

Polymorphism of the Kap 1.1, Kap 1.3 and K33 Genes in Chios, Kivircik and Awassi ^[1]

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Abstract

Keratin-associated proteins are major structural components of hair and wool fibres. They also play an important role in determining the properties of the fibre. This study is designed to examine variation in the genes encoding key keratin and keratin –associated proteins in Chios, Kivircik and Awassi sheep. Variation at these loci has the potential to be developed as genetic markers deal with wool traits. Blood samples were taken into 2 ml sterilized tubes with EDTA from vena jugularis for having genomic DNA samples. Genomic DNA isolation was obtained by standart salting out method. Regions were amplified for the determination of KAP 1.1, KAP 1.3 and K33 polymorphisms in DNA samples by using PCR. Gene polymorphisms and chi-square test were used to determine whether the populations are in Hardy-Weinberg equilibrium using by POPGENE32 software. Results of the sequence analysis of the regions were evaluated by using MEGA 5. The results have shown a possibility to improve the quality of the wool traits on those local species by doing selection trials targeting on those mentioned genes.

Keywords: KAPs, Sequencing, Turkish sheep, Wool trait

Sakız, Kivircik ve İvesi Koyun Irklarında KAP 1.1, KAP 1.3 ve K33 Gen Polimorfizmlerinin Belirlenmesi

Özet

Kreatin ilişkili proteinler, yün ve kıl yapısının komponentleridir. Lifli yapının özelliklerinin belirlenmesinde önemli rol oynarlar. Yapılan bu çalışmada amaç, Sakız, Kivircik ve İvesi koyun ırklarında kreatin ve kreatin ilişkili proteinleri kodlayan genlerdeki varyasyonu incelemektir. Bu lokuslardaki olası varyasyonlar, yün verimi ile ilişkili genetik markör oluşturabilir. Kan örnekleri 2 ml'lik EDTA'lı tüplere vena jugularis den alınmıştır. Standart salting out yöntemiyle genomik DNA izole edilmiştir. KAP 1.1, KAP 1.3 ve K33 için bölgeleri belirlemek için PCR yöntemi kullanılmıştır. Elde edilen sonuçlar POPGEN 32 istatistik programı ile değerlendirilmiştir. Sekans analizi sonuçları için MEGA 5 programı kullanılmıştır. Sonuçlar göstermiştir ki, çalışmadaki yerli ırklarda, söz konusu genlere odaklı seleksiyon uygulamalarının, yün verim ve kalitesini geliştirmek için önem oluşturabilir.

Anahtar sözcükler: KAPs, Sekans, Yerli koyun ırkları, Yün verimi

INTRODUCTION

Most of the proteins in hair and wool are categorized in two types; keratin intermediate filament-forming proteins (known as keratins) and keratin-associated proteins (KAPs). The studies show that there has been an increase in the number of KAP genes defined in humans and other species and progressively accounts of variation in these genes ^[1]. Keratin - associated proteins are a structural constituent of

hair and wool fibres. They play a critical role in determining the physic-mechanical properties of hair and wool fibres ^[2-5].

The wool fibre consists of three main parts, the cuticle, the cortex and medulla ^[6]. The cortex includes 90% of the wool fibre. It also consists of filamentous microfibrils ^[7,8]. Keratin intermediate filaments are formed by microfibrils. KAPs have 3 groups based on amino acids compositions; the high-sulphur proteins (KAP 1.n, KAP 2.n, and KAP 3.n),



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ultra high sulphur proteins (KAP 4.n, KAP 5.n, and KAP 10.n) and the last one; high glycine tyrosine proteins (KAP 6.n, KAP7.n, KAP 8.n) [7,9,10]. KAPs are encoded by a large number of genes which are polymorphic [11]. The KAP genes between two and nine alleles have been determined [12].

Gene expression can be affected by the structure and function of the encoded proteins [13]. It may also be affected by the variation in KAP genes and as a result of that wool traits are influenced. Wool quality might be improved if we can identify the KAP genes which affect wool traits. On the other side only 16 functional genes have been identified until now so this number of sheep KAP genes, may not be sufficient to improve the wool quality. Still many genes have not been determined in human which are homologues in the sheep genome [14-16].

MATERIAL and METHODS

One hundred and thirty five Chios, Awassi and Kivircik sheep were investigated. Information from the breeder producer was considered in order to avoid family connections. Blood from sheep was collected into tubes containing EDTA. Genomic DNA was isolated by using salt-out method [17].

The primers used to amplify the KAP 1.1 locus were designed from a published gene sequence, Gen Bank nos. AY835603- AY835605 to amplify a 311 bp fragment of the KAP1.1. and F :5'-CAACCCTCTCAACCCAACTCC-3'; R :5'-CGCTGCTACCCACCTGGCCAT A-3'.

The KAP 1.3 locus were designed from a published sequence, Gen Bank nos. AY835589-AY835597 to amplify a 598 bp fragment of the KAP 1.3; F :5'-GGGTGGAACAA GCAGACCAAACCTC-3'; R :5'-TAGTTTGTGGGACTGTACACT GGC-3'. The K33 locus were designed from a published sequence, Gen Bank nos. AY835598-AY835602 to amplify a 480 bp fragment of the K33;

F:5'-CACAACCTCTGGCTTGGTGAACCTG-3';

R:5'-CTTAGCCATATCTCGGATCCCTC-3' [18].

Amplification consisted of; initial denaturation 95°C for 1 min, followed by 30 cycles of 95°C for 30 s, annealing at 65°C for 30 s and extension at 72°C for 30 s with a final extension of 72°C for 7 min.

The Polymerase Chain Reaction (PCR) volume of 25 µl contained; 1 U Taq DNA Polymerase (Fermentas Life Sci., Canada), 2-2.5 mL 10X PCR buffer, 1.5 mM MgCl₂, 50-100 ng genomic DNA, 100mM dNTP (TaKaRa Biotechnology Co., Ltd., Japan), and 10 pmol of each primer [18]. PCR products was visualised after electrophoresis on a 2% agarose gel with a long-wavelength UV transilluminator (Thermo Fisher Scientific, Germany).

Sequencing was performed by using an ABI-3100

sequencer (PE Biosystems) and the BigDye™ terminator cycle sequencing kit after the purification of the PCR products. Forward primer was used to sequence the PCR products. Single nucleotide polymorphisms (KAP1.1) in codons 74, 111, 177, 207, 241, 262, 289 were checked directly. For KAP 1.3; codons 53, 60, 66, 67, 160, 178, 184, 232, 241, 264, 313, 337, 352, 364, 380, 486, 557, 598 and for K33; codons 127, 160, 184, 208, 223, 251, 307, 308, 340 were checked directly.

Genotype and allele frequencies of each polymorphism were calculated by using the PopGene 32 software program and also the chi-square tests (χ^2) was used to check whether the populations were in Hardy-Weinberg equilibrium using PopGene32 software [19].

Statistical Analysis

Results of the sequence analysis of the regions were evaluated by using MEGA 5 (Mega Software, USA www.megasoftware.net). MEGA5 is a collection of maximum likelihood (ML) analyses for inferring evolutionary trees, selecting best-fit substitution models (nucleotide or amino acid), inferring ancestral states and sequences (along with probabilities), and estimating evolutionary rates site-by-site [20].

RESULTS

Information on product size, genotype frequencies and allele frequencies for KAP 1.1 gene, KAP 1.3 gene and K33 gene are listed in *Table 1*.

KAP 1.1: Three amplimers of different length were obtained and designated A, B and C. The length of amplimers was 341, 311 and 281 bp respectively. For KAP 1.1 gene, the obtained results indicated that A and B alleles frequencies were higher in Awassi than the others. It was found that C allele frequency was higher in Chios than the Awassi and Kivircik.

KAP 1.3: Nine amplimers of different length were obtained and designated A-I. The results noticed that the C allele was the highest in Awassi, the G allele was the highest in Kivircik. Both of the A and D alleles were not determined in all three breed.

K33: Five unique SSCP banding were observed and designated A-E. The D allele was not obtained in all three breeds. The results indicated that the A allele was the highest in Chios and the B allele, the C allele and the E allele were the highest in Kivircik.

DISCUSSION

This study reports three alleles at the KAP 1.1 locus. The length of the amplimers from KAP 1.1 alleles A, B and C were 341, 311, 281 bp respectively. The length

Table 1. Loci, product size, genotypes and allele frequencies for polymorphisms in three candidate genes**Table 1.** Üç aday genin, polimorfizmde lokus, ürün, genotip ve allel frekans verileri

Locus	Breed	n	Product Size	Genotype Frequency (%)								Allele Frequency (%)										(χ ²)				
				AA	AB	BB	CC							A	B	C										
KAP 1.1			311	AA	AB	BB	CC							A	B	C										
	Sakız	45		7	8	20	10							0.24	0.53	0.23										56.67***
	Kivircik	45		1	9	27	8							0.12	0.7	0.18										47.70***
	İvesi	45		10	10	24	1							0.33	0.64	0.03										100.58***
KAP 1.3			598	AA	BB	CC	DD	EE	FF	GG	HH	II	A	B	C	D	E	F	G	H	I					
	Sakız	45			8	15		3	2	6	4	7		0.18	0.34		0.06	0.05	0.13	0.09	0.15					308.00***
	Kivircik	45				11		3		25	3	3			0.24		0.07		0.55	0.07	0.07					207.00***
	İvesi	45				34				6	4	1			0.75				0.14	0.08	0.03					188.57***
K33			480	AA	BB	CC	DD	EE					A	B	C	D	E									
	Sakız	45		41		3		1					0.92	0.01	0.07											53.43***
	Kivircik	45		35		5		5					0.81	0.04	0.11		0.04									108.92***
	İvesi	45		38		3		4					0.87	0.03	0.07		0.03									112.00***

¹ Hardy-Weinberg equilibrium, *** P≤0.001

polymorphism of the KAP 1.1 gene determined in this investigation has previously been reported in Romney sheep and in Merino sheep [18,21]. Our results showed that A and B alleles frequencies were higher in Awassi than the others and also the C allele frequency was the highest in Chios breed.

Five alleles were identified at the K33 locus in Merino sheep which was reported by Itenge *et al.* [18]. By the way Roger *et al.* [21] determined a diallelic polymorphism at this same loci. In our study, the D allele wasn't obtained in all three breed. Addition to this, the E allele was not obtained for Chios sheep.

Itenge *et al.* [18] reported eight alleles (except for B allele) in a 598 bp KAP 1.3 amplimer from Merino sheep. The results found by Powell *et al.* [22] also supported these findings. Roger *et al.* [21] determined six alleles from Romney sheep. In this study we obtained seven alleles for Chios sheep, five alleles for Kivircik sheep and four alleles for Awassi sheep in a 598 bp KAP 1.3 amplimer. Both of the A and D alleles were not determined in all of them. Our investigation showed that the C allele was the highest in Awassi, 0.75 by the way 0.34 for Chios and 0.24 for Kivircik, the G allele was the highest in Kivircik whereas the G allele was 0.13 in Chios and 0.14 in Awassi.

As a conclusion it has been known that Kivircik sheep breed has been preferred for wool quality and Awassi sheep breed has been preferred for wool trait among the domestic sheep breed in Turkey. Their wool quality ranges from carpet to medium-wool quality [23]. It might be a relationship between these alleles and wool quality and wool product. To support this suggestion, further linkage

analyses are necessary. This will allow future analysis of how these genes may affect wool traits.

The allele numbers at the KAP1.3 and the K33 locus in Chios, Awassi and Kivircik were determined lower than the Merino sheep and Romney sheep breed. It might be suggested to make further trials on those loci to possible improvement on wool quality and wool trait in those breeds.

REFERENCES

- Hua G, Huitong Z, Grant W, Zhidong Y, Stefan C, Jolon MD, Jeffrey EP, Mathew WW, Reena A, Simon B, Yulin C, Jinquan L, Jonathan GHH:** An updated nomenclature for keratin-associated protein (KAPs). *Int J Biol Sci*, 8, 258-264, 2012. DOI: 10.7150/ijbs.3278
- Powell BC, Rogers GE:** The role of keratin proteins and their genes in the growth, structure and properties of hair. *In*, Jolles P, Zahn H, Hocker E (Eds): Formation and Structure of Human Hair. 59-148, Birkhauser Verlag, Basel, 1995.
- Powell BC:** The keratin proteins and genes of wool and hair. *Wool Tech Sheep Breed*, 44, 100-118, 1996.
- Rogers MA, Winter H, Langbein L, Wollschlager A, Praetzel-Wunder S, Jave-Suarez S, Schweizer J:** Characterization of human KAP24.1, a cuticular hair keratin associated protein with unusual amino-acid composition and repeat structure. *J Invest Dermatol*, 127, 1197-1204 2007.
- Hua G, Huitong Z, Hickford JGH:** Diversity of the glycine/tyrosine-rich keratin associate protein 6 gene (KAP 6) family in sheep. *Mol Biol Rep*, 38, 31-35, 2011. DOI: 10.1007/s11033-010-0074-6
- Onions WJ:** Wool - an introduction to its properties, varieties, uses and production. 278, Ernest Benn Ltd, London, United Kingdom, 1962.
- Marshall, RC, Orwin DFG, Gillespie JM:** Structure and biochemistry of mammalian hard keratin. *Electron Microsc Rev*, 4, 47-83, 1991. DOI: 10.1016/0892-0354(91)90016-6
- Powell BC, Rogers GE:** Hair keratin: Composition, structure and biogenesis. *In*, Bereiter-Hahn J, Matoltsy AG, Richards KS (Eds): Biology of

the Integument. 695-721, Heidelberg Springer-Verlag, Berlin, 1986.

9. Parry D, Steinert PM: Intermediate filament structure. *Cur Opin Cell Biol*, 4, 94-98, 1992. DOI: 10.1016/0955-0674(92)90064-J

10. Plowman JE: Proteomic database of wool components. *J Chrom B*, 787, 63-76, 1996. DOI: 10.1016/S1570-0232(02)00211-8

11. Rogers MA, Schweizer J: Human KAP genes, only the half of it? Extensive size polymorphisms in hair keratin-associated protein genes. *J Invest Dermatol*, 124, 7-9, 2005. DOI: 10.1111/j.0022-202X.2005.23728.x

12. Gong H, Zhou H, McKenzie GW, Hickford JGH, Yu Z, Clerens S, Dyer JM, Plowman JE: Emerging issues with the current keratin-associated protein nomenclature. *Int J Tricholog*, 2, 104-105, 2010. DOI: 10.4103/0974-7753.77519

13. Elmaci C, Sahin S, Oner Y: Distribution of different alleles of aromatase cytochrome P450 (CYP19) and melatonin receptor 1A (MTRN1A) genes among native Turkish sheep breeds. *Kafkas Univ Vet Fak Derg*, 19, 929-933, 2013. DOI: 10.9775/kvfd.2013.8900

14. Cottle DJ, Bryson WG, Aitken GD: A review of markers for wool and sheep carcass quality traits. *Wool Tech Sheep Breed*, 50, 401-409, 2002.

15. Schweizer J, Bowden PE, Coulombe PA, Langbein L, Lane EB, Magin TM, Maltais L, Omary MB, Parry DA, Rogers MA, Wright MW: New consensus nomenclature for mammalian keratins. *J Cell Biol*, 174, 169-174, 2006. DOI: 10.1083/jcb.200603161

16. Huitong Z, Hua G, Wei Y, Yuzhu L, Hickford JGH: Identification and sequence analysis of the keratin-associated protein 24-1(KAP24-

1) gene homologue in sheep. *Gene*, 511, 62-65, 2012. DOI: 10.1016/j.gene.2012.08.049

17. Miller SA, Dykes DD, Polesky HE: A simple salting out procedure for extracting DNA from human nucleated cells. *Nucleic Acids Res*, 16 (3): 1215, 1998.

18. Itenge-Mweza TO, Forresta RHJ, McKenzie GW, Hogan A, Abbott J, Amofo O, Hickford JGH: Polymorphism of the KAP1.1, KAP1.3 and K33 genes in Merino sheep. *Mol Cell Prob*, 21, 338-342, 2007. DOI: 10.1016/j.mcp.2007.04.002

19. Yeh FC, Yang RC, Boyle T: Popgene version 1.31, Microsoft Windows®-Based Freeware for Population Genetic Analysis. <http://www.ualberta.ca/~fyeh/pop32.exe>. Last modified August 1999. Accessed: April 12, 2013.

20. Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S: MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance and maximum parsimony methods. *Mol Biol Evol*, 28, 2731-2739, 2011. DOI: 10.1093/molbev/msr121

21. Roger GR, Hickford JGH, Bickerstaffe R: Polymorphism in two genes for B2 high sulphur proteins of wool. *Anim Genet*, 25, 407-415, 1994. DOI: 10.1111/j.1365-2052.1994.tb00531.x

22. Powell BC, Sleight MJ, Ward KA, Rogers GE: Mammalian keratin gene families: Organization of genes coding for the B2 high-sulphur proteins of sheep wool. *Nucleic Acids Res*, 16, 5327-5346, 1983.

23. Mason IL: A world dictionary of livestock breeds, types and varieties. Fourth ed., CAB International, Wallingford, United Kingdom, 273, 1996.