SNPs Detected in the SIRT1 and H-FABP Genes and Their Association with Growth Traits in Yak

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Abstract

The aims of this study were to investigate whether the Sirtuin 1 (*SIRT1*) and Hear fatty acid binding protein (*H-FABP*) genes could be used as candidate genes in the breeding of yak. In 409 indigenous Chinese yaks, two single nucleotide polymorphisms (SNPs) were identified through DNA sequencing technology, including a SNP within the 5'UTR of *SIRT1* (g.1906A>G), and a SNP in the exon 1 of *H-FABP* (g.6643C>T). The chi-square test suggested that all the variations were in Hardy-Weinberg equilibrium (P>0.05). An association analysis suggested that g.1906A>G of *SIRT1* gene and g.6643C>T of *H-FABP* gene exhibited significant correlation with body weight and body length (P<0.01). These results indicated that these SNPs could be used as meritorious and available genetic markers in yak growth traits breeding.

Keywords: SIRT1 gene, H-FABP gene, Growth traits, Yak, Single nucleotide polymorphism

Yak Sığırında SIRT1 ve H-FABP Genlerinde Belirlenen Tek Nükleotid Polimorfizmleri ve Büyüme Özellikleri İle İlişkisi

Öz

Bu çalışmanın amacı Yak sığırı yetiştiriciliğinde Sirtuin 1 (*SIRT1*) ve Kalp tipi yağ asidi bağlayıcı protein (*H-FABP*) genlerinin kullanılabilirliğinin araştırılmasıdır. Yerel 409 Çin yak sığırında *SIRT1* geninin (g.1906A>G) 5'UTR'sinde ve *H-FABP* geninin (g.6643C>T) ekzon 1'inde olmak üzere iki adet tek nükleotid polimorfizmi (SNP) DNA sekanslama teknolojisi ile tespit edildi. Ki kare testi tüm varyasyonların Hardy-Weinberg denklemi içerisinde olduğunu gösterdi (P>0.05). İlişki analizi, *SIRT1* geninin g.1906A>G ve *H-FABP* geninin g.6643C>T'sinin vücut ağırlığı ve uzunluğu ile anlamlı derecede ilişkili olduğunu gösterdi (P<0.01). Bu sonuçlar, SNP'lerin Yak sığırı büyüme özelliklerine bağlı yetiştiricilikte kullanılabilecek önemli genetik belirteçler olabileceğini göstermiştir.

Anahtar sözcükler: SIRT1 geni, H-FABP geni, Büyüme özellikleri, Yak, Tek nükleotid polimorfizmi

INTRODUCTION

Yak (Bos grunniens) was distributed mainly in the Qinghai-Tibetan Plateau at altitudes from 3000 m to 5000 m above sea level, which could survive in conditions of extreme harshness with extreme cold, poor oxygen concentrations, and low air pressure ^[1,2]. At present, the total population of yak in China were estimated to be 15 million and accounted for over 90% of those distributed all over the world ^[3]. This species provided hides, meat, and milk for local Tibetans needs and played a crucial

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role in contribution to the animal husbandry economy in Qinghai-Tibetan Plateau^[4].

Mammalian Sirtuin 1 (*SIRT1*) was localized in the nucleus, wherein it influenced the activity of transcription factors via deacetylate histones ^[5]. In vivo and vitro, *SIRT1* regulated body growth through modulated insulin resistance and body glucose equilibrium ^[6,7]. In response to fasting, the secretion of insulin in *SIRT1^{-/-}* mice was significant restrained ^[8], when compared with wild-type littermates in pancreatic β -cells. In turn, β -cells-specific *SIRT1*-overexpression transgenic

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mice influenced adenosine triphosphate (ATP) production by repressing uncoupling protein 2 (*UCP2*), consequently contributing to enhanced glucose-stimulated insulin secretion ^[9]. *SIRT1* deacetylated peroxisome proliferatoractivated receptor gamma coactivator 1-alpha (*PGC-1a*) at multiple lysine sites, resulting in alteration of genetic programs for gluconeogenesis and glycolysis in the liver ^[10]. In addition, *SIRT2* deacetylated forkhead box O 1 (*FoxO1*), in parallel with influences the activation of NeuroD and MafA, thereby inhibiting the secretion of insulin in β -cells ^[11].

Hear type fatty acid binding protein (*H-FABP*) was expressed predominantly in hear and skeletal muscle ^[12], wherein played a crucial role in signal transduction pathways such as the uptake or utilization of long chain fatty acids ^[13]. The *H-FABP*-nullt mice exhibited improved insulin sensitivity, which was perhaps related to the increased reliance on glucose ^[14]. The expressions of *UCP2*, *UCP3*, *ACOX1*, and *PGC-1a* involving in lipid and glycolysis metabolism were modified by the knockdown of *H-FABP* gene in brown adipocytes ^[15]. As a transcriptional factor, *KLF15* gene regulated diverse arrays of biological processes including cell proliferation, differentiation and apoptosis ^[16]. Previous studies demonstrated that the *KLF15* gene might modify the core promoter of *H-FABP* gene, thereby influencing the growth-related traits in mammal ^[17].

Based on the functional role in metabolism, polymorphism of *SIRT1* and *H-FABP* genes had been previously demonstrated in pig ^[18], cattle ^[19], and human ^[20]. However, there were no reports on associations between the variations of these two genes and growth traits in yak. The main purpose of this study was to evaluate the genetic association between polymorphism of the *SIRT1* and *H-FABP* genes and growth traits in Chinses domestic yak.

MATERIAL and METHODS

Experimental Animals

In total, 409 female yak aged from 12 to 24 months were randomly collected, which belonged to five farm reared

in Qilian county, Qinghai Province of China. They were reared under the same environmental dry-lot nutrition standard conditions. Blood samples were collected from the jugular vein, and stored at -20°C. Genomic DNA was extracted using a DNA extraction Kit (OMGAM Bio-Tek, Doraville, USA) following instructions provided in the attached protocol. The DNA concentration was estimated spectrophotometrically, and then the DNA was diluted to 50 ng/uL. Meanwhile, the phenotypes traits including body weight, body length, withers height and chest circumference were measured according to Gilbert's method ^[21]. For the accuracy of the results, all individuals were measured once by the same person.

SNP Screening and Genotyping

Available sequence information from yak SIRT1 gene (Genbank accession no NW_005395486.1) and H-FABP (Genbank accession no NW_005395183.1) were used to design PCR primers (Table 1). 5 primer pairs were synthesized by Sangon Biotech (Shanghai, Chian) Co., Ltd. The PCR was carried out in 30 uL mixture containing 50 ng DNA, 10 pM of each primer, 0.2 mM dNTP, 2.5 mM MgCl₂, and 0.5 U Taq DNA polymerase. The cycling protocol was as follows: initial denaturation at 94°C for 5 min, with 35 cycles of denaturation at 94°C for 30 s, annealing for 30 s at optimum temperature, and elongation at 72°C for 30 s. The final extension was performed at 72°C for 10 min. All PCR products were sequenced in forward direction by using the ABI 3730 sequencer (Applied Biosystems, Foster City, CA, USA), and the results were analyzed by DNAMAN software version 5.2.2 (Madison, WI, USA). In this study, all 409 samples were genotyped by directly sequencing method of PCR products respectively.

Statistical Analysis

Genotyping and allele frequencies, Hardy-Weinberg equilibrium (HWE) and polymorphism information content (PIC) were calculated by online website (*http://www.msrcall.com/Gdicall.aspx*). The effect of genotypes of SNPs on the

Table 1. Primers used for PCR amplification					
Primer Name	Primer Sequence (5' to 3')	Annealing Temperature	Product Length	Amplified Region	
SIRT1-L1	CCTGATTTCATTGGGATA		777 h.c	5'UTR	
SIRTI-LI	AAGGCTGAGCAAATAACC	02.5 C	777 bp	SUIK	
SIRT1-L2	CTTGGACTTGGCATTCTC	C0.0°C	251 ha	Fever 4	
SIRTT-L2	TGGGCTCTTTACCACTCT	60.0℃	351 bp	Eexon 4	
	TTTTGGCTTACAGGAACT	50.2°C	740 hrs	3′UTR	
SIRT1-L3	AGGCGTTTACTAATCTGC	58.3℃	749 bp		
H-FABP-L1	CTATGTAACGTCTTTGAAGG	61.0°C	500 hr	Eexon 1	
	ACAGGCAACAGGTAGATGCT	01.0 C	509 bp		
	GGCTGGCTGAGCTCTGGCTC	60.5%	549 hr	Eexon 2	
H-FABP-L2	AGTGAGGCTTTGTGCTCTGC	60.5℃	548 bp		

growth traits was analyzed by SPSS software (Version 18.0). The model applied was: $Y_{ijk} = \mu + G_i + A_j + F_k + E_{ijk}$, where Y_{ijk} is the trait value for each individual, μ is the overall population mean, G_i is the fixed effect associated with the *i*th genotype, A_j is the fixed effect of the *i*th age, F_k is the fixed effect of the *i*th age, F_k is the fixed effect of the *i*th farm and E_i is the random error. In this model, P-values less than 0.05 were considered to be statistically significant.

RESULTS

SNP Detection and Genetic Diversity Analyses

As is presented in *Fig.* 1, two variations were identified through DNA sequencing analysis, including one SNP within 5'UTR of *SIRT1* gene (g.1906A>G), and one SNP located in exon 1 of *H-FABP* gene (g.6643C>T). The g.1906A>G locus had 3 genotypes (AA, AG and GG, respectively). The g.6643C>T locus had 2 genotypes, the TT genotype was not observed in the sampled animals.

Table 2 illustrates the genotypic and allelic frequencies at locus of *SIRT1* and *H-FABP* genes. The "A" allele of g.1906A>G (*SIRT1*) and the "C" allele of g.6643C>T (H-FABP) were found to be predominant (72.98% and 90.10%, respectively). In addition, the g.1906A>G had moderate polymorphic status (0.25 <PIC<0.05).

Association Analysis

In the present study, one SNP (g.1906A>G) in the 5'UTR of *SIRT1* gene was identified. Statistical results showed that the animals with genotype AA exhibited significantly higher body weight and body length compared to genotype GG (P=0.000, P=0.004, respectively).

For the *H-FABP* gene, statistical analyses indicated animals with genotype CC of g.6643C>T had significantly higher body weight and body length than those with genotype CT (P=0.000 and P=0.031, respectively), demonstrating that allele "C" might be associated with an increase in growth traits in yak (*Table 3*).

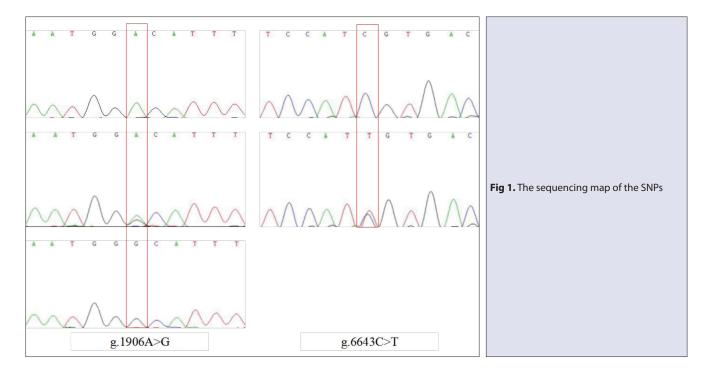


Table 2. Distribution of genotype and allele frequencies in yak						
Site	Genotype (N)	Genotypic Frequency (%)	Alleles	Allele Frequency (%)	χ²(HWE)	PIC
	AA (225)	55.01	G	72.98	P>0.05	0.32
g.1906A>G (<i>SIR</i> T1)	AG (147)	35.94	Т	27.02		
	GG (37)	9.05				
	CC (328)	80.20 C 90.10	P>0.05	0.16		
g.6643C>T (<i>H-FABP</i>)	CT (81)	19.80	Т	9.90		
	TT (0)	0.00				
χ² (HWE): Hardy-Weinbe	erg equilibrium χ² va	lue, Hard-Weinberg equilibriu	m (P>0.05), Harc	dy-Weinberg disequilibriu	um (P<0.05)	

6 14 -	Traits	Genotype (Mean±SE)			5141
Site		AA	AG	GG	P Value
	BW (kg)	175.09±0.87 ^A	165.58±1.10 ^B	156.90±2.15 ^c	0.000
	BH (cm)	106.98±0.63	104.64±0.78	105.84±1.55	0.313
g.1906A>G (<i>SIRT1</i>)	BL (cm)	120.89±0.37 ^A	116.89±0.46	113.27±0.91 ^B	0.004
	CC (cm)	144.91±0.39	142.48±0.49	140.16±0.94	0.147
		СС	СТ		
	BW (kg)	172.41±0.75 ^A	160.38±1.24 ^B		0.000
g.6643C>T (<i>H-FABP</i>)	BH (cm)	107.62±0.28	106.30±0.56		0.464
	BL (cm)	119.99±0.30ª	113.79±0.63 ^b		0.031
	CC (cm)	143.87±0.33	142.52±0.67		0.232

DISCUSSION

Through deacetylating specific transcription factors (i.e., PPARy and FOXO1) [22], the SIRT1 gene modulated mitochondrial capacity ^[23], insulin secretion ^[9] and plasma glucose levels ^[24]. Previously, a polymorphism (g.-274A>G) within the promoter region of SIRT1 gene in a Nanyang cattle breed was significantly correlated with body height and body weight ^[25]. Three SNPs (g.25764G>A, g.25846A>G and g.25868T>C) of SIRT1 gene were identified in Qinchuan cattle, and were observed to be associated with body length and withers height [26]. Additionally, the novel 12bp indel of SIRT1 gene were associated with body weight, chest circumference and height at hip cross Chinese beef cattle^[27]. In the present study, the statistical results showed that the individuals with genotype AA of g.4845C>T of SIRT1 gene were significantly associated with body weight and body length than the other genotype. Those results were consistent with the previous findings of other species [18-20].

The g.1906A>G was located in region of 5'UTR of *SIRT1* gene, without changing the structure of amino acid. Recently, there were some reports about the effects of variants located in region of 5'UTR on the gene expression pattern ^[28]. A SNP within 5'UTR region of zinc finger, BED-type containing 6 (*ZBED6*) gene resulted in transcription factor change, and then altered the transcription activity and mRNA expression in beef cattle ^[29]. A 5'UTR-region SNP of growth hormone-releasing hormone receptor (*GHRHR*) gene locus had been found to be associated with body weight and average daily gain in Chinese cattle ^[30]. Thus, it was an interesting work to find out the mechanism for the association between these 5'UTR-region SNPs and the growth traits in yak.

The *H-FABP* gene contained four exons, and was expressed predominantly in skeletal muscle and subcutaneous fat ^[12]. Because of its important role in metabolism, association between *H-FABP* polymorphisms and economic characters

in livestock was extensively studied. In Berkshire pig breed, one SNP (*H-FABP*/*Hinf*1) was found to be associated with the live weight and fatty acid composition ^[31]. Similarly, this SNP had a significant effect on moisture, tenderness and flavor score of Korean native pig ^[32]. For the g.6643C>T of *H-FABP* locus, only 2 genotypes were detected. The result was due to the lack of genotype in the population or the small size of experimental population. The genotypic frequencies of g.6643C>T of *H-FABP* gene conformed to Hardy-Weinberg equilibrium might be a result of random mating in yak ^[33]. Additionally, our experimental populations belonged to intermediate genetic diversity. This reflected that this genetic marker could provide more reasonable and effective genetic information ^[34].

Sequence alignment showed that the g.6643C>T of *H-FABP* gene was synonymous (Thr \rightarrow Thr), and thus did not change the structure of the encoded protein. It had been demonstrated previously that synonymous SNPs could modify the stability of mRNA ^[35,36], thereby influencing the phenotypic traits in mammal. A synonymous mutation (g.25557A>G) in the silent information regulator 6 (*SIRT6*) gene was found to be associated with intramuscular fat in Qinchuan cattle ^[37]. Additionally, synonymous mutation (c.72G>A) of the *UCP2* gene was related to growth performance, carcass characteristics and meat quality in rabbits ^[38].

Results of the present study suggested that g.1906A>G of *SIRT1* and g.6643C>T of *H-FABP* were significantly correlated with growth traits in yak. It inferred that these SNPs could modify stability/expression of mRNA, therefore influencing the growth-related phenotypes in yak.

In summary, genotypes of *SIRT1* and *H-FABP* were confirmed to be significantly associated with body weight and body length in yak. Our investigation provided evidence that both *SIRT1* and *H-FABP* genes could be used as molecular markers for better growth traits of yak.

CONFLICTS OF INTEREST

The authors declare no conflict of interest.

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REFERENCES

1. Shi Y, Hu Y, Wang J, Elzo MA, Yang X, Lai S: Genetic diversities of MT-ND1 and MT-ND2 genes are associated with high-altitude adaptation in yak. *Mitochondrial DNA A*, 29, 485-494, 2018. DOI: 10.1080/24701394.2017.1307976

2. Xia W, Osorio JS, Yang Y, Liu D, Jiang MF: Characterization of gene expression profiles related to yak milk protein synthesis during the lactation cycle. *J Dairy Sci*, 101, 11150-11158, 2018. DOI: 10.3168/jds.2018-14715

3. Cai X, Mipam TD, Zhang H, Yue B: Abundant variations of *MC4R* gene revealed by Phylogenies of Yak (*Bos grunniens*) and other mammals. *Mol Biol Rep*, 38, 2733-2738, 2011. DOI: 10.1007/s11033-010-0418-2

4. Wang K, Hu Q, Ma H, Wang L, Yang Y, Luo W, Qiu Q: Genome-wide variation within and between wild and domestic yak. *Mol Ecol Resour*, 14, 794-801, 2014. DOI: 10.1111/1755-0998.12226

5. Guarente L: Calorie restriction and sirtuins revisited. *Genes Dev*, 27, 2072-2085, 2013. DOI: 10.1101/gad.227439.113

6. Iyer S, Han L, Bartell SM, Kim HN, Gubrij I, de Cabo R, O'Brien CA, Manolagas SC, Almeida M: Sirtuin1 (Sirt1) promotes cortical bone formation by preventing beta-catenin sequestration by FoxO transcription factors in osteoblast progenitors. *J Biol Chem*, 289, 24069-24078, 2014. DOI: 10.1074/jbc.M114.561803

7. Schenk S, McCurdy CE, Philp A, Chen MZ, Holliday MJ, Bandyopadhyay GK, Osborn O, Baar K, Olefsky JM: Sirt1 enhances skeletal muscle insulin sensitivity in mice during caloric restriction. *J Clin Invest*, 121, 4281-4288, 2011. DOI: 10.1172/JCI58554

8. Yoshizaki T, Schenk S, Imamura T, Babendure JL, Sonoda N, Bae EJ, Oh DY, Lu M, Milne JC, Westphal C, Bandyopadhyay G, Olefsky JM: SIRT1 inhibits inflammatory pathways in macrophages and modulates insulin sensitivity. *Am J Physiol Endocrinol Metab*, 298, E419-E428, 2010. DOI: 10.1152/ajpendo.00417.2009

9. Bordone L, Motta MC, Picard F, Robinson A, Jhala US, Apfeld J, McDonagh T, Lemieux M, McBurney M, Szilvasi A, Easlon EJ, Lin SJ, Guarente L: Sirt1 regulates insulin secretion by repressing UCP2 in pancreatic β cells. *PLoS Biol*, 4 (2): e31, 2006. DOI: 10.1371/journal. pbio.0040031

10. Rodgers JT, Lerin C, Haas W, Gygi SP, Spiegelman BM, Puigserver P: Nutrient control of glucose homeostasis through a complex of PGC-1alpha and SIRT1. *Nature*, 434, 113-118, 2005. DOI: 10.1038/nature03354

11. Kitamura YI, Kitamura T, Kruse JP, Raum JC, Stein R, Gu W, Accili D: FoxO1 protects against pancreatic β cell failure through NeuroD and MafA induction. *Cell Metab*, 2, 153-163, 2005. DOI: 10.1016/j.cmet.2005.08.004

12. Smathers RL, Petersen DR: The human fatty acid-binding protein family: Evolutionary divergences and functions. *Hum Genomics*, 5, 170-191. 2011. DOI. 10.1186/1479-7364-5-3-170

13. Chen QM, Wang H, Zeng YQ, Chen W: Developmental changes and effect on intramuscular fat content of H-FABP and A-FABP mRNA expression in pigs. *J Appl Genet*, 54, 119-123, 2013. DOI: 10.1007/s13353-012-0122-0

14. Shearer J, Fueger, PT, Bracy, DP, Wasserman, DH, Rottman, JN: Partial gene deletion of heart-type fatty acid-binding protein limits the severity of dietary-induced insulin resistance. *Diabetes*, 54, 3133-3139,

2005. DOI: 10.2337/diabetes.54.11.3133

15. Vergnes L, Chin R, Young SG, Reue K: Heart-type fatty acid-binding protein is essential for efficient brown adipose tissue fatty acid oxidation and cold tolerance. *J Biol Chem*, 286, 380-390, 2011. DOI: 10.1074/jbc. M110.184754

16. Prosdocimo DA, Sabeh MK, Jain MK: Kruppel-like factors in muscle health and disease. *Trends Cardiovasc Med*, 25, 278-287, 2015. DOI: 10.1016/j.tcm.2014.11.006

17. Li A, Wu L, Wang X, Xin Y, Zan L: Tissue expression analysis, cloning and characterization of the 5'-regulatory region of the bovine *FABP3* gene. *Mol Biol Rep*, 43, 991-998, 2016. DOI: 10.1007/s11033-016-4026-7

18. Cho KH, Kim MJ, Jeon GJ, Chung HY: Association of genetic variants for *FABP3* gene with back fat thickness and intramuscular fat content in pig. *Mol Biol Rep*, 38, 2161-2166, 2011. DOI: 10.1007/s11033-010-0344-3

19. Gui L, Hao R, Zhang Y, Zhao X, Zan L: Haplotype distribution in the class I sirtuin genes and their associations with ultrasound carcass traits in Qinchuan cattle (*Bos taurus*). *Mol Cell Probes*, 29, 167-171, 2015. DOI: 10.1016/j.mcp.2015.03.007

20. Shimoyama Y, Suzuki K, Hamajima N, Niwa T: Sirtuin 1 gene polymorphisms are associated with body fat and blood pressure in Japanese. *Transl Res*, 157, 339-347, 2011. DOI: 10.1016/j.trsl.2011.02.004

21. Gilbert RP, Bailey DR, Shannon NH: Linear body measurements of cattle before and after 20 years of selection for postweaning gain when fed two different diets. *J Anim Sci*, 71, 1712-1720, 1993.

22. Qiang L, Wang L, Kon N, Zhao W, Lee S, Zhang Y, Rosenbaum M, Zhao Y, Gu W, Farmer SR, Accili D: Brown remodeling of white adipose tissue by SirT1-dependent deacetylation of Ppargγ. *Cell*, 150, 620-632, 2012. DOI: 10.1016/j.cell.2012.06.027

23. Jung HY, Lee D, Ryu HG, Choi BH, Go Y, Lee N, Lee D, Son HG, Jeon J, Kim SH, Yoon JH, Park SM, Lee SV, Lee IK, Choi KY, Ryu SH, Nohara K, Yoo SH, Chen Z, Kim KT: Myricetin improves endurance capacity and mitochondrial density by activating SIRT1 and PGC-1a. *Sci Rep*, 7:6237, 2017. DOI: 10.1038/s41598-017-05303-2

24. Gilbert RE, Thai K, Advani SL, Cummins CL, Kepecs DM, Schroer SA, Woo M, Zhang Y: SIRT1 activation ameliorates hyperglycaemia by inducing a torpor-like state in an obese mouse model of type 2 diabetes. *Diabetologia*, 58, 819-827, 2015. DOI: 10.1007/s00125-014-3485-4

25. Li M, Sun X, Hua L, Lai X, Lan X, Lei C, Zhang C, Qi X, Chen H: SIRT1 gene polymorphisms are associated with growth traits in Nanyang cattle. *Mol Cell Probes*, 27, 215-220, 2013. DOI: 10.1016/j.mcp.2013.07.002

26. Gui L, Wang H, Wei S, Zhang Y, Zan L: Molecular characterization, expression profiles, and analysis of *Qinchuan* cattle *SIRT1* gene association with meat quality and body measurement traits (*Bos taurus*). *Mol Biol Rep*, 41, 5237-5246, 2014. DOI: 10.1007/s11033-014-3393-1

27. Jin Y, Yang Q, Gao J, Tang Q, Duan B, Yu T, Qi X, Liu J, Wang R, Dang R, Lei C, Chen H, Lan X: Detection of insertions/deletions within SIRT1, SIRT2 and SIRT3 genes and their associations with body measurement traits in cattle. *Biochem Genet*, 56, 663-676, 2018. DOI: 10.1007/s10528-018-9868-3

28. Pagani F, Baralle FE: Genomic variants in exons and introns: Identifying the splicing spoilers. *Nat Rev Genet*, 5, 389-396, 2004. DOI: 10.1038/nrg1327

29. Huang YZ, Li MX, Wang J, Zhan ZY, Sun YJ, Sun JJ, Li CJ, Lan XY, Lei CZ, Zhang CL, Chen H: A 5'-regulatory region and two coding region polymorphisms modulate promoter activity and gene expression of the growth suppressor gene *ZBED6* in cattle. *PLoS One*, 8 (11): e79744, 2013. DOI: 10.1371/journal.pone.0079744

30. Zhang CF, Chen H, Zhang ZY, Zhang LZ, Yang DY, Qu YJ, Hua LS, Zhang B, Hu SR: A 5'UTR SNP of GHRHR locus is associated with body weight and average daily gain in Chinese cattle. *Mol Biol Rep*, 39, 10469-10473, 2012. DOI: 10.1007/s11033-012-1927-y

31. Lee SH, Choi YM, Choe JH, Kim JM, Hong KC, Park HC, Kim BC: Association between polymorphisms of the heart fatty acid binding protein gene and intramuscular fat content, fatty acid composition, and meat quality in Berkshire breed. *Meat Sci*, 86, 794-800, 2010. DOI: 10.1016/j.meatsci.2010.06.024 **32. Li X, Kim SW, Choi JS, Lee YM, Lee CK, Choi BH, Kim TH, Choi YI, Kim JJ, Kim KS:** Investigation of porcine *FABP3* and *LEPR* gene polymorphisms and mRNA expression for variation in intramuscular fat content. *Mol Biol Rep*, 37, 3931-3939, 2010. DOI: 10.1007/s11033-010-0050-1

33. Huang YZ, Wang J, Zhan ZY, Cao XK, Sun YJ, Lan XY, Lei CZ, Zhang CL, Chen H: Assessment of association between variants and haplotypes of the *IGF2* gene in beef cattle. *Gene*, 528, 139-145, 2013. DOI: 10.1016/j. gene.2013.07.035

34. Ma X, Guan L, Xuan J, Wang H, Yuan Z, Wu M, Liu R, Zhu C, Wei C, Zhao F, Du L, Zhang L: Effect of polymorphisms in the *CAMKMT* gene on growth traits in Ujumqin sheep. *Anim Genet*, 47, 618-622, 2016. DOI: 10.1111/age.12455

35. Duan J, Wainwright MS, Comeron JM, Saitou N, Sanders AR, Gelernter J, Gejman PV: Synonymous mutations in the human

dopamine receptor D2 (*DRD2*) affect mRNA stability and synthesis of the receptor. *Hum Mol Genet*, 12, 205-216, 2003. DOI: 10.1093/hmg/ ddg055

36. Chamary JV, Hurst LD: Evidence for selection on synonymous mutations affecting stability of mRNA secondary structure in mammals. *Genome Biol*, 6 (9): R75, 2005. DOI: 10.1186/gb-2005-6-9-r75

37. Gui L, Jiang B, Zhang Y, Zan L: Sequence variants in the bovine silent information regulator 6, their linkage and their associations with body measurements and carcass quality traits in Qinchuan cattle. *Gene*, 559, 16-21, 2015. DOI: 10.1016/j.gene.2015.01.008

38. Liu WC, Lai SJ: A synonymous mutation of *uncoupling protein 2 (UCP2)* gene is associated with growth performance, carcass characteristics and meat quality in rabbits. *J Anim Sci Technol*, 58:3, 2016. DOI: 10.1186/ s40781-016-0086-4