

Hsp-70 Gene Expression Analyses in the Different Ages of Rainbow Trout

Gonca ALAK *  Abdulkadir ÇİLTAŞ * Orhan ERDOĞAN *

[1] This work was the entire of master thesis entitled the expression analyses of Hsp-70 gene in the different ages rainbow trouts

* Atatürk University, Agriculture Faculty, Department of Fisheries, TR-25240 Erzurum - TURKEY

Makale Kodu (Article Code): KVFD-2009-757

Summary

The purpose of this study was to determinate the association between heat shock protein 70 (Hsp-70) expression and the age in rainbow trout. Total RNA was extracted with TRIzol Reagent (Life Technologies) and the first strand cDNA was synthesized using Super Script III Reverse Transcriptase. Gene amplification was done by PCR method and quantitative determination of gene expression was using beta actin gene as control. Consequently, we demonstrated that Hsp70 expression significantly increased the age.

Keywords: Rainbow trouts (*Oncorhynchus mykiss*), Age, Gene expression, Heat shock protein, Hsp70

Farklı Yaşlardaki Gökkuşuğu Alabalıklarında (*Oncorhynchus mykiss*) Hsp-70 Geninin Ekspresyon Analizi

Özet

Bu çalışmanın amacı gökkuşuğu alabalıklarında (*Oncorhynchus mykiss*) Hsp-70 geni ekspresyonu ile yaş arasındaki ilişkiyi belirlemektir. TRIzol Reagent (Life Technologies) ile toplam RNA, Super Script III Reverse Transcriptase ile de cDNA kütüphanesi hazırlandı. Gen amplifikasyonu PCR kullanılarak, ekspresyonun miktar belirlenmesi ise beta aktin kullanılarak yapıldı. Sonuçta Hsp-70 ekspresyonunun yaşla birlikte önemli oranda arttığı gözlemlendi.

Anahtar sözcükler: Gökkuşuğu alabalığı (*Oncorhynchus mykiss*), Yaş, Gen ekspresyonu, Isı şok protein, Hsp70

INTRODUCTION

Heat shock proteins (Hsps) are a group of intracellular proteins that have an unusually high degree of identity at the amino acid level, among diverse organisms. As this family of proteins is induced by stressors other than heat, they are also commonly referred to as 'stress proteins' in the literature. The term stress proteins also may refer to several other groups of proteins that respond to stressors¹.

The naming of Hsps are generally based on their molecular mass (kilodaltons, kDa). Heat shock proteins are also grouped according to function (eg chaperonin), DNA sequence, and antibody cross-reactivity. The

commonly used categories are: 100 kDa; 90 kDa; 70 kDa; 60 kDa; and the 16±30 kDa group, and are usually referred to as Hsp-100, Hsp-90, Hsp-70, Hsp-60, and the low molecular weight (LMW) class of proteins, respectively².

In the last three decades, there has been an exponential increase in the interest and research activity concerning the description, classification and functional significance of these proteins. Heat shock proteins are constitutively expressed in cells to maintain a number of critical cellular processes relating to protein folding, fidelity and translocation. These proteins also are induced

 İletişim (Correspondence)

 +90 442 2311426

 galak@atauni.edu.tr

in cells in response to a variety of stressors and enhance survival by protecting vital cellular functions¹.

Research in this area continues to grow at an ever-increasing rate and applications to problems and opportunities in human health and environmental monitoring are developing rapidly. Heat shock protein studies in fish are still in the early stages compared with those in bacteria, yeast and mammals. Studies in fish are still in the descriptive stages of documenting novel proteins that are produced in various tissues in response to a variety of biological and abiotic stressors. For example, Hsp-70, which are expressed in response to trace metal exposure, organic pollutants changes in temperature, hypoxia, exposure to ultraviolet radiation, exercise, hyperthermia, oxidative stress, metabolic challenge, ethanol, free radicals, amino acid analogs, heavy metals and aging^{3,4}.

A characteristic feature of aging is a progressive impairment in the ability to maintain homeostasis in the face of environmental challenges. Age-related decline in Hsp expression may contribute to this dysfunctional homeostasis. Specifically, Hsp mRNA and protein production in response to heat shock decreases with age in a variety of species from flies to rats⁵. However, the age-related changes in Hsp response in rainbow trout, a heavily utilized research animal, have not been reported to date. Fish have long been used as an experimental model for aging^{6,8}. The adaptability of the rainbow trout⁹ to mutagenesis studies¹⁰ makes it an attractive vertebrate model to study the aging process¹.

In this paper, we have evaluated effect of age on expression of Hsp-70 in muscle of rainbow trout in different age.

There are many studies involving Hsp-70 expression but none only aging¹¹. Nor is there any aging study relation to Hsp-70 expression in fish. In this study, there was a relationship Hsp-70 expression and age. Thus, in this study it was seen that if fish have aging some genes can be expressed in their body such as Hsp-70. Therefore this study can become a reference for future research on relation between Hsp expression and age or plan of rearing density. And it provides information on age-related cellular and molecular changes that occur in fish species.

MATERIAL and METHODS

Fish Material

The study used different ages (8, 18 and 36 months) rainbow trout (*Oncorhynchus mykiss*), obtained from

Ataturk University, Agricultural Faculty, Fishery Department Research and Extension Center Faculty of Agriculture. They were stabulated, with inconsistent mortality, at biomass density (250 kg/m³) in a 10*2*1.3 m³ pond connected to a water recycling system served with about 24 water refilling per day. Water parameters were strictly controlled: temperature 10±1°C, total hardness 102 mg as CaCO₃, free O₂ 8±0.5 ppm, pH 7.8. At the average size of 8 months 150 g, 18 months 500 g and 36 months 1500 g. Food was automatically distributed daily ensured as a 1.5% body mass ratio, with extruded pellet for marine fish (Bleacksea Food). Temperature, O₂ and pH were continuously monitored, while other parameters were analyzed at a weekly rate. After 15 days, three fish for each group was randomly sampled, immediately stunned and slaughter. Skeletal muscle were removed from dorsal region, frozen in liquid N₂ and stored at 80°C until molecular biology analyses.

RNA Preperation and cDNAs Synthesis

Total RNA was isolated using TRIzol® Reagent according to the manufacturer's recommendation (Invitrogen), and the RNA pellets were resuspended in TE buffer (Tris-EDTA, pH 8.0). To eliminate possible genomic DNA contamination, the RNA samples were treated with a DNase I (Sigma) by the manufacturer's instructions. The total RNA was quantified by measuring the absorbance at 260 nm using a UV-VIS Spektrofotometre (Aquamate), and the purity was assessed by determining the ratio of the absorbance at 260 and 280 nm. All samples had 260/280 nm ratios <1.6. Additionally, the integrity of the RNA preparations was verified by visualization of the 18S and 28S ribosomal bands stained with ethidium bromide after electrophoresis on 1.0% agarose gels. Total RNA was reverse transcribed using a commercially available cDNA synthesis kit SuperScript™ III Reverse Transcriptase (Invitrogen). cDNA was stored at -20°C.

Quantitative RT-PCR

For PCR reaction ~100 ng template, 1X PCR buffer (1.5 mM MgCl₂), 200 µL of each dNTP, 0.2 µL gene-specific hsp-forward (5'- TGCACCTAGGTTTTTCATAGAAT - 3') and hsp-reverse (5'- ATGGAGGTGTAGAAGTCGATGC- 3') primers, and 2.5 units of Taq DNA polymerase were mixed for a total reaction volume of 15 µL. Thermal cycling conditions were as follows: initial activation at 80°C for 1 min, at 94°C for 3 min, 40-PCR cycles at 94°C for 45s, 58°C for 45s, 72°C for 45s, and a final extension at 72°C for 5 min. For actin gene, Primers actinF (5'TGGGGCAGTATGGCTTGTATG3') and actinR (5' CTCTGGCACCCCT AATCACCTCT-3') were used as the internal control¹¹. PCR cycles for actin gene was in initial

activation at 80°C for 1 min, at 94°C for 3 min, 35-PCR cycles at 94°C for 20s, 54°C for 20s, 72°C for 20s, and a final extension at 72°C for 5 min. The PCR products were loaded into 1% agarose gel and run in TAE 1X buffer at 100 mV for 30 min. A distinct band estimated at ~954bp nucleotides (Hsp) and ~165bp (actin) nucleotides were generated and amplified products quantified by using ImageJ 1.37c (<http://rsb.info.nih.gov/ij/>).

Statistical Analysis

The Student's t-test and standard deviation were used for data analysis. Significance of differences was defined as $P < 0.01$ ¹².

RESULTS

In the present study, we have obtained for Hsp-70 (Gen Bank accession number AB062281), and we have completed a cDNA of 3746 bp by SuperScript™ III Reverse Transcriptase (Invitrogen). We amplified a section of 954 bp of Hsp-70 gene and comprehensively compared their mRNA amplified products quantified by using ImageJ

1.37c. β -actin was amplified as 165 bp and used as a control ¹³, Fig. 1.

Quantitative mRNA levels of Hsp-70 were plotted by using gel density in 40th PCR cycle. β -actin was used as control (Fig. 2). As far as quantitative mRNA is concerned, the results show that age fairly effect on fish. If it is compared with all age, quantitative mRNA levels more than ten times as high in 36 months as other age.

DISCUSSION

There are many experiments about Hsp on organisms. The expression of Hsp-70 has been studied in relation to various stressors such as heat shock ¹⁴⁻¹⁹, aquatic toxicants ²⁰⁻²⁵ and psychological stress ^{26,27}. In all of this studies, Hsp expression has been increased dramatically similar our results. However, the expression of various types of Hsp protein and mRNA levels in response to heat stress has been shown to decrease with age ^{18,28-33}. Pahlavani ³⁴, showed that the induction of Hsp-70 expression by heat shock decreases 40-50% with age. There are only a few reports on the effects of aging on

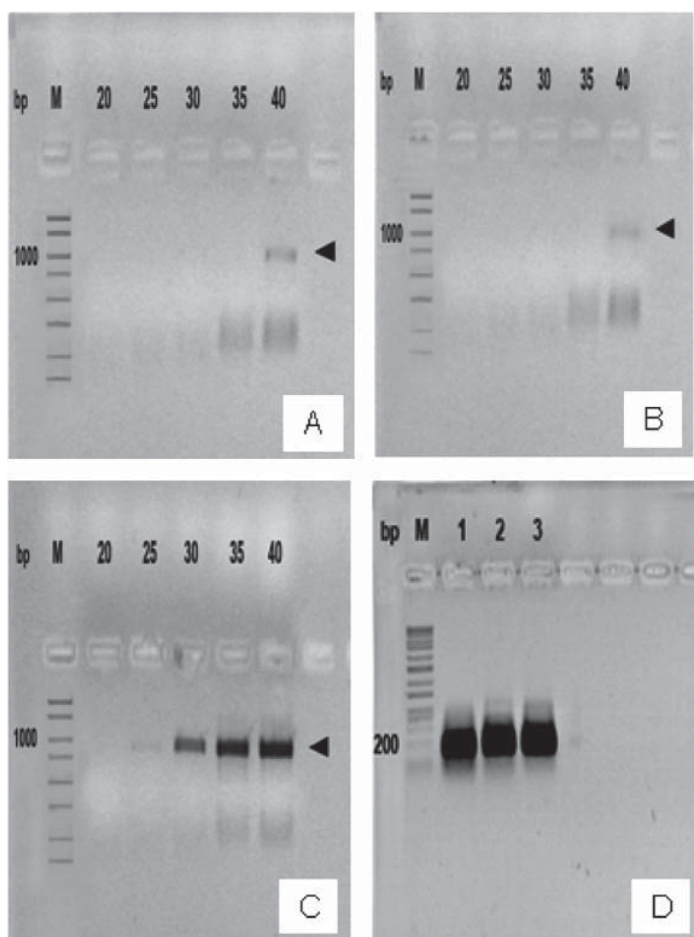


Fig 1. HSP-70 gene agarose gel patterns, 8 months (A), 18 months (B), 36 months (C), 20-40: PCR cycling (A,B,C); β -actin control: Line 1: 8 months, Line 2: 18 months, Line 3: 36 months (D)

Şekil 1. HSP-70 geninin agaroz jel görüntüleri; 8 aylık (A), 18 aylık (B), 36 aylık (C), 20-40 cycle (A,B,C); beta aktin kontrol: 1. Sıra: 8 aylık, 2. Sıra: 18 aylık, 3. Sıra : 36 aylık (D)

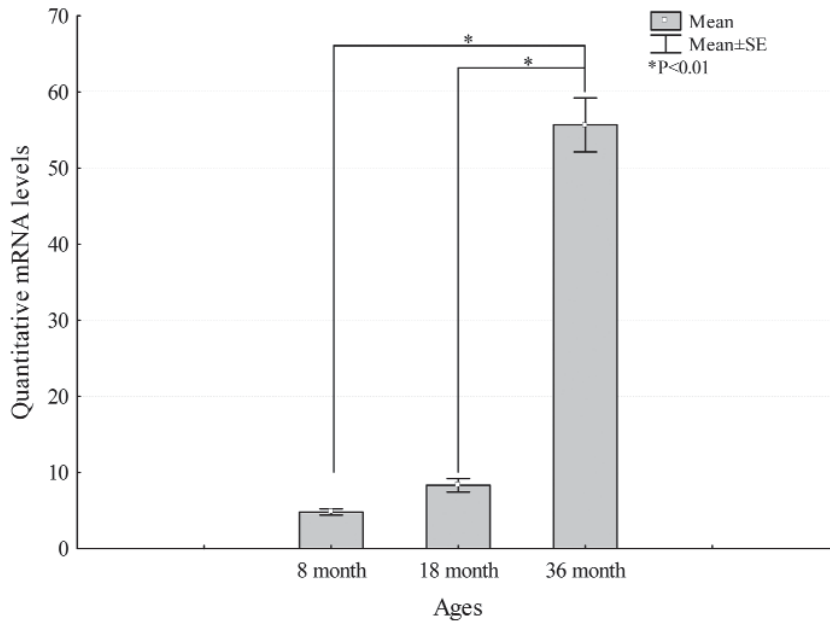


Fig 2. Relatives expression of Hsp-70 in sketal muscle of different ages groups. Asterisks denotes significant difference from age groups ($P < 0.01$). $n=7$

Şekil 2. Hsp-70 geninin farklı yaş gruplarında ekspresyon ilişkisi. Şekilde yaş grupları arasında önemli farklılıklar belirlenmiştir ($P < 0.01$). Örnek sayısı=7

fish⁶⁻⁸. These studies provide only limited information on age-related cellular and molecular changes that occur in fish species.

There are many studies involving Hsp expression. But, there isn't any experiment related with heat shock protein and age. Thus, the purpose of the present study was to isolate Hsp genes from rainbow trout and to analyze their expression profiles at the mRNA levels. In this regard, we quantitatively compared the mRNA levels between different ages under rearing the same condition using PCR analysis. The present work demonstrated the expression of Hsp-70 gene was increased with fish age.

In this study the effect of age on Hsp-70 gene expression in different age fish was investigated. Consequently, Hsp-70 gene expression increase with fish age.

REFERENCES

1. Iwama GK, Thomas PT, Forsyth RB, Vijayan MM: Heat shock protein expression in fish. *Rev Fish Biol Fish*, 8, 35-56, 1998.
2. Morimoto RI, Tissieres A, Georgopoulos C: The Biology of Heat Shock Proteins and Molecular Chaperones (Cold Spring Harbor Monograph Series, Vol. 26), p. 496, Plainview, Cold Spring Harbor Laboratory Press, NY, 1994.
3. Kregel KC: Molecular biology of thermoregulation invited review: Heat shock proteins: Modifying factors in physiological stress responses and acquired thermotolerance. *J Appl Physiol*, 92, 2177-2186, 2002.
4. Lewis S, Handy RD, Cordi B, Billingham Z, Depledge MH: Stress proteins (HSP's): Methods of detection and their use as an environmental biomarker. *Ecotoxicology*, 8, 351-368, 1999.
5. Soti C, Csermely P: Aging and molecular chaperones. *Exp Gerontol*, 38, 1037-1040, 2003.
6. Patnaik BK, Mahapatro N, Jena BS: Ageing in fishes. *Gerontology*, 40, 113-132, 1994.
7. Woodhead AD: Fish in studies of aging. *Exp Gerontol*, 13, 125-140, 1978.
8. Woodhead AD: Aging, the fishy side: An appreciation of Alex Comfort's studies. *Exp Gerontol*, 33, 39-51, 1998.
9. Kubilay A, Altun S, Didinen BI, Ekici S, Diler Ö: Gökkuşluğu alabalığı (*Oncorhynchus mykiss*) işletmelerinde *Flavobacterium psychrophilum* izalasyonu. *Kafkas Univ Vet Fak Derg*, 15, 709-715, 2009.
10. Knapik EW, Goodman A, Ekker M, Chevrette M, Delgado J, Neuhaus S, Shimoda N, Driever W, Fishman MC, Jacob HJ: A microsatellite genetic linkage map for zebrafish (*Danio rerio*). *Nature Genetics*, 18, 338-343, 1998.
11. Erdogan O, Atamanalp M, Sisman T, Aksakal E, Alak G: Effects of 2,2-Dichlorovinyl Dimethyl Phosphate (DDVP) on Hsp-70 Gene Expression in Rainbow Trout. *Israeli Journal of Aquaculture-Bamidgeh*, 59 (4): 230-234, 2007.
12. Ergün G, Aktaş S: ANOVA modellerinde kareler toplamı yöntemlerinin karşılaştırılması. *Kafkas Univ Vet Fak Derg*, 15 (3): 481-484, 2009.
13. Ojima N, Yamashita M, Watabe S: Quantitative mRNA expression profiling of heat-shock protein families in rainbow trout cells. *Biochem Biophys Res Commun*, 329, 51-57, 2005.
14. Lele Z, Engel S, Krone PH: Hsp47 and Hsp70 gene expression is differentially regulated in a stress- and tissue-specific manner in Zebrafish embryos. *Developmental Genetic*, 21, 123-133, 1997.
15. Currie S, Moyes CD, Tufts BL: The effects of heat shock

and acclimation temperature on Hsp70 and Hsp30 mRNA expression in rainbow trout: *In vivo* and *in vitro* comparisons. *J Fish Biol*, 56, 398-408, 2000.

16. Palmisano AN, Winton JR, Dickhoff WW: Tissue-specific induction of hsp90 mRNA and plasma cortisol response in Chinook salmon following heat shock, seawater challenge, and handling challenge. *Mar Biotechnol*, 2, 329-338, 2000.

17. Mesa MG, Welland LK, Wagner P: Effects of acute thermal stress on the survival, predator avoidance, and physiology of juvenile fall chinook salmon. *Northwest Sci*, 76, 118-128, 2002.

18. Murtha JM, Keller ET: Characterization of the heat shock response in mature Zebrafish (*Danio rerio*). *Exp Gerontol*, 38, 683-691, 2003.

19. Bowen L, Werner I, Johnson ML: Physiological and behavioral effects of zinc and temperature on coho salmon (*Oncorhynchus kisutch*). *Hydrobiologia*, 559, 161-168, 2006.

20. Williams JH, Farag AM, Stansbury MA, Young PA, Bergman HL, Petersen NS: Accumulation of Hsp70 in juvenile and adult rainbow trout gill exposed to metal contaminated water and or diet. *Environ Toxicol Chem*, 15, 1324-1328, 1996.

21. Vijayan MM, Pereira C, Kruzynski G, Iwama GK: Sublethal concentrations of contaminant induce the expression of hepatic heat shock protein 70 in two salmonids. *Aquat Toxicol*, 40, 101-108, 1998.

22. Ar-T-Ar'Ssa S, Ausseil O, Palluel O, Vindimian E, Garnier-La J, Porcher JM: Biomarker responses in juvenile rainbow trout (*Oncorhynchus mykiss*) after single and combined exposure to low doses of cadmium, zinc, PCB77 and 17 β -oestradiol. *Biomarker*, 8, 6-9, 2003.

23. Weber LP, Diamond SL, Bandiera SM, Janz DM: Expression of HSP70 and CYP1A protein in ovary and liver of juvenile rainbow trout exposed to betanaphthoflavone. *Comp Biochem Physiol C Toxicol Pharmacol*, 131, 387-94, 2002.

24. Feng Q, Boone AN, Vijayan MM: Copper impact on heat shock protein 70 expression and apoptosis in rainbow trout

hepatocytes. *Comp Biochem Phys C*, 135, 345-355, 2003.

25. Shen H, Wang X.-R, Zhang JF, Liu H, Zhao YJ: Western blotting responses of heat shock protein (HSP70) in the liver of young fish, *Carassius auratus* to lower concentration of zinc (Zn super [2+]). *J Agro-Environ Sci*, 23, 441-443, 2004.

26. De Wachter B, Scholliers A, Blust B: Semiquantitative immunoblot detection of 70kDa stress proteins in the carp *Cyprinus carpio*. *Bull Environ Contam Toxicol*, 60, 37-44, 1998.

27. Gornati R, Papis E, Rimoldi S, Terova G, Saroglia M, Bernardini G: Rearing density influences the expression of stress-related genes in sea bass (*Dicentrarchus labrax*, L.). *Gene*, 341, 111-118, 2004.

28. Pardue S, Groshan K, Raese JD, Morrison-Bogorad M: Hsp70 mRNA induction is reduced in neurons of aged rat hippocampus after thermal stress. *Neurobiol Aging*, 13, 661-672, 1992.

29. Heydari AR, Wu B, Takahashi R, Strong R, Richardson A: Expression of heat shock protein 70 is altered by age and diet at the level of transcription. *Mol Cell Biol*, 13, 2909-2918, 1993.

30. Rogue PJ, Ritz MF, Malviya AN: Impaired gene transcription and nuclear protein kinase C activation in the brain and liver of aged rats. *FEBS Lett*, 334, 351-354, 1993.

31. Wu B, Gu MJ, Heydari AR, Richardson A: The effect of age on the synthesis of two heat shock proteins in the hsp70 family. *J Gerontol*, 48, 50-56, 1993.

32. Locke M, Tanguay RM: Diminished heat shock response in the aged myocardium. *Cell Stress Chaperones*, 1, 251-260, 1996.

33. Keller ET, Murtha JM: The use of mature zebrafish (*Danio rerio*) as a model for human aging and disease. *Comp Biochem Phys C*, 138, 335-341, 2004.

34. Pahlavani MA, Harris MD, Moore SA, Weindruch R, Richardson A: The expression of heat shock protein 70 decreases with age in lymphocytes from rats and rhesus monkeys. *Exp Cell Res*, 218, 310-318, 1995.