused increments in sperm motility and reductions in abnormal sperm in boars [9], improved fertility in turkeys [10] as well as increased total of sperm count in rams [11]. On the other hand, the addition of fish oil as a source of omega-3 fatty acids to diet of boars has been demonstrated to possess no significant improvement effect on the quality of frozen-thawed sperm [12]. Similarly, incorporation of DHA as an omega-3 fatty acid in the diet has been found to improve significantly the motility, live sperm and normal morphology of fresh semen [13,14]. Addition of DHA from fish oil to the egg yolk extender was found to be effective for increase the progressive motility and membrane integrity in post-thawed boar semen [15]. Although there are many evidences about the influence of DHA enriched extenders on freezing of boar semen, however, there are limited numbers of reports about the effect of fish oil and/or DHA enriched extender on the quality of frozen-thawed bull semen [16,17]. The objective of this study was to evaluate the effect of fish oil addition to the extender on live sperm rate, motility and abnormality of post-thawed Bali bull spermatozoa.

## MATERIAL and METHODS

### Semen Collection

Semen was collected by artificial vagina from four Bali bulls raised in Center Insemination Banjarbaru, province of south Kalimantan-Indonesia. All the bulls of at least 4-6 years of age and average weight of 625-650 kg were raised under a similar grazing system (various kinds of grass) and supplemented with concentrat at the rate of 4.5 kg/head/day. Two ejaculates were obtained from each bulls every week for one month. Immediately after semen collection, the ejaculates were immersed in the warm water bath at 37°C until their assessments in the laboratory. Then, semen parameters were assessed based on the macroscopic and microscopic characteristics. Macroscopic evaluations included volume, pH and color. Microscopic evaluations included live sperm rate, motility and abnormality. The main extender consisted of 10% skim milk, 5% egg-yolk, 1% glucose, 8% glycerol, Streptomycine (1 mg/mL) and Penicilin (1000 IU/mL).

This study was designed to compare the effect different concentrations of fish oil (*liquid*) with doses 0 (control), 50 mg, 100 mg, 150 mg and 200 mg in the 100 mL skim milk-egg yolk extender. The cryoprotective extender for the control group was the same as that for the treatment groups except that it was not supplemented with fish oil [(natural, 1000 mg) containing Omega-3 Marine; Triglycerides (300 mg) as EPA (180 mg) and DHA (120 mg) from Nature's Care (Manufacture Pty, Ltd. Minna Close Belrose, Australia)].

### Freezing, Thawing and Evaluation

Semen was diluted to obtain a final concentration of  $25 \times 10^6$  sperm/mL. Diluted semen was packed into 0.25 ml straws (Biovet, France). After sealing, straws were placed horizontally on a cold rack (5°C) and lowered into nitrogen vapors (-50°C), 4 cm above the surface of liquid nitrogen. After 3 min, when the temperature reached -120°C, the frozen straws were transferred into goblets of appropriate size and transferred into a liquid nitrogen tank. After two weeks of storage, the straws per treatment were randomly chosen, thawed in water at 37°C for 30 sec and evaluated for post-thawed semen quality including live sperm rate, motility and abnormality.

To evaluate sperm motility, a small droplet ( $\approx 10 \mu\text{L}$ ) of post-thawed semen was placed in the center of a pre-warmed slide and covered with cover slip. It was transferred to a heated microscope stage at 37°C and subjectively calculated by phase contrast microscopy (400 $\times$  magnification). For assessment of live sperm percentage, a modification of the eosin-nigrosin procedure described by Evans and Maxwell [18] was used. 10  $\mu\text{L}$  of post-thawed semen was mixed with 30  $\mu\text{L}$  of eosin-nigrosin stain. A thin smear was prepared and 200 spermatozoa was observed under a microscope (400 x). The same eosin-nigrosin stained slides were also used to define sperm abnormality for which 200 spermatozoa were examined for defects associated with tail region, mid-piece and sperm head.

### Statistical Analysis

The data are expressed as mean  $\pm$  SEM. A one-way analysis of variance (ANOVA) with tukey's multiple comparison test was applied to determine differences among the treatments using SPSS statistical software (version 16.0). Differences with P value 5% were considered to be statistically significant.

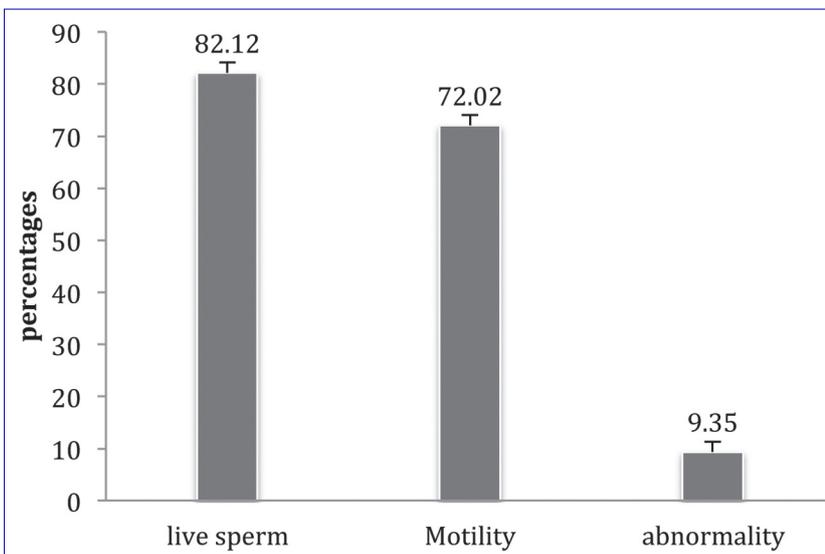
## RESULTS

The effect of addition of different doses of fish oil on post-thawed live sperm rate, motility and abnormality is shown in *Table 1* and *Fig. 1*. Addition of 100 mg fish oil to the 100 mL extender significantly ( $P < 0.05$ ) increased both live sperm rate and motility when compared to the control and other treatments. However, the treatment with 200 mg fish oil caused significant ( $P < 0.05$ ) reduction in live sperm

**Table 1.** Post-thawed sperm parameters including different dose fish oil Bali bulls

Treatments	Sperm parameters		
	Live Sperm Rate (%)	Motility (%)	Abnormality (%)
Control	54.64±2.41 <sup>b</sup>	42.50±2.50 <sup>ab</sup>	18.83±23.06 <sup>a</sup>
50 mg/100 mL	57.32±2.15 <sup>bc</sup>	43.33±1.44 <sup>b</sup>	21.17±50.49 <sup>ab</sup>
100 mg/100 mL	60.36±1.76 <sup>c</sup>	46.67±1.42 <sup>c</sup>	23.66±17.01 <sup>b</sup>
150 mg/100 mL	53.34±2.25 <sup>b</sup>	43.33±1.37 <sup>b</sup>	22.67±48.05 <sup>b</sup>
200 mg/100 mL	47.30±2.02 <sup>a</sup>	40.01±0.21 <sup>a</sup>	22.33±09.21 <sup>b</sup>

<sup>a,b,c</sup> Values in the same column with different superscripts indicate significant difference at  $P < 0.05$ , (n= 24)

**Fig 1.** Characteristic fresh semen of Bali bulls spermatozoa (Mean±SEM) (n= 24)

rate in comparison with the control and other treatments. In addition, 200 mg fish oil supplementation also caused significant ( $P < 0.05$ ) reduction when compared with the other treatment groups but not control group. A significant ( $P < 0.05$ ) increment was observed in abnormality rate of frozen-thawed spermatozoa in comparison with the control group when all doses of fish oil except 50 mg/100 mL were added.

## DISCUSSION

In the present study was evaluated the effects of fish oil supplementation on post-thaw semen quality in Bali bull spermatozoa. We hypothesized that fish oil supplement can enhanced semen quality in terms of live sperm rate and motility but not on sperm abnormality. The results obtained from this study are generally in agreement with the findings of previous studies conducted in other species such as turkeys [10], pigs [8] and rams [10], in which fish oil supplement improved semen quality. Meanwhile, reactive oxygen species (ROS) produced from cryopreservation

can also induce acrosome reaction and spermatozoa damage [19]. The toxicity effects of ROS eventually results in protein ionization and inactivation, peroxidation of lipids, in particular polyunsaturated fatty acids, and DNA damage which destabilize spermatozoa plasma membrane [20,21].

Live sperm rate and motility of semen are important factors because spermatozoa must travel from the vagina and uterus if semen is deposited by natural mating and AI, respectively. Cryopreservation, freezing and thawing processes may induce spermatozoa damages especially to the plasma membrane and organelles [22,23]. The determination of live sperm rate and motility is important for the evaluation of post-thaw quality of semen. In the present study, it was observed that addition of 100 mg fish oil to the 100 mL extender significantly increased both live sperm rate and motility, but treatment with 200 mg fish oil caused significant reduction in live sperm rate in comparison with the control and other treatments. In addition, 200 mg fish oil supplementation also caused significant ( $P < 0.05$ ) reduction when compared with the other treatment groups but not control group. This situation is shown that the dose of 100 mg fish oil provides an ideal concentration in diluent so as to provide comfort to the spermatozoa,

thus preventing damage to the process freezing, cryopreservation and thawing. These results are confirmed by previous reports by Abdi-Benemar et al. [24], who revealed that all frozen-thawed sperm characteristics are significantly enhanced with a surge in DHA levels from fish oil. One of the components of fish oil is PUFA with long chains and has been found in the spermatozoa of various species including ram, man and bull. These fatty acids improve the fluidity of the sperm plasma membrane which is then responsible for increased resistance of the sperm to cold conditions [25]. On the other hand, DHA can be presented as a cryoprotectant that will be able to penetrate the plasma membrane of the sperm and thus act intracellularly. Therefore, modifications in fatty acid composition of surrounding diluents with fish oil supplement can cause changes in the fluidity and elasticity of the sperm plasma membrane by incorporation its fatty acids in the lipids of sperm [26]. In the present study, 150 and 200 mg fish oil supplementations were observed to decrease the percentages of live sperm and motility. This result are

supported by the data previously reported by Kandelousi et al.<sup>[27]</sup>, who stated that higher levels of omega-6 fatty acids result in decreased sperm concentration, motility, and altered morphology.

In the present study, a significant increment was observed in abnormality rate of frozen-thawed spermatozoa in comparison with the control group when all doses of fish oil except 50 mg/100 mL were added. Majority of sperm abnormalities detected in this research were minor or secondary defects like simple coiled or bent tails that usually encounter in response to change in temperature, but not the defects of major type such as acrosome defect, defects associated through the sperm head and mid-piece, which are irreversible and genetically related<sup>[28]</sup>.

In conclusion, the results indicate that addition of 100 mg fish oil to 100 mL of skim milk-egg yolk extender could be beneficial for the improvement of the post-thaw quality of Bali bull spermatozoa including live sperm and motility. Therefore, fish oil addition at the dose of 100 mg per 100 mL skim milk-egg yolk extender can be recommended to bovine semen production centers.

## REFERENCES

1. Demir K, Öztürk Bakırer G, Cirit Ü, Bozkurt HH, Aktaş A, Bırler S, Ak K, Pabuçcuoğlu S: Effects of cooling rate on membrane integrity and motility parameters of cryopreserved ram spermatozoa. *Kafkas Univ Vet Fak Derg*, 21 (1): 61-67, 2015. DOI: 10.9775/kvfd.2014.11726
2. Isachenko E: Vitrification of mammalian spermatozoa in the absence of cryoprotectants: From past practical difficulties to present. *Reprod Biol*, 6, 191-200, 2003. DOI: 10.1016/S1472-6483(10)61710-5
3. Khalili B, Farshad A, Zamiri MJ, Rashidi A, Fazeli P: Effects of sucrose and trehalose on the freezability of Markhoz goat spermatozoa. *Asian Australas J Anim Sci*, 22, 1614-1619, 2009. DOI: 10.5713/ajas.2009.90286
4. Purdy PH: A review on goat sperm cryopreservation. *Small Ruminant Res*, 63, 215-225, 2006. DOI: 10.1016/j.smallrumres.2005.02.015
5. Hammerstedt RH: Maintenance of bioenergetic balance in sperm and prevention of lipid peroxidation: A review of the effect on design of storage preservation systems. *Reprod Fertil Dev*, 5, 675-690, 1993. DOI: 10.1071/RD9930675
6. Mandal R, Damodar B, Jitamanyu C: Role of membrane lipid fatty acids in sperm cryopreservation. *Adv Androl*, 2014 (2014), Article ID: 190542. DOI: 10.1155/2014/190542
7. Stoeckel K, Nielsen LH, Fuhrmann H, Bachmann L: Fatty acid patterns of dog erythrocyte membranes after feeding of a fish-oil based DHA-rich supplement with a base diet low in n-3 fatty acids versus a diet containing added n-3 fatty acids. *Acta Vet Scand*, 6, 53-57, 2011. DOI: 10.1186/1751-0147-53-57
8. Nichols PD, Glencross B, Petrie JR, Singh SP: Readily available sources of long-chain omega-3 oils: Is farmed Australian seafood a better source of the good oil than wild-caught seafood. *Nutrients*, 6, 1063-1079, 2014. DOI: 10.3390/nu6031063
9. Rooke JA, Shao CC, Speake BK: Effects of feeding tuna oil on the lipid composition of pig spermatozoa and *in vitro* characteristics of semen. *Reproduction*, 121 (2): 315-322, 2001.
10. Blesbois E, Douard V, Germain M, Boniface P, Pellet F: Effects of n-3 polyunsaturated dietary supplementation on the reproductive capacity of male turkeys. *Theriogenology*, 61, 537-549, 2004. DOI: 10.1016/S0093-691X(03)00207-3
11. Samadian F, Towhidi A, Rezayazdi K, Bahreini M: Effects of dietary n-3 fatty acids on characteristics and lipid composition of ovine sperm. *Animal*, 4 (12): 2017-2022, 2010. DOI: 10.1017/S1751731110001308
12. Castellano CA, Audet I, Laforest JP, Matte JJ, Suh M: Fish oil diets alter the phospholipid balance, fatty acid composition, and steroid hormone concentrations in testes of adult pigs. *Theriogenology*, 76, 1134-1145, 2011. DOI: 10.1016/j.theriogenology.2011.05.022
13. Aksoy Y, Aksoy H, Altinkaynak K, Aydin HR, Ozkan A: Sperm fatty acid composition in subfertile men. *Prostaglandins Leukot Essent Fatty Acids*, 75, 75-79, 2006. DOI: 10.1016/j.plefa.2006.06.002
14. Am-in N, Kirkwood RN, Techakumphu M, Tantasuparuk W: Lipid profiles of sperm and seminal plasma from boars having normal or low sperm motility. *Theriogenology*, 75, 897-903, 2011. DOI: 10.1016/j.theriogenology.2010.10.032
15. Kaeoket K, Sang-urai P, Thamniyom A, Chanapiwat P, Techakumphu M: Effect of docosahexaenoic acid on quality of cryopreserved boar semen in different breeds. *Reprod Domest Anim*, 45, 458-463, 2010. DOI: 10.1111/j.1439-0531.2008.01239.x
16. Maldjian A, Pizzi F, Gliozzi T, Cerolini S, Penny P, Noble R: Changes in sperm quality and lipid composition during cryopreservation of boar semen. *Theriogenology*, 63, 411-421, 2005. DOI: 10.1016/j.theriogenology.2004.09.021
17. Chanapiwat P, Kaeoket K, Tummaruk P: Effects of DHA-enriched hen egg yolk and L-cysteine supplementation on quality of cryopreserved boar semen. *Asian J Androl*, 11, 600-608, 2009. DOI: 10.1038/aja.2009.40
18. Evans G, Maxwell WMS: Salamon's Artificial Insemination of Sheep and Goats. University Press, Sydney, NSW, Australia, 1987.
19. Mammoto A, Masumoto N, Tahara M, Ikebuchi Y, Ohmichi M, Tasaka K, Miyake A: Reactive oxygen species block sperm-egg fusion via oxidation of sperm sulfhydryl proteins in mice. *Biol Reprod*, 55, 1063-1068, 1996.
20. Aitken RJ: Free radicals, lipid peroxidation and sperm function. *Reprod Fertil Dev*, 7, 659-668, 1995. DOI: 10.1071/RD9950659
21. Aitken RJ, Gordon E, Harkiss D, Twigg JP, Milne P, Jennings Z, Irvine DS: Relative impact of oxidative stress on the functional competence and genomic integrity of humanspermatozoa. *Biol Reprod*, 59, 1037-1046, 1998. DOI: 10.1095/biolreprod59.5.1037
22. Estes MC, Fernández SMR, Soler AJ, Garde JJ: Head dimensions of cryopreserved red deer spermatozoa are affected by the thawing procedure. *Cryoletters*, 24, 261-268, 2003.
23. Munoz OVL, Amirat-Briand T, Diaz L, Va'squez E, Schmidt S, Desherces M, Anton D, Bencharif D, Tainturier D: Effect of semen dilution to low-sperm number per dose on motility and functionality of cryopreserved bovine spermatozoa using low-density lipoproteins (LDL) extender: Comparison to Triladyl® and Bioxcell®. *Theriogenology*, 71, 895-900, 2009. DOI: 10.1016/j.theriogenology.2008.10.010
24. Abdi-Benemar H, M Jafaroghli B, Khalili MJ, Zamiri H, Ezazi AA, Shadparvar: Effects of DHA supplementation of the extender containing egg yolk and - tocopherol on the freezability and post-thawing fertility of ram semen. *Small Ruminant Res*, 130, 166-170, 2015. DOI: 10.1016/j.smallrumres.2015.06.013
25. Wathes DC, Abayasekara DR, Aitken RJ: Polyunsaturated fatty acids in male and female reproduction. *Biol Reprod*, 77, 190-201, 2007. DOI: 10.1095/biolreprod.107.060558
26. Conquer JA, Martin JB, Tummon I, Watson L, Tekpetey F: Effect of DHA supplementation on DHA status and sperm motility in asthenozoospermic males. *Lipids*, 35, 149-154, 2000. DOI: 10.1007/BF02664764
27. Kandelousi SMA, Arshami J, Naserian AA, Abavisani A: The effects of addition of omega-3, 6, 9 fatty acids on the quality of bovine chilled and frozen-thawed sperm. *Open Vet*, 3 (1): 47-52, 2013.
28. Chenoweth PJ: Genetic sperm defects. *Theriogenology*, 64, 457-468, 2005. DOI: 10.1016/j.theriogenology.2005.05.005