

The Investigation of Recombinant Human IGF-I (Insulin-Like Growth Factor-I) Injection on Rats Liver Malondealdehyde and Reduced Glutathione Levels

Aysel GÜVEN*

Meryem DEMİRCAN POYRAZ**
Hacı Ahmet DEVECİ*

Alpaslan Kadir DEVRİM***
İnan KAYA*

* Department of Biochemistry, Faculty of Science, University of Kafkas, 36100-Kars, TURKEY

** Department of Histology and Embriology, Faculty of Veterinary Medicine, University of Kafkas, 36040-Kars, TURKEY.

***Department of Biochemistry, Faculty of Veterinary Medicine, University of Kafkas, 36100-Kars, TURKEY

Yayın Kodu: 2006/27-A

Summary

The effect of insulin-like growth factor-I (IGF-I) administration on lipid peroxidation and reduced glutathione in liver of rats was studied. Total of 30 female rats (Wistar albino), weighing 95 ± 5 g, were divided into two groups and kept in two separate cages. The second group (n=15) was rh IGF-I (100 ng/kg/day) in 0.01 M NaHCO₃ was subcutaneously injected for 7 days period. Also, first group (control group (n=15)) was subcutaneously received 0.01 M NaHCO₃ in same manner with rh IGF-I treated group. At the end of injection period, at 8th, 15th and 22nd days, rats have been weighed and liver tissues were removed. At the 7th injection day, removed liver tissues from control group has been examined and has not been seen any significant changes statistically. At the 15th injection day, has seen that MDA levels are increased (P<0.01) and GSH levels are decreased (P<0.01) statistically. On the other hand at the 22nd day has been recorded that both MDA and GSH levels are decreased. These results showed that IGF-I has been important metabolic effect on the MDA and GSH.

Keywords: Insulin like growth factor I, lipid peroxidation, reduced glutathione, rat

Recombinant Human IGF-I (Insülin-Like Growth Factor-I) Enjeksiyonu Yapılan Ratlarda Karaciğer Malondialdehit ve Redükte Glutasyon Düzeylerinin Araştırılması

Özet

Bu çalışmada deney hayvanı olarak, ağırlıkları 95 ± 5 g arasında değişen 30 adet dişi rat (Wistar albino), kontrol ve deney olarak 15'er hayvanlık iki gruba ayrıldı ve ayrı kafeslere alındı. Deney grubundaki ratlara 7 gün boyunca her 24 saatte bir 100 ng/kg/gün dozunda rekombinat human İnsülin-Like Growth Factor-I (rh IGF-I), kontrol grubuna ise sadece 0.01 M NaHCO₃ subkutan olarak enjekte edildi. Yedi günlük enjeksiyonu takip eden 8, 15 ve 22. günlerde ratlar tartılarak karaciğer dokuları alındı.

Enjeksiyonun 7. gününde kontrol grubundan alınan karaciğer örneklerinde malondialdehit (MDA) ve redükte glutasyon (GSH) düzeylerinde istatistiksel olarak önemli bir değişim görülmedi. Onbeşinci günde MDA düzeylerinde istatistiksel olarak önemli bir artış görülürken, GSH düzeyinde aynı oranda bir azalış görüldü. Yirmiikinci günde hem MDA hem de GSH düzeylerinde önemli bir azalma kaydedildi (p<0.01). Bu sonuçlar IGF-I'in MDA ve GSH üzerinde metabolik bir etkiye sahip olduğunu göstermektedir.

Anahtar sözcükler: İnsülin benzeri büyüme faktörü-I, lipid peroksidasyonu, redükte glutasyon, rat.

İletişim (Correspondence)

Phone: +90 474 2120201

e-mail: ayselguven@hotmail.com

INTRODUCTION

Oxidative stress, resulting from the increased production of free radicals and reactive oxygen species and/or a decrease in antioxidant system, causes severe damage to biologic macromolecules and dysregulation of normal metabolism and physiology^{1,2}. Oxygen radicals are known to produce membrane peroxidation and MDA formation³, which are both detrimental to cellular function. Peroxidation can increase membrane permeability, whereas MDA can inactivate membrane transporters³ by forming intra and intermolecular cross links^{3,4}. Such events may be more damaging in the long term⁵. To minimize free radical damage, there is a complex antioxidant system, which includes the interception of free radicals with antioxidants to form less reactive compounds. Antioxidants prevent the organism from the harmful effects of free radicals by scavenging or inhibiting their formation. Cells maintain their vital functions against oxidative damage with the help of a system that involves antioxidant enzymes.

It is generally known that the GSH-related thiols participate in many important biological reactions, including the protection of cell membranes against oxidative damage⁶. The endogenous antioxidant glutathione (GSH) is the most abundant antioxidant in the body and is responsible for neutralizing reactive oxygen species produced in the mitochondria^{7,8}. GSH scavenges super oxide radicals and hydrogen peroxide resulting in the formation of the disulfide glutathione (GSSG). GSSG is subsequently reduced by GSSG reductase and several other antioxidant enzymes are used to process and detoxify reactive oxygen species ROS⁸. However, the effect of IGF-I on the anti-oxidative enzymes, especially in liver tissue, should be clarified.

The insulin-like growth factors (IGF-I and IGF-II) compare a family of polypeptide hormones structurally related to insulin with multifunctional metabolic and anabolic properties⁹. Growth hormone released from the anterior pituitary gland increases IGF-I synthesis and secretion from the liver and, at the cellular level, promotes the synthesis of DNA, RNA and protein. Although these actions are most commonly associated with long bone growth and the increase in lean body mass that occurs during normal mammalian development, studies in our laboratory and others have focused on a possible relationship between oxidative

stress and IGF-I and liver^{10,11}.

Insulin growth factor-I is a growth factor with anabolic effects¹², which is mainly produced by hepatocytes. Patients with advanced liver cirrhosis show low serum concentrations of this hormone¹³. Previous works have shown beneficial effects of low doses of IGF-I on experimental cirrhosis including liver function tests and oxidative liver damage¹⁴. To better understand the possible pathways behind the beneficial effect of IGF-I, the aim of this work is to investigate several parameters involved in oxidative damage in hepatic tissue.

MATERIALS and METHODS

Animals

Wistar albino female rats, weighing 95±5 g, were fasted for 12h, but allowed free access to water before the experiments. The animals were kept in wirebottom cages, in a room at a constant temperature (22±2°C) with 12-h light and dark cycles, and were fed a standart rat chow (Table 1) and tap water ad libitum. The rats were randomly divided into the two groups of fifteen each. The second group was rh IGF-I (100 ng/kg/day) in 0.01 M NaHCO₃ was subcutaneous injected for a 7 days period. Also, control group (n=15) was subcutaneous received 0.01 M NaCHO₃ in the same manner with rh IGF-I treated group.

Table 1. Diyet bileşimi

Table 1. Diet composition

Ingredients	%
Wheat	10
Corn	23
Barley	15
Wheat bran	8
Soybean	26
Fish flour	8
Meat-bone flour	4
Pelleted	5
Salt	0.8
Vitamin mineral mix*	0.2

* Vitamin A, D₃, E, K₃, B₁, B₂, B₆ and B₁₂, nicotinamide, folic acid, biotin, Mn, Fe, Zn, Cu, I, Co, Se, antioxidants (butyldroxytoluol) and Ca.

Biochemical Analysis

Analysis of malondialdehyde in the liver

Lipid peroxidation in the liver tissue was assessed by estimation of malondialdehyde (MDA) according to the method of Ohkawa et al¹⁵. Reduced glutathione (GSH) was measured estimated in the liver tissue. GSH was assayed by the method of Beutler et al¹⁶. GSH concentrations were calculated for 1 g protein content from the 13000 g supernatant fraction and protein concentration, in supernatants were determined by Folin-phenol reagent with bovine serum albumin as the standard¹⁷.

Statistical Analysis

The collected values as mean±SE were statistically analyzed (student's t-test) (IFFC, 1987) and evaluated using the SPSS 6.0 (1993) softwove programe. P values< 0.01, 0.05 were considered significant¹⁸.

RESULTS

Lipid peroxidation is thought to be an important mechanism of liver injury and malondialdehyde (MDA) is one of its end products. Thus, measurement of MDA can be used to assess lipid peroxidation.

DISCUSSION

Antioxidants prevent new radical species formation by converting existing free radical species into less harmful molecules or by preventing transformation of free radicals from other molecules. To avoid from ROS-induced injury to tissue, a complex antioxidant system, consisting of both enzymatic and non enzymatic defenses, has evaluated. Traditionally, antioxidants have been defined as substances that prevent the formation of ROS or other oxidants, scavenge them, or repair the damage that they cause¹⁹⁻²². There is a considerable evidence that IGF-I decreases ROS and free radicals production both in vivo and invitro, and depressed IGF-I synthesis in rat is considered to be responsible for enhanced oxidative stress^{14,23}. Liver damage has been associated with oxidative stress and lipid peroxidation^{16,24}. Recent studies from laboratory in rats with carbon tetrachloride-induced cirrhosis have demonstrated that short courses of treatment with low doses of IGF-I are associated to hepatoprotective^{11,23,24}.

In order to give a better insight into the pathways by which IGF-I seems to exert its the hepatoprotective actions, this study was aimed to analyze several parameters involved in oxidative stress in the liver,

Table 2. rh-human IGF-I verilen ratlarda karaciğer MDA ve GSH düzeyleri
Table 2. Liver MDA and GSH levels in rats after rh-human IGF-I administration

Time after administration (day)	n	Control		Treatment	
		MDA (nmol/g prot)	GSH (nmol/g prot)	MDA (nmol/g prot)	GSH (nmol/g prot)
7	5	794.9±32.2	3.32±0.3	789.4±32.2	3.27±0.3
15	5	792.3±28.4	3.25±0.2	804.7±26.8*	2.56±0.4*
22	5	794.8±27.4	3.33±0.3	776.8±32.6*	2.94±0.3*

* p<0.01

Table 2 shows the biochemical values, GSH and MDA levels of IGF-I-treatment animals and normal control subjects. GSH and MDA levels in liver did not differ significantly in the liver tissues of IGF-I-treated groups (7th day), compared to those of controls group. On the 15th day it has been seen that MDA level has been increased statistically on the other hand, GSH level has been decreased at the same ratio with MDA. On the 22th day it has been seen that both MDA and GSH levels have decreased.

such as MDA, GSH both in IGF-I treated and untreated rats.

In order to find a relationship between the studied parameters and oxidative liver damage, MDA levels, an index of lipid peroxidation, were evaluated. Hepatic levels of MDA were increased in untreated rats compared with control group as it was previously reported in cirrhotic rat similar protocols¹⁰⁻¹¹. The same result is shown in our study in Table 2. However, the reduction in the antioxidant defense systems

recovered to control levels at 22nd day, the evaluated MDA and GSH decreased with the recovery of radical scavenging enzymes ($p < 0.01$) compared to those of control group. GSH is known to be counter-regulatory to a number of cytokines including IGF-I, which is being investigated as a possible surrogate marker for breast cancer risk. GSH conjugation of IGF-I is a major pathway of excretion of IGF-I²⁵.

These data indicate that the antioxidant defense system against oxidative damage was impaired after IGF-I-induced. In addition, the recovery of this system in the liver is mainly caused by the degradation of lipid peroxide, resulting in a decrease in MDA. Those findings support the hypothesis that IGF-I enhances the activity of the pathway and impairs the antioxidant status, particularly glutathione redox cycle, resulting in a higher defense against oxidative stress^{10,24}.

REFERENCES

- 1 **Knight JA:** Free radicals: Their history and current status in aging and disease. *Ann Clin Lab Sci*, 28, 331-46, 1998.
- 2 **Toyokuni S:** Reactive oxygen species-induced molecular damage and its application in pathology. *Pathol Int*, 49, 91-102, 1999.
- 3 **Kramer JH, Mak T, Webllicki WB:** Differential sensitivity of canine cardiac sarcolemmal and microsomal enzymes to inhibition by free radical induced lipid peroxidation. *Circ Res*, 55, 120-124, 1985.
- 4 **Rajguru SU, Yeargans GS, Seidler NW:** Exercise causes oxidative damage to skeletal muscle microsomes while increasing cellular subhydrials. *Life Sci*, 54, 149-157, 1993.
- 5 **Nair V, Turner GA, Offerman R:** Novel adducts from the modification of nucleic acid bases by malondialdehyde. *J Am Chem Soc*, 106, 3370-3371, 1984.
- 6 **Bacon BR, Britton RS:** Hepatic injury in chronic iron overload. Role lipid peroxidation. *Chem Biol Interactions*, 70, 183-226, 1989.
- 7 **Bush AI:** Metals and neuroscience. *Curr Opin Chem Biol*, 4, 184-191, 2000.
- 8 **Lash LH, Putt DA:** Renal cellular of exogeneous glutathione: Heterogeneity at physiological and pharmacological concentrations. *Biochem Pharm*, 58, 897-907, 1999.
- 9 **Sara VR, Hall K:** Insulin-like growth factor-I and their binding protein. *Physiol Rew*, 70, 591-613, 1990.
- 10 **Muguerza B, Castilla-Cortazar I, Garcia M, Quiroga J, Santidrian S, Prieto J:** Antifibrogenic effect in vivo of low doses of insulin-like growth factor-I in cirrhotic rats. *Biochem Biophys Acta*, 1536, 185-195, 2001.
- 11 **García-Fernández M, Castilla-Cortázar I, Díaz-Sánchez M, Navarro I, Puche JE, Castilla A, Casares AD, Clavijo E, González-Barón S:** Antioxidant effects of insulin-like growth factor-I (IGF-I) in rats with advanced liver cirrhosis. *BMC Gastroenterol*, 5, 7-13, 2005.
- 12 **Jones JI, Clemmons DR:** Insulin growth factors and their binding proteins: biological actions. *Endocr Rev*, 16, 3-34, 1995.
- 13 **Caufriez A, Reding P, Urbain D, Goldstein J, Copinschi G:** Insuline like growth factor-I: A good indicator of functional hepatocellular capacity in alcoholic liver cirrhosis. *J Endocrinol Invest*, 14, 317-321, 1991.
- 14 **Picardi A, Costa De Oliveira A, Muguerza B, Tosar A, Quiroga J, Castilla-Cortáza I, Santidrián S, Prieto J:** Low doses of insulin-like growth factor-I improve nitrogen retention and food efficiency in rats with early cirrhosis. *J Hepatol*, 24, 267-279, 1997.
- 15 **Ohkawa H, Ohishi N, Yagi KL:** Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Biochem*, 95, 351-358, 1979.
- 16 **Beutler E, Duron O, Kelly BM:** Improved method for the determination of blood glutathione. *J Lab Clin Med*, 61, 882-888, 1963.
- 17 **Lowry OH, Rosenbrough NJ, Far AL, Randal RJ:** Protein measurement with the folin-phenol reagent. *J Biol Chem*, 193-265, 1951.
- 18 **International Federation of Clinical Chemistry (IFCC):** *Eur J Clin Chem Biochem*, 25, 337-342, 1987.
- 19 **Sun Y:** Free radicals, antioxidant enzymes and carcinogenesis. *Free Rad Biol Med*, 8, 583-589, 1990.
- 20 **Meister ME:** Glutathione. *Ann Rev Biochem*, 52, 711-760, 1983.
- 21 **Güven A, Kaya N:** Determination of reduced glutathione, glutathione S-transferase and selenium levels in goose liver cells with damage induced by carbon tetrachloride and ethanol. *Tr J Vet Anim Sci*, 29, 1233-8, 2005.
- 22 **Güven A, Güven A, Gülmez M:** The effect of kefir on the activities of GSH-Px, GST, CAT, GSH and LPO levels in carbon tetrachloride-induced mice tissues. *J Vet Med B*, 50, 412-416, 2003.
- 23 **Castilla-Cortázar I, Aliaga-Montilla MA, Salvador J, Garcia M, Delgado G, Gonzalez-Baron S, Quiroga J, Prieto J:** Insuline-like growth factor-I restores the reduced somatostatinergic tone controlling growth hormone secretion in cirrhotic rats. *Liver*, 21, 405-409, 2001.
- 24 **Castilla-Cortázar I, Garcia M, Muguerza B, Perez R, Quiroga J, Santidrián S, Prieto J:** Hepatoprotective effects of insulin-like growth factor-I in rats with carbon-tetrachloride-induced cirrhosis. *Gastroenterology*, 113, 1682-1691, 1997.
- 25 **Daughaday WH:** A personal history of the origin of the somatomedin hypothesis and recent challenges to its validity. *Perspect Biol Med*, 32, 194-211, 1989.