

Changes in Antioxidant Sytem, Lipid Peroxidation, Heart and Respiratory Rate and Rectal Temperature with Ketamine and Ketamine-Xylazine Anaesthesia in Tuj Rams

Nadide Nabil KAMILOĞLU *  Alkan KAMILOĞLU** Ebru BEYTUT*

* Department of Physiology, Faculty of Veterinary Medicine, University of Kafkas, Kars - TURKEY

** Department of Surgery, Faculty of Veterinary Medicine, University of Kafkas, Kars - TURKEY

Yayın Kodu (Article Code): 2008/101-A

Summary

The objective of this study was to determine if a ketamine or ketamine-xylazine had any effect on the antioxidant defence system and on lipid peroxidation as thiobarbituric acid substance (TBARS) in rams. The experiment was carried out on 16 clinically healthy Tuj rams each weighing an average 56-60 kg and aged 3-4 years old. Rams were divided into two groups of eight ram in each. To produce anaesthesia the Group I was given 10 mg/kg ketamine HCl, the Group II received 50 µg/kg xylazine HCl and 10 mg/kg ketamine HCl, intramuscularly. Blood samples were taken from the jugular vein before and at 5, 10, 15, 30 and 60th minutes after drugs injection to measure the levels of vitamin E and TBARS in plasma and the activity of glutathione peroxidase (GSHPX) and the levels of glutathione (GSH) in erythrocytes. Heart and respiratory rates and rectal temperature were also recorded for 60 min. in two groups. Although plasma levels of TBARS and vitamin E were significantly ($P<0.001$) lowered at the 15th min. after ketamine injection, the activity of GSHPx and levels of GSH in RBC were significantly increased at the 10th min. after ketamine injection ($P<0.001$) in Group I. While significant changes were observed in heart rate and respiratory rate at the 5th minute after anaesthesia. Also, the activity of GSHPx and levels of GSH in RBC in Group II were significantly increased at the 15th min. after ketamine-xylazine injection ($P<0.001$). Levels of TBARS in plasma and vitamin E in plasma were not changed with ketamine-xylazine injection. Heart and respiratory rates began to increase at the 10th min. after ketamine-xylazine injection and decreased to a value approximately that of before analgesia by the 15th min. and remained stable throughout the anaesthesia. The plasma levels of TBARS in the ketamine group were lower than those in the ketamine-xylazine group at the 15 and 30th min. after injections ($P<0.001$). GSH levels and GSHPx activity in RBCs of rams have a similar increase in two groups. There were no significant differences observed in rectal temperature during anaesthesia in Group I and II. In conclusion, the results of this study shows that ketamine and kombination of ketamine-xylazine injection at a therapeutic concentration can suppress free radical generation and support antioxidant system of rams.

Keyword: Ketamine, Xylazine, Ram, Lipid peroxidation, Antioxidants

Koyunlarda Ketamin ve Ketamin-Xylazin Anestezisi ile Eritrosit Antioksidan Sistemi, Lipid Peroksidasyon, Kalp Hızı, Solunum Sayısı ve Rektal Isıda Meydana Gelen Değişiklikler

Özet

Bu çalışma, ketamin ve ketamin-xylazinin koyunlarda kullanılan anestezi dozunun plazma ve eritrosit antioksidan sistemi ve thiobarbituric acid substance (TBARS) düzeyleri üzerine etkilerini belirlemek amacıyla 3-4 yaşında, ortalama 56-60 kg ağırlığında 16 klinik olarak sağlıklı koç üzerinde yürütüldü. Her grupta 8 koç olacak şekilde iki grup oluşturuldu. Anestezisi oluşturmak amacıyla I. Gruba 10 mg/kg ketamin HCl, II. Gruba 10 mg/kg ketamin HCl ve 50 µg/kg Xylazin HCl IM yolla enjekte edildi. Plazma E vitamini ve TBARS olarak lipid peroksidasyon düzeylerini, glutatyon peroksidaz (GSHPX) aktivitesini ve eritrosit glutatyon (GSH) düzeylerini belirlemek için, kan numuneleri ketamin ve ketamin-xylazin enjeksiyonundan önce ve 5, 10, 15, 30 ve 60. dakikalarda v. jugularis'ten alındı. Kalp hızı, solunum sayısı ve rektal ısı da aynı zaman aralıklarında 60 dak süresince belirlendi. I. Grupta plazma TBARS ve E vitamini düzeylerinin ketamin enjeksiyonundan 15 dak sonra azaldığı ($P<0.001$) gözlenirken, eritrosit GSHPx aktiviteleri ve GSH düzeylerinin ketamin enjeksiyonundan 10 dak sonra önemli düzeyde arttığı ($P<0.001$) belirlendi. I. Grupta, kalp hızı ve solunum sayısının anestezinin 5 dak sonra azaldığı gözlenirken, rektal ısıda bir farklılık tespit edilmedi. II. Grupta eritrosit GSHPx aktiviteleri ve GSH düzeylerinin ketamin-xylazin enjeksiyonundan 15 dak sonra arttığı belirlenirken, plazma E vitamini ve TBARS düzeylerinin ketamin-xylazin enjeksiyonu ile değişmediği belirlendi. Plazma TBARS düzeyleri I. Grupta anestezinin 15 ve 30. dak'larında II. Gruba göre daha düşük bulundu ($P<0.001$). GSHPx aktiviteleri ve GSH düzeylerinin her iki grupta da benzer şekilde arttığı gözlemlendi. II. Grupta kalp hızı ve solunum sayısının anestezinin 10. dak'sına kadar arttığı, ancak 15. dak'dan sonra başlangıç değerlerine döndüğü ve anestezisi süresince bu değerlerde bir değişiklik belirlenmediği gözlenirken, rektal ısıda bir farklılık saptanmadı. Çalışma sonuçları, ketamin ve ketamin-xylazin enjeksiyonunun serbest radikal oluşumunu baskıladığını ve antioksidan sistemi desteklediğini göstermektedir.

Anahtar sözcükler: Ketamin, Xylazin, Koyun, Lipid Peroksidasyon, Antioksidanlar



İletişim (Correspondence)



+90 474 2426807/1144



nkamiloglu@hotmail.com

INTRODUCTION

In clinical veterinary practice there is demand for a safe and reliable injectable method of anaesthesia for use in routine surgery of short to intermediate duration. Ketamine, a dissociative anaesthetic, have been used for this purpose by intra venously and intra muscularly. Ketamine produce poor muscle relaxation, good somatic analgesia and an increased sympathetic tone. The major problems occur during recovery and include tonic-clonic cramps and convulsion-like signs and also increases blood pressure. However, it does not affect heart rate or respiratory rate when used for general anaesthesia or premedication ^{1,2}.

On the other hand, evidences suggest some interactions between anaesthetic agents and reactive oxygen species (ROS) ³. Also, some anaesthetic agents which used in routine surgery such as ketamine, thiopental, midazolam and propofol were investigated in terms of antioxidative effects and effects on lipid peroxidation production. Clinical plasma concentrations of ketamine have minimal effects on ROS production ⁴. Lupp et al.⁴ suggested that ketamine act as a radical scavenger and inhibitor of the oxidative function of microsomal cytochrome P₄₅₀ that has a role in the metabolism of endogenous pools of arachidonic acid and also play important physiological roles in the control of tissue and body homeostasis. Studies from several laboratories have suggested important roles for the P₄₅₀ arachidonic acid monooxygenase in the pathophysiology of experimental hypertension. Also, it is reported that ketamine inhibit nitric oxide synthase activity in rat brain ⁵.

An alpha 2 agonist-xylazine combination with ketamine is often used in veterinary anaesthesiology for anaesthetizing or immobilizing laboratory or domestic animals ^{6,7}. Depending on the dose of ketamine has sedative, analgetic, psychomimetic, cataleptic and anaesthetic effects. It stimulates blood circulation ⁸. It does not significantly alter the pattern of breathing ⁸. Xylazine markedly reduces ketamine dosage and the subsequent reduction of the undesirable psychomimetic effects ⁹. It markedly potentiates hypnotic and analgetic effect of ketamine ¹⁰. Breathing is slightly influenced when clinical doses are used ¹¹. Xylazine reduce the dose of ketamine by 40–60%, induce muscle relaxation and reduce the occurrence of psychomotor symptoms after ketamine. It is known that,

breathing is not significantly influenced with this combination and ketamine-xylazine anaesthesia so reliable with respect to the incidence of cardiac arrhythmias ^{6,12}.

Therefore this study was undertaken in order to evaluate the effect of ketamine and combination of ketamine-xylazine anaesthesia on the antioxidant system and lipid peroxidation in rams.

MATERIAL and METHODS

The experiment was carried out on 16 clinically healthy male Tuj sheep, each weighing an average 56-60 kg and aged 3-4 years old. The sheepes were divided into two groups of eight sheep in each. To produce sedation the first group (Group I) was given 10 mg/kg ketamine HCl (Ketamine, Parke Davis, Eczacıbaşı), the other group (Group II) received 50 µg/kg xylazine HCl (%2 Alfazyne, Alfasan) injectable solution and 10 mg/kg ketamine hydrochloride (Ketamine, Parke Davis, Eczacıbaşı) five min after xylazine administration. Drugs were injected intra muscularly into the rams.

Blood samples were taken from the jugular vein before ketamine injection and at 5, 10, 15, 30, 45 and 60 minutes to measure the levels of vitamin E and lipid peroxidation (LPO) in plasma and the activity of glutathione peroxidase (GSHPX; EC: 1.11.1.9) and the levels of glutathione (GSH) in erythrocytes.

Hearth rate (beats/per min), respiratory rate (breaths/ per min) and rectal temperature were recorded for each animal at the above mentioned time intervals. Hearth rate was calculated from ECG records. The ECGs were recorded by a direct writing electrocardiograph. All ECGs were standardized at 10 mm/mV, with chart speed of 25 mm/min. The rectal temperature was obtained by a thermometer placed into the rectum. Respiratory rate was determined by direct observation. Motor functions were tested as full response to needle prick to various areas of the body. Sensory functions were tested as response to palpebral and corneal reflexes.

Blood was collected using heparinized vacutainer tubes. The plasma and the red blood cells (RBC) were separated by centrifugation (2500 g, for 15 min at 4°C). The plasma was frozen (-20°C) until

further analysis. The RBC samples were washed three times with 0.9% sodium chloride and then haemolysed by exposure to a nine-fold volume of distilled water followed by freezing (-20°C for 18 h) and thawing before analysis. The plasma specimens were used for the determination of vitamin E and LPO, and the RBC samples were used for the determination of GSHPX and GSH.

The end product of polyunsaturated fatty acid peroxidation, malondialdehyde (MDA), reacting with thiobarbituric acid (TBA) in serum samples was determined by the method of Matkovichs et al.¹³. The values of MDA reactive material (TBARS) were expressed in terms of the malondialdehyde (MDA) content (nmol/ml plasma), which served as a standard of 1,1,3,3-tetraethoxypropane (Sigma, Chemical Company St. Louis, MO, USA).

The vitamin E (α -tocopherol) levels of plasma were determined in the frozen serum samples by the method described by McMurray et al.¹⁴. The relevant wavelengths for vitamin E detection were 292 and 330 nm. Calibration was performed using a standard solution of α -tocopherol in methanol.

The GSH levels of haemolysed RBC were

measured spectrophotometrically using Ellman's reagent¹⁵. GSH-Px (EC 1.11.1.9) activity was determined using cumene hydroperoxide and reduced GSH as co-substrates, and the loss of GSH following enzymic reaction was measured spectrophotometrically with Ellman's reagent at 37°C and 412 nm according to Lawrence and Burk¹⁶. The haemoglobin concentration in lysed erythrocytes was determined by the cyanmethemoglobin method¹⁷.

Measurements were compared with baseline values (time: 0) by performing One-way parametric ANOVA test and post-hoc differences in variables between groups performed by Tukey test using MINITAB statistical program (Minitab,1998). All results were expressed as the mean \pm standard deviation (SD). P value <0.05 was considered to be statistically significant

RESULTS

Plasma TBARS and vitamin E concentrations and erythrocytes GSH levels and GSHPx activity according to time and to the experimental groups are shown in the *Figures 1. A, B, C and D* respectively.

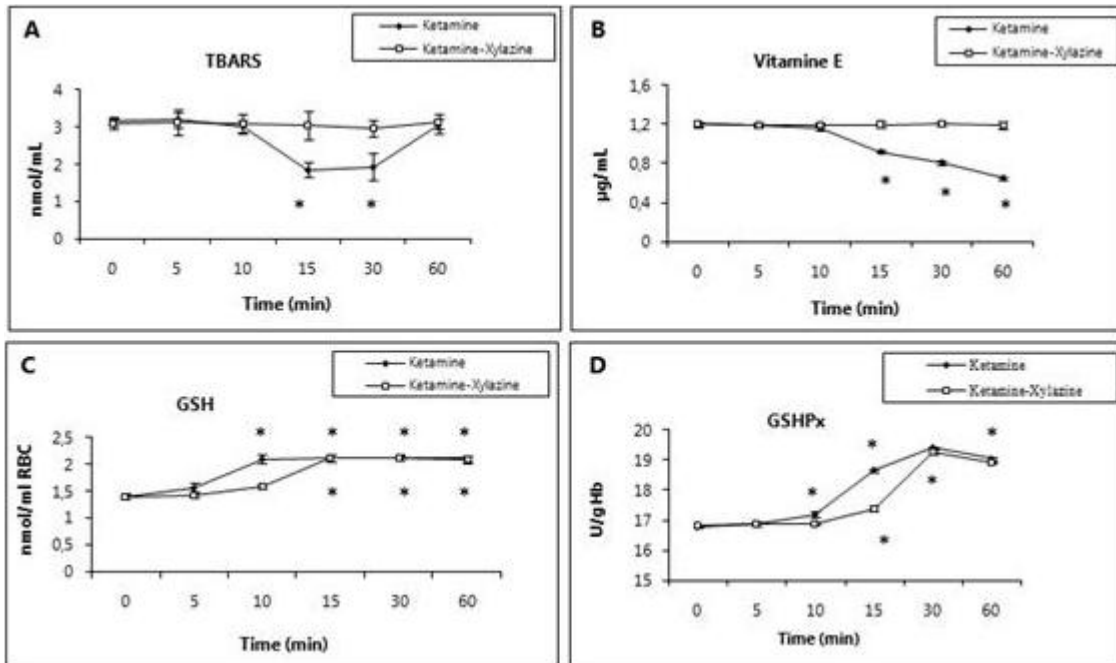


Fig 1. Line graphs showing the plasma TBARS and vitamin E concentrations and erythrocytes GSH levels and GSHPx activity at different time points from the experimental groups. * P<0.001 in comparison to the 0th point of the time. Values are expressed as mean \pm SD

Şekil 1. Grafikler ketamin ve ketamin-ksilazin enjeksiyonu ile plazma TBARS ve E vitamini düzeyleri ile eritrosit GSH ve GSHPx aktivitelerinde zamana göre belirlenen değişiklikleri göstermektedir. *: P<0.001 (0. dakikaya göre istatistiksel önemi göstermektedir. Değerler Ortalama \pm Standart Sapma olarak verilmiştir

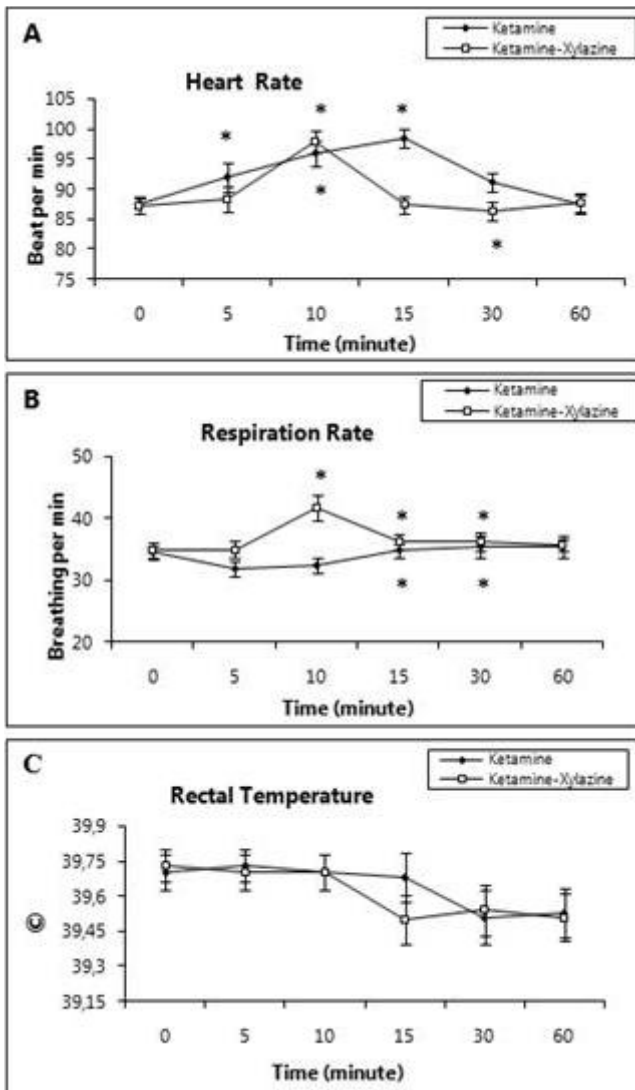


Fig 2. Line graphs showing the alteration in heart rate, respiratory rate and rectal temperature at different time points from the experimental groups. * $P < 0.001$ in comparison to the 0th point of the time. Values are expressed as mean \pm SD

Şekil 2. Grafikler ketamin ve ketamin-ksilazin enjeksiyonu ile kalp hızı, solunum sayısı ve rektal ısıda zamana göre belirlenen değişiklikleri göstermektedir. * : $P < 0.001$ (0. dakikaya göre istatistiksel önemliliği göstermektedir. Değerler Ortalama \pm Standart Sapma olarak verilmiştir

Although plasma levels of TBARS and vitamin E content was significantly ($P < 0.001$) lowered at the 15th minute after ketamine injection, the activity of GSHPx and levels of GSH in RBC were significantly increased at the 10th minute after ketamine injection ($P < 0.001$) in Group I when compared baseline values. Also, the activity of GSHPx and levels of GSH in RBC in Group II were significantly increased at the 15th minute after ketamine-xylazine injection ($P < 0.001$). Levels of TBARS in plasma and vitamin E in plasma were not changed with ketamine-xylazine injection in comparison with baseline values. On the other

hand when compared groups, plasma levels of vitamin E were higher than in the ketamine-xylazine group than in the ketamine group at the 15, 30, 60th min after anesthesia ($P < 0.001$). TBARS concentration remained relatively constant in the two groups until 15th min. The plasma levels of TBARS in the ketamine group were lower than those in the ketamine-xylazine group at the 15 and 30th min after injections ($P < 0.001$) GSH levels and GSHPx activity in RBCs of rams have a similar increase in two groups. There is no significant difference between groups in GSH levels. GSHPx activity in RBCs of rams in the ketamine group higher than that in the ketamine-xylazine group only at the 15th min after anesthesia.

Figure 2 (A, B and C) shows the alteration in heart rate, respiratory rate and rectal temperature before and after injections in the experimental groups. In Group I, significant increase were observed in heart rate and respiratory rate at the 5th minute after anesthesia, then decreased to a value approximately that of the control (0. min) by the 60th min. On the other hand, in Group II, heart and respiratory rates began to increase at the 10th minutes after ketamine-xylazine injection and decreased to a value approximately that of before analgesia by the 15th min and remained stable throughout the anaesthesia. There were no significant differences observed in rectal temperature during anaesthesia in Group I and II.

DISCUSSION

In this study, we analyzed the effect of widely used dose of ketamine and combination of ketamine/xylazine anesthesia on oxidant-antioxidant parameters as well as the effects of these agents on cardio-pulmoner activity in sheep. It is known that a cyclohexylamine analogue, ketamine, produces a cataleptoid state involving unconsciousness and somatic analgesia but no muscular relaxation. Also, ketamine induces anaesthesia rapidly and causes minimal depression of the respiratory and cardiovascular systems, and has a wide margin of safety^{18,19}. Because of these advantages of ketamine, it is widely used in veterinary practise by alone or in combination with xylazine¹². On the other hand, xylazine is a potent hypnotic with powerful central muscular relaxant properties. Its main disadvantages are that it produces significant cardiac arrhythmias in all

species, interfering with normal electrical activity in the heart⁷. This study showed that respiration and heart rate changed significantly ($P < 0.001$) after ketamine injection. However, from the clinical point of view, cardiovascular stability is good and breathing is not significantly influenced with combination of ketamine/xylazine. Similar to results of previous studies^{6,9,20}, it is observed that combination of ketamine/xylazine effectively reduced some of the undesirable effects of ketamine, such as tachycardia, apnea on cardio-pulmonary activity. However, neither ketamine nor combination of ketamine-xylazine effected rectal temperature.

Oxygen free radicals are formed continuously in the organism as part of enzymatic reactions or as a product of oxidation processes. They play an important role in various pathophysiological events, including respiratory distress and myocardial vasoconstriction²¹. In sheep, however, there is little information available on oxidant-antioxidant change with the anaesthesia induced by ketamine and a combination of xylazine and ketamine. A study by Berlinskii and Berlinskii²² suggested that ketamine in combination with clofelin helped reduce the activity of free radical lipid peroxidation generation and showed minimal changes in the activities of red cell peroxide resistance, also lowest level of accumulation of MDA in children. Moreover, ketamine has been shown to be a cytochrome P₄₅₀ inducer. Also, Lupp et al.⁴ demonstrate that ketamine decreased LPO and H₂O₂ production and also act as radical scavengers and inhibitor of the oxidative function of P₄₅₀. On the other hand, there is strong evidence of formation of free radicals during ketamine use that are responsible for increased lipid peroxidation process and products in serum²³. In addition, exposure to an anaesthetic agent, may reduce flow of body and pulmonary oxygen content, as seen in xylazine anaesthesia^{24,25}. Cases of pulmonary oedema and death have been reported after administration of xylazine in sheep (26). Pulmonary oedema caused by increased permeability is characterized by the presence of cellular damage and toxicity mediated by oxygen free radicals²⁷. However, there is limited study about antioxidant activity of combination of ketamine/xylazine. A study by Helmer et al.²⁸ suggest that xylazine/ketamine combination is capable of diminishing lipopolysaccharide-induced nitric

oxide synthase in vivo. In our study, we found that while ketamine reduce TBARS production, combination of ketamine/xylazine did not affect on lipid peroxidation. This result may be the result of an antioxidant activity of ketamine and also the similar response to increased oxidative stress of this combination.

Oxidative stress occurs when production of active oxidants overwhelms the antioxidant defense mechanisms^{29,30}. Cells are protected from the damaging effects of oxidant by superoxide dismutase (SOD), glutathione peroxidase (GSHPx) and catalase (CAT). GSHPx plays a major role in detoxification of lipid peroxides that are formed in vivo especially in the absence of vitamin E and other antioxidants or in animals undergoing oxidative stress. GSHPx system may be a compensatory physiological adaptive response to reduce the oxidative stress. Increased GSHPx activity may be due to the quantity of H₂O₂ and other organic peroxides. Also, increase of GSHPx activity protected the cells from the formation of lipid peroxides by reducing H₂O₂ levels which in turn attenuated OH• generation. Reduced GSH serves as substrate for GSHPx providing reducing equivalents for the metabolism of H₂O₂ and lipid peroxides. It is the major non-protein thiol in mammalian cells and tissues and also plays an important role in the cellular defenses against free radicals and peroxides^{31,32}. Therefore, the role of GSH in the detoxification of highly reactive oxygen species (ROS) is important³³. The present study found a significant increase ($P < 0.001$) in GSHPx activity and levels of GSH 10-15 min after ketamine and combination of ketamine/xylazine injection in RBC of rams respectively. This increase may be explained by antioxidant activity of this anaesthetic agent prevent the utilization of antioxidant enzyme.

Our literature review found no studies related to the levels of vitamin E in plasma during anaesthesia or analgesia. Vitamin E has been known an essential structural component of biological membranes contributing to their stability. This lipid-soluble compound is needed for the mitochondria electron-transport function and prevents oxidation of various compounds. Also, vitamin E diminishes the peroxidation of PUFA through scavenging and chain breaking free radicals³². We showed in our results that oxidative stress may have been potentiated by diminished

vitamin E concentration with ketamine anesthesia. Stable levels of vitamin E during xylazine/ketamine anesthesia may be explained by the antioxidant properties of this combination suppress the utilization of these antioxidants.

In conclusion, the results of this study shows that ketamine and combination of ketamine-xylazine injection at a therapeutic concentration can suppress free radical generation and support antioxidant system of rams.

REFERENCES

- Haskins SC, Farver TB, Patz JD:** Ketamine in dogs. *Am J Vet Res*, 46 (9):1855-1860, 1985.
- Lumb WV, Jones EW:** Veterinary Anesthesia. Third ed., Williams & Wilkins, 1996.
- Johns RA:** Endothelium, anesthetics, and vascular control. *Anesthesiol*, 79 (6):1381-1391, 1993.
- Lupp A, Kerst S, Karge E, Quack G, Klinger W:** Investigation on possible antioxidative properties of the NMDA-receptor antagonists ketamine, memantine, and amantadine in comparison to nicanartine in vitro. *Exp Toxicol Pathol*, 50 (4-6): 501-506, 1998.
- Galley HF, Webster NR:** Brain nitric oxide synthase activity is decreased by intravenous anesthetics. *Anesth Analg*, 83(3):591-4, 1996.
- Lin HC, Purohit RC, Powe TA:** Anesthesia in sheep with propofol or with xylazine-ketamine followed by halotene. *Vet Surg*, 26 (3):247-252, 1997.
- Maze M, Tranquilli W:** Alpha-2-adrenoceptor agonists: defining the role in clinical anesthesia. *Anesthesiology*, 74, 581-605, 1991.
- Aantaa R, Scheinin M:** Alfa-2-Adrenergic agents in anaesthesiology. *Acta Anaesthes Scand*, 37, 433-448, 1993.
- Coulson NM, Januszkiewicz AJ, Dodd KT, Ripple GR:** The cardiorespiratory effects of diazepam-ketamine and xylazine-ketamine anesthetic combinations in sheep. *Lab Anim Sci*, 39 (6): 591-597, 1989.
- Green SA, Thurmon JC:** Xylazine: a review of its pharmacology and use in veterinary medicine. *J Vet Pharma Therap*, 11, 295-313, 1988.
- Kamiloglu NN, Kamiloglu A, Beytut E, Baran V:** Effect of xylazine on cardiopulmonary activity, antioxidant status and lipid peroxidation in rams. *Indian Vet J*, 82, 1043-1045, 2005.
- Sumitra M, Manikandan P, Rao KV, Nayeem M, Manohar BM, Puvanakrishnan R:** Cardiorespiratory effects of diazepam-ketamine, xylazine-ketamine and thiopentone anesthesia in male Wistar rats-a comparative analysis. *Life Sci*, 75 (15): 1887-1896, 2004.
- Matkovics B, Szabo L, Varga IS:** Determination of enzyme activities in lipid peroxidation and glutathione pathways (in Hungarian). *Lab Diagn*, 15, 248-249, 1988.
- McMurray CH, Blanchflower WJ, Rice DA:** Influence of extraction technique on the determination of α -tocopherol in animal feedstuffs. *J Assoc Off Anal Chem*, 63, 1258-1261, 1980.
- Sedlak J, Lindsay RH:** Estimation of total protein-bound and non-protein sulfhydryl groups in tissue with ellman's reagent. *Anal Biochem*, 25, 192-205, 1968.
- Lawrence RA, Burk RF:** Glutathione-peroxidase activity in selenium-deficient rat liver. *Biochem Biophys Res Commun*, 71, 952-955, 1976.
- Drabkin DL:** Haemoglobin spectrometric studies, part XIV. *J Biol Chem*, 164, 703-708, 1946.
- Traber DL, Wilson RD, Priano LL:** The effect of alpha-adrenergic blockade on the cardiopulmonary response to ketamine. *Anesth Analg*, 50 (5): 737-742, 1971.
- Flecknell PA:** Injectable anaesthetics. In: Hall Taylor, Editor, Bailliere Tindall, WB Saunders, Philadelphia, pp. 129-156, 1994.
- Nowrouzian I, Schels HF, Ghodsian I, Karimi H:** Evaluation of the anaesthetic properties of ketamine and a ketamine/xylazine/atropine combination in sheep. *Vet Rec*, 108 (16): 354-356, 1981.
- Boyd MR:** Biochemical mechanisms in chemical-induced lung injury: Role of metabolic activation. *Crc Crit Rev Toxicol*, 7, 103-176, 1980.
- Berlinskii VV, Berlinskii VF:** Ketamine-clofelin anesthesia in children. *Anesteziol Reanimatol*, 5, 38-40, 1995.
- Reinke LA, Kotake Y, Moore DR, Nanji AA:** Free radical formation during ketamine anesthesia in rats: A cautionary note. *Free Rad Biol Med*, 24 (6): 1002-1006, 1998.
- Celly CS, McDonnell WN, Young SS, Black WD:** The comparative hypoxaemic effect of four alpha 2 adrenoceptor agonists (xylazine, romifidine, detomidine and medetomidine) in sheep. *J Vet Pharmacol Ther*, 20 (6): 464-471, 1997.
- Tranquilli WJ, Maze M:** Clinical pharmacology and use of α_2 -adrenergic agonists in veterinary anaesthesia. *Anaesth Pharma Rev*, 1, 297-309, 1993.
- Ugla A, Lindqvist A:** Acute pulmonary oedema as an adverse reaction to the use of xylazine in sheep. *Vet Rec*, 111, 42, 1983.
- Amouzadeh HP, Sangiah SCW, Qualls JR, Cowell RL, Mauromoustakos A:** Xylazine-induced pulmonary edema in rats. *Toxicol Appl Pharmacol*, 108, 417-427, 1991.
- Helmer KS, Cui Y, Dewan A, Mercer DW:** Ketamine/Xylazine attenuates LPS-induced iNOS expression in various rat tissues. *J Surgical Res*, 112, 70-78, 2003.
- Halliwell B:** Reactive oxygen species and the central nervous system: A Review. *J Neurochem*, 59 (5):1609-1623, 1992.
- Shi X, Castranova V, Halliwell B, Vallyathan V:** Reactive oxygen species and silica-induced carcinogenesis. *J Toxicol Environ Health B Crit Rev*, 1 (3):181-197, 1998.
- Halliwell B, Gutteridge JM:** Free radicals, lipid peroxidation and cell damage. *Lancet*, 2 (8411): 1095, 1984.
- Gutteridge JM, Halliwell B:** Free radicals and antioxidants in year 2000. A historical look to the future. *Ann NY Acad Sci*, 895, 136-147, 2000.
- Meister A, Anderson ME:** Glutathione. *Annu Rev Biochem*, 52, 711-760, 1983.