REVIEW ARTICLE

Saponins and Their Role as Vaccine Adjuvant Against Coccidiosis in **Poultry**

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How to cite this article?

Alsayeqh AF: Saponins and their role as vaccine adjuvant against coccidiosis in poultry. Kafkas Univ Vet Fak Derg, 31 (3): 351-359, 2025.

DOI: 10.9775/kvfd.2025.34196

Article ID: KVFD-2025-34196 Received: 10.04.2025 Accepted: 09.06.2025 Published Online: 13.06.2025

Abstract

Coccidiosis, induced by various Eimeria species, has been one of the most important health threats and performance of poultry around the world. Other than the current treatment, successful vaccination approaches have been realized. Advances in saponin biochemistry, from Quillaja saponaria and Yucca schidigera plants, have supported the development of vaccine adjuvants based on these natural glycosides. Saponins also activate innate immune pathways and can assist with antigen presentation, enhancing humoral and cell-mediated responses. Saponin-based adjuvants, including QS-21 and Quil A, can enhance adjuvant efficacy by inducing higher antibody responses and promoting long-lasting protective immunity privation. Nonetheless, challenges including toxicity issues about the saponin fractions and the variable adjuvant activity among different saponins, have also been reported. Future studies aim to improve saponin adjuvant formulations, determine their harmlessness, and investigate new transport systems, such as immunostimulating complexes. By enhancing poultry health, such advancements contribute to safer meat and egg products, directly supporting food safety. Moreover, reducing disease-related losses in poultry farms promotes food security by ensuring stable and efficient protein production. The focus of this review article is to highlight the role of saponins as vaccine adjuvants to enhance immunity against Eimeria species in poultry.

Keywords: Coccidiosis, Eimeria, Food safety, Phytochemicals, Quil A, Saponins, Vaccine adjuvants

Introduction

Coccidiosis is one of the most important health problems [1] affecting animals worldwide, causing disease with major economic losses [2]. It is caused by obligate intracellular parasitic protozoa known as Eimeria, belonging to the order Apicomplexa [3]. Eimeria remains one of the most economically important species that cause disease in livestock globally [4]. Eimeria is remarkably receptive in the poultry industry [5], this is because the transmission of parasites is highly favored by the bulk number of susceptible birds [6,7]. Eimeria spp. results in coccidiosis disease that hinders the expansion of the poultry industry [8]. The infection may get worse because of poor management techniques such as excessive stocking densities, contaminated feeders, drinkers, and damp litter that encourage oocyst sporulation, and inadequate ventilation facilities [9]. Among the most virulent Eimeria spp., E. tenella is the most common, followed by E. acervulina and E. maxima [10]. Due to increased mortality, stunted growth, and a low feed conversion ratio, the

disease directly affects the production potential of affected livestock, resulting in significant financial losses [11,12]. In the poultry industry, globally there was a huge economic loss of more than 3 billion US\$ annually [13]. Such economic losses not only reduce profitability but also threaten food security by limiting the availability of affordable protein sources. Additionally, disease-related contamination in meat and eggs poses a serious risk to food safety across the supply chain.

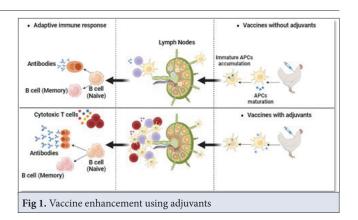
The coccidial parasites can enter various animal hosts and effectually exploit the immune system [4]. This creates a serious dispute against the action of control. Many drugs have been developed against the coccidia parasite [8]. In avian coccidiosis, Eimeria species develop frequent resistance when new and effective drugs are detected in the body [14]. Alongside commercial losses, using chemotherapeutics in poultry can also generate harmful residues in eggs and meat $^{[15]}$. According to studies, the use of vaccines is the most efficient way to prevent and reduce the prevalence of infectious diseases [16,17]. Though



there is an emerging line of action as prophylactic vaccines, these are said to be a successful approach against coccidia, but these vaccines are minimal and in short supply [4]. Anticoccidial drugs have successfully prevented coccidiosis over the decades, with certain restrictions related to production costs [18]. There are various kinds of vaccines: live attenuated, inactivated vaccines (killed), and recombinant vaccines (subunits) [19]. Mostly, the commercial vaccines against coccidiosis are based on killed or live virulent organisms [20]. Nonetheless, the assurance of a live attenuated vaccine is uncertain because of the possibility of severe regression [21]. When comparing, it shows that a durable solution and safer options are provided by subunit vaccines [22,23]. However, isolated antigens from different host systems can be less antigenic and immune stimulating than killed or live attenuated vaccine variants [24]. To stimulate an effective immune response by delivering these antigens to the immune system is a challenging task [25]. In this matter, it is generally acknowledged that to increase efficacy and immunity, we need further components to be added to vaccines. These components are macromolecules and their complexes, either compounds or molecules, commonly known as adjuvants [26,27].

Adjuvants were first discovered in 1920 by Gaston Ramon [28], a French scientist who observed that if aluminum salts are included in a vaccine, they increase its potential [29]. It is derived from the Latin word "Adjuvare" which means to aid [30,31]. Vaccine adjuvants are used to increase the potential of the immune system response to combined antigens [32]. Most adjuvants are chemicals, macromolecules, or compounds that improve innate immunity by combining with the antigens, and enhance the immune response [16,33]. Many efforts have been made to reduce the intricacy of antigens, such as pure antigens, recombinants [34], artificially manufactured peptides, and proteins, as an alternative use of whole inactive organisms without adjuvants to induce immunity [35,36]. The non-toxic adjuvants can boost and direct immune response. Certain adjuvants, including mineral gels or water in oil emulsions, regulate the antigen at the injection site [37]. Adjuvants increase the secondary type of immune response by slowly releasing the restrained antigen into the immune system [38, 39]. The enhancement in the action of vaccines using adjuvants is shown in *Fig. 1*.

Although the mechanism of action of adjuvants is still under investigation, significant progress has been made in recent years to identify them [40]. The potential and virulent nature of adjuvants should be maintained in order to provide protected stimulants with very fewer reactions [41], it depends on how adjuvants are being used [36,42]. In several years, adjuvants have been used in many experimental subunit vaccines that are often too weak to stimulate



immune response alone [31], however, not all vaccines need adjuvants [36].

Many studies have shown that the plant-derived compounds of herb spices known as phytochemicals, play a vital role in antimicrobial, coccidiostat activities in animals [43-46], as they improve gut health, immunity, growth enhancement, and adsorption of nutrients [47,48]. Adjuvants derived from phytochemistry, such as saponins, carbohydrates, protein lectins, and heatshock proteins, are efficient immune stimulants with little toxicity [49]. Different studies marked the effectiveness of phytochemicals as adjuvants against various diseases. These have proved to be useful as potential adjuvants against coccidiosis. Saponins are the most significant phyto-biotics for use as adjuvants in vaccines among all other phytochemicals [50]. They are natural compounds produced by plants that play an important role in defense mechanisms due to their antimicrobial, fungicidal, and insecticidal properties. Furthermore, many plant saponins can activate the immune system, which leads to considerable interest in their potential as vaccine adjuvants [3]. This review deliberates the potential application of plantderived saponin compounds in the development of vaccines against poultry coccidiosis. It also comprehensively explains the immunological effects, functions, constraints, and postulated mechanisms of action of saponins.

SAPONINS

Saponins are amphiphilic, heat-stable, and glycosidic secondary metabolites derived from plants ^[51]. These steroidal aglycones and triterpenoids are significantly used in the pharmaceutical industry ^[52]. They have immunomodulatory and antioxidant qualities that mainly make them useful as immunizing adjuvants against coccidiosis ^[53,54]. The bark, roots, leaves, and seeds of the *Quillaia Saponaria* tree yield an extract known to have immune-modulatory qualities. This tree is widely distributed throughout South America ^[3,52,55]. Non-polar

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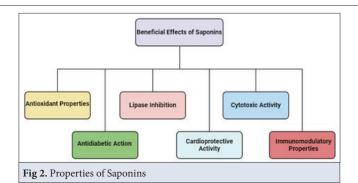


Table 1. Adjuvant active saponins Source Adjuvant **Poultry** Eimeria spp. Antigen Route Results Ref. Weight gain, antibody [63] Saponins QCDC/RT Broiler E. acervulina rProfilin Subcutaneous production, and intestinal lesions were reduced Mitogen-induced lymphocyte production and increased Chicken rProfilin body weight Saponins OCDC-R E. acervulina Subcutaneous Reduced intestinal lesions. No impact on oocyst shedding. Purified saponin from Quillaja Intramuscular and E. spp. [65] QS-21 QS-21 Chicken IgG2a saponica, used to stabilize lipid Intranasal emulsions Chicken Increased growth. Reduced Saponins QCDC E. maxima rProfilin Non-encapsulated Embryo oocyst shedding Anemone 3- monodesmoside, The serum antibody titer in Chicken [55, 67] IgGs Subcutaneous radiant E. spp. 3,28-bisdesmoside chicken activates macrophages Saponins No effect on oocyst, IgG [68] Saponins QCDC Broiler E. acervulina rProfilin Subcutaneous increased ISCOMs from E. tenella all Increased IgGs protected Broiler Intranacal Saponins E. tenella endemic plants antigens against infection Antigens from Decreased lesion score and [70] **ISCOMs** Broiler E. tenella Intranasal Saponins oocyst shedding sporozoite Increased total body weight **ISCOMs** Broiler [71] Saponins E. tenella AgP27 Diet Supplement Conferred partial protection against infection

aglycones make up the chemical structure of saponins, which is connected with chains of carbohydrates (polar) ^[56,57]. The presence of both polar and non-polar groups provides surface-active properties ^[58]. Saponins have a lot of beneficial effects as shown in *Fig. 2*.

They can stimulate a broad range of cytokine secretions ^[59], thereby enhancing both humoral and cellular immune responses ^[60]. Their ability to modulate innate immunity suggests that they could be used to design new vaccines that induce specific immune responses tailored to various pathogens and can reduce the drug resistance problem ^[61,62]. Adjuvant active saponins are shown in *Table 1*.

SAPONIN-DERIVED COMPOUNDS

Summarized in Table 2.

Immunostimulating Complexes (ISCOMs)

One innovative approach in utilizing saponins while mitigating their toxicity involves formulating them into Immunostimulating complexes "ISCOMs" ^[72]. ISCOMs are particulate adjuvant systems that combine saponins with cholesterol and phospholipids ^[73], significantly reducing toxicity while retaining potent adjuvant properties ^[74]. They provide a sophisticated approach to coccidiosis vaccination in poultry, taking advantage of their unique structure and composition to efficiently boost immune responses ^[3]. It mimics the immune system's innate detection of bacterial and viral structures to trigger potent humoral and cellular immune responses in poultry ^[75]. It is essential to activate both immune systems while treating intracellular parasites such as *Eimeria* species ^[76].

Table 2. Saponin-Derived Compounds								
Source	Common Name	Adjuvant	Mode of Action	Benefits	Ref.			
Peptasan	Sikakai	Acacia concinna	Immune modulation enhances adaptive and innate immune reactions Antioxidant and anti-inflammatory characteristics	Enhanced immune response Reduced parasite load Natural and safe for poultry	[91]			
Yucca schidigera	Yucca	Y. schidigera	Saponins boost the immune response by boosting cytokine synthesis and T-cell proliferation Antioxidant characteristics	Improved gut health Reduced ammonia levels Enhanced vaccine efficacy	[92]			
Quillaja Saponaria	Soapbark	Q. Saponaria	Saponins boost the immune response by boosting dendritic cell development and antigen presentation Adjuvant for vaccinations	Increased vaccine efficacy Enhanced antibody production Natural adjuvant	[50]			
Norponin XO2	Yucca and Fenugreek	Yucca schidigera and Trigonella foenum- graecum	Yucca and Fenugreek saponins work together to enhance immunological response Improved intestinal health and decreased parasite load	The synergistic effect enhances immune response Improved gut health	[93]			

Research has demonstrated encouraging outcomes in terms of improving vaccination efficacy and offering a more comprehensive defense against many *Eimeria* strains that are common in the production of chickens [45,77,78].

Quil A

Dalsgaard purified an adjuvant derived from Q. saponaria called Quil A containing a mixture of saponins in 1978 [79]. The mixture is enriched to use as an adjuvant for immunity induction via other isotypes of antibodies [80]. It is also used commercially in many vaccines [81,82]. The toxicity differs significantly between the components of Quil A [83,84]. The efficacy of Quil A as an adjuvant has been hindered by its apparent toxicity [85], which has been found in small animals following parenteral immunization with this adjuvant, and such toxicity may limit its utility [86]. However, nontoxic immunostimulatory fractions of Quil A have been identified, which may reduce or eliminate this issue [87]. While Quil A does not appear to be an effective mucosal adjuvant, its use as part of ISCOMs appears to be critical to the system's efficacy [88]. Structural and functional relationships of Saponin-adjuvant compounds are of interest because various fractions are used in immunological research.

QS-21

It can play an important role in poultry coccidiosis by improving antigen presentation and helping in T cells and antibody development ^[75]. Regardless of its efficacy, it is linked to formulation stability issues and some toxicity problems ^[89], which has prompted ongoing research to address these issues. The goal of continuing research is to create QS-21 variations that are less toxic while maintaining immunostimulatory activity to maximize its

potential as an adjuvant in vaccines against coccidiosis [90]. QS-21 shows promising results as an adjuvant in boosting immune responses against coccidiosis. However, their efficacy and safety profiles in this setting necessitate additional research and optimization for their efficient application in poultry vaccination programs [44].

MECHANISM OF ACTION

Immunomodulatory Effects and Immune Boosting

In poultry, saponins enhance the immune response by affecting immune organ maturation, increasing antibody levels, and providing better defense against coccidiosis [94]. They are well-recognized for their potent immune modulatory effects [95]. The primary mechanism involves their interaction with immune cells, leading to the enhancement of the immune response [96]. They attract a variety of cells including macrophages, DCs (dendritic cells), and lymphocytes [55]. Saponins activate DCs by enhancing their capacity to process T lymphocytes. Maturation of DCs and upregulation of co-stimulatory chemicals allow for the efficient activation of T-cells [73,95]. B cells can create large amounts of antibodies. Saponins do this by boosting antibody isotype switching, resulting in a more effective immune response to pathogens.

Antioxidant Properties

Saponins have a great antioxidant capacity, which helps to manage reactive oxygen species and minimize diseases linked with oxidative stress, a major role in coccidiosis [97,98]. Reactive oxygen species (ROS) are produced during coccidia infection, resulting in oxidative stress and tissue damage. Saponins' antioxidant activities serve to neutralize ROS, lowering oxidative stress and its related

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diseases [99]. This antioxidant activity not only protects the host from ROS-induced damage but also improves the immune response by lowering pro-inflammatory cytokine production and increasing overall antioxidant levels [100-102].

Mechanisms at the Cellular Level

Saponins regulate several essential systems that underpin the immune response. These include the effects on antigen presentation and cytokine generation [103]. Saponins have a crucial role in antigen presentation, which initiates the adaptive immune response [104]. When antigen-presenting cells (APCs) are enhanced, dendritic cells (DCs) and macrophages can efficiently process and present antigens, thereby initiating immune responses [78,105]. It involves the upregulation of major histocompatibility complex (MHC) molecules on the surface of APCs, that are important for presenting antigens to T-cells. Boosting antigen presentation by APCs leads to activation of helper T cells (Th cells) and cytotoxic T cells [106].

Comparative Studies with Other Adjuvants

Comparative studies help to position saponins within the broader context of coccidiosis control strategies and highlight their advantages and limitations. A comparative study by [107] assessed the efficacy of saponins compared to synthetic anticoccidial drugs in broiler chickens. The results demonstrated that saponins were as effective as synthetic drugs in controlling coccidiosis, with the added benefit of being natural and reducing the risk of resistance development. This study also emphasized the potential of saponins to be used in combination with other control methods to enhance their efficacy [107]. In another study, saponins were compared with ionophore anticoccidials, which are widely used in the poultry industry. The findings indicated that saponins were comparable to ionophores in reducing lesion scores and oocyst shedding. Moreover, saponins exhibited a more favorable safety profile, with fewer side effects reported in treated birds. This highlights the potential of saponins as safer alternatives to traditional anticoccidials [78,105].

LIMITATIONS OF PHYTOCHEMICAL ADJUVANT VACCINATION OF COCCIDIOSIS

Phytochemicals may have anti-nutritional qualities, resulting in decreased feed intake, growth suppression, and negative effects on body growth [108]. Some may be harmful at high quantities but harmless at lower levels [109]. The effectiveness of phytochemicals in treating coccidiosis varies depending on the specific plant compounds employed, their quantities, and their ratios [110-112]. The

lack of standardization has the potential to undermine the dependability and efficacy of phytochemical adjuvants ^[113]. The extensive use of phytochemicals, like anticoccidial chemical compounds, has the potential to cause resistance in *Eimeria* spp. ^[108]. As a result, it is critical to apply techniques to prevent resistance development, such as alternating phytochemical adjuvants with other management methods and employing them in conjunction with other interventions ^[114,115]. More research is needed to understand the mechanisms of action of phytochemical adjuvants and to identify the most effective chemicals for coccidiosis reduction.

Prospects

Advances in biotechnology and synthetic biology offer exciting opportunities for producing saponins with enhanced properties [112], genetic engineering techniques can be used to modify microorganisms to produce saponins with specific structural features that enhance their stability and immunostimulatory effects. Moreover, the integration of saponins with advanced delivery systems, such as nanocarriers and immunostimulating complexes, holds great potential. These delivery systems can protect saponins from degradation, enhance their targeting of immune cells, and provide a controlled release of antigens [115]. The task at hand involves developing standardized animal models that replicate the illnesses of the intended species for use in vaccination regimens and determining the parameters that must be assessed accurately to determine the effectiveness of vaccines [3].

Conclusion

In summary, the reviewed data suggest that saponins as adjuvants allow the opportunity to achieve the main objective of adjuvant research in vaccines, as a safe option that is mostly non-toxic and able to boost immune response as it may be included in many vaccine formulations against coccidiosis. However, the difficulty lies in choosing the best adjuvant that is most suitable for immunization protocols. Despite that, their decreased immunogenicity eventually became a crucial component as a refined antigen of several vaccinations. In addition to their amazing qualities, the non-toxic nature of most saponins eases the main worry about manufactured compounds having severe effects. Saponins can increase the efficacy of coccidiosis vaccines, which are made using live attenuated, inactivated, and recombinant techniques by increasing immune response. Formulations such as Quil A, QS-21, and ISCOMs have shown considerable promise in increasing vaccine efficacy while minimizing toxicity when appropriately modified. Unlike synthetic adjuvants, saponins are derived from plants and often exhibit fewer side effects, with added antioxidant and immunomodulatory benefits. However, comparative studies suggest that saponins can be as effective as conventional anticoccidials, while also reducing the risk of drug resistance. It is demonstrated that they can dramatically boost the immune response, increasing protection against illness and saponins are safer substitutes for traditional adjuvants. All things considered, the use of saponins in vaccines is an essential tactic in the prevention and control of coccidiosis, and future research should concentrate on the creation of safer and more effective adjuvants and vaccines to tackle this serious health issue.

DECLARATION

Availability of Data and Materials: Data and materials for this research are available upon request.

Acknowledgments: The researcher would like to thank the Deanship of Graduate Studies and Scientific Research at Qassim University.

Conflict of Interest: The author declares that there is no conflict of interest.

Generative Artificial Intelligence: No Generative Artificial Intelligence was used in this research

REFERENCES

- 1. Khan MM, Lillehoj HS, Lee Y, Adetunji AO, Omaliko PC, Kang HW, Fasina YO: Use of selected plant extracts in controlling and neutralizing toxins and sporozoites associated with necrotic enteritis and coccidiosis. *Appl Sci*, 14 (8):3178, 2024. DOI: 10.3390/app14083178
- 2. Shahininejad H, Rahimi S, Torshizi MAK, Arabkhazaeli F, Ayyari M, Behnamifar A, Abuali M, Grimes J: Comparing the effect of phytobiotic, coccidiostat, toltrazuril, and vaccine on the prevention and treatment of coccidiosis in broilers. *Poult Sci*, 103 (5):103596, 2024. DOI: 10.1016/j. psj.2024.103596
- **3. Sander VA, Corigliano MG, Clemente M:** Promising plant-derived adjuvants in the development of coccidial vaccines. *Front Vet Sci*, 6:20, 2019. DOI: 10.3389/fvets.2019.00020
- **4. Gao Y, Sun P, Hu D, Tang X, Zhang S, Shi F, Yan X, Yan W, Shi T, Wang S:** Advancements in understanding chicken coccidiosis: From *Eimeria* biology to innovative control strategies. *One Health Adv*, 2 (1): 1-19, 2024. DOI: 10.1186/s44280-024-00039-x
- **5. Nasiri V, Jameie F, Morovati Khamsi H:** Detection, identification, and characterization of *Eimeria* spp. from commercial chicken farms in different parts of Iran by morphometrical and molecular techniques. *Acta Parasitol*, 69 (1): 854-864, 2024. DOI: 10.1007/s11686-024-00818-x
- **6. Blake DP, Tomley FM:** Securing poultry production from the everpresent *Eimeria* challenge. *Trends Parasitol*, 30 (1): 12-19, 2014. DOI: 10.1016/j.pt.2013.10.003
- **7. Saeed Z, Alkheraije KA:** Botanicals: A promising approach for controlling cecal coccidiosis in poultry. *Front Vet Sci*, 10:1157633, 2023. DOI: 10.3389/fvets.2023.1157633
- **8. Johnson WT:** Avian coccidiosis. *Poult Sci*, 2 (5): 146-163, 1923. DOI: 10.3382/ps.0020146
- 9. Nowaczewski S, Janiszewski S, Kaczmarek S, Kaczor N, Racewicz P, Jarosz Ł, Ciszewski A, Ślósarz P, Hejdysz M: Evaluation of the effectiveness of alternative methods for controlling coccidiosis in broiler chickens: A field trial. *Anim Sci Pap Rep*, 41 (2): 97-110, 2023. DOI: 10.2478/aspr-2023-0001
- 10. Anwar F, Mm D, Khan MS, Noreen S, Malik M, Nouroz F, Anwar MZ, Khan MSZ: Exploring the diversity of *Eimeria* species and prevalence of clinical coccidiosis in suspected broiler chickens. *Res Sq.* 2024 (preprint article). DOI: 10.21203/rs.3.rs-3400201/v1
- 11. Ahad S, Tanveer S, Nawchoo IA, Malik TA: Coccidiosis in poultry A

- review. Life Sci J, 20 (7): 44-50, 2023. DOI: 10.7537/marslsj200723.06
- **12. Imran A, Alsayeqh A:** Anticoccidial efficacy of *Citrus sinensis* essential oil in broiler chicken. *Pak Vet J*, 42 (4): 461-466, 2022. DOI: 10.29261/pakvetj/2022.082
- **13. Borgonovo F, Ferrante V, Grilli G, Guarino M:** An innovative approach for analysing and evaluating enteric diseases in poultry farm. *Acta IMEKO*, 13 (1): 1-5, 2024. DOI: 10.21014/actaimeko.v13i1.1627
- **14. Hou Y, Han B, Lin Z, Liu Q, Liu Z, Si H, Hu D:** Effects of six natural compounds and their derivatives on the control of coccidiosis in chickens. *Microorganisms*, 12 (3):601, 2024. DOI: 10.3390/microorganisms12030601
- 15. Hussain K, Alsayeqh AF, Abbas A, Abbas RZ, Rehman A, Zaib W, Rehman TU, Mahmood MS: Potential of *Glycyrrhiza glabra* (Licorice) extract an alternative biochemical and therapeutic agent against coccidiosis in broiler chickens. *Kafkas Univ Vet Fak Derg*, 28 (5), 2022. DOI: 10.9775/kyfd.2022.27620
- **16.** Nooraei S, Sarkar Lotfabadi A, Akbarzadehmoallemkolaei M, Rezaei N: Immunogenicity of different types of adjuvants and nano-adjuvants in veterinary vaccines: A comprehensive review. *Vaccines*, 11 (2):453, 2023. DOI: 10.3390/vaccines11020453
- 17. Liao S, Lin X, Zhou Q, Wang Z, Yan Z, Wang D, Su G, Li J, Lv M, Hu J: Epidemiological investigation of coccidiosis and associated risk factors in broiler chickens immunized with live anticoccidial vaccines in China. *Front Vet Sci*, 11:1375026, 2024. DOI: 10.3389/fvets.2024.1375026
- 18. Rahmani A, Ahmed Laloui H, Kara R, Dems MA, Cherb N, Klikha A, Blake DP: The financial cost of coccidiosis in Algerian chicken production: A major challenge for the poultry sector. *Avian Pathol*, 53 (5): 368–379, 2024. DOI: 10.1080/03079457.2024.2336091
- **19. Saeed Z, Alsayeqh A:** Evaluation of anthelmintic, hematological and serum biochemical effects of herbal dewormer on the cattle. *Solven Vet Res*, 60, 353-362, 2023. DOI: 10.26873/svr-1624-2022
- **20.** Hauck R, Macklin KS: Vaccination against poultry parasites. *Avian Dis*, 67 (4): 441-449, 2024. DOI: 10.1637/aviandiseases-d-23-99989
- **21. Vashishtha VM, Kumar P:** The durability of vaccine-induced protection: An overview. *Expert Rev Vaccines*, 23 (1): 389-408, 2024. DOI: 10.1080/14760584.2024.2331065
- **22.** Innes EA, Bartley PM, Rocchi M, Benavidas-Silvan J, Burrells A, Hotchkiss E, Chianini F, Canton G, Katzer F: Developing vaccines to control protozoan parasites in ruminants: Dead or alive? *Vet Parasitol*, 180 (1-2): 155-163, 2011. DOI: 10.1016/j.vetpar.2011.05.036
- 23. Chavda VP, Ghali ENHK, Balar PC, Chauhan SC, Tiwari N, Shukla S, Athalye M, Patravale V, Apostolopoulos V, Yallapu MM: Protein subunit vaccines: Promising frontiers against COVID-19. *J Control Release*, 366, 761-782, 2024. DOI: 10.1016/j.jconrel.2024.01.017
- **24. Rautenschlein S, Schat KA:** The immunological basis for vaccination. *Avian Dis*, 67 (4): 366-379, 2024. DOI: 10.1637/aviandiseases-d-23-99996
- **25.** Zheng L, Zhang L, Tan F, Zhang H, Wang L, Zheng M: *Lactococcus lactis* NZ3900/pNZ8149-IL-4-IL-2 as an adjuvant to reduce vaccine dose in chicken coccidia live mixed vaccine. *Animal Res One Health*, 2 (1): 50-58, 2024. DOI: 10.1002/aro2.12
- **26. Petrovsky N, Aguilar JC:** Vaccine adjuvants: Current state and future trends. *Immunol Cell Biol*, 82 (5): 488-496, 2004. DOI: 10.1111/j.0818-9641.2004.01272.x
- **27. Oladejo M, Tijani AO, Puri A, Chablani L:** Adjuvants in cutaneous vaccination: A comprehensive analysis. *J Control Release*, 369, 475-492, 2024. DOI: 10.1016/j.jconrel.2024.03.045
- **28. Alsayeqh AF, Rao ZA:** Nutritional supplements for the control of avian coccidiosis A review. *Ann Anim Sci*, 23 (4): 993-1007, 2023. DOI: 10.2478/aoas-2023-0013
- **29.** Chippaux JP: Gaston Ramon's big four. *Toxins*, 16 (1): 33, 2024. DOI: 10.3390/toxins16010033
- **30. Mustafa S, Alsayeqh AF:** Role of plant phytochemicals/extracts for the control of *Dermanyssus gallinae* in poultry and its zoonotic importance. *Poult Sci*, 104 (4):104899, 2025. DOI: 10.1016/j.psj.2025.104899
- 31. Goetz M, Thotathil N, Zhao Z, Mitragotri S: Vaccine adjuvants for infectious disease in the clinic. *Bioeng Transl Med*, 2024 (9):e10663, 2024.

- DOI: 10.1002/btm2.10663
- **32.** Verma SK, Mahajan P, Singh NK, Gupta A, Aggarwal R, Rappuoli R, Johri AK: New-age vaccine adjuvants, their development, and future perspective. *Front Immunol*, 14:1043109, 2023. DOI: 10.3389/fimmu.2023.1043109
- **33.** Chen X: Emerging adjuvants for intradermal vaccination. *Int J Pharm*, 632:122559, 2023. DOI: 10.1016/j.ijpharm.2022.122559
- **34. Gupta S, Pellett S:** Recent developments in vaccine design: From live vaccines to recombinant toxin vaccines. *Toxins*, 15 (9):563, 2023. DOI: 10.3390/toxins15090563
- **35. Min W, Kim WH, Lillehoj EP, Lillehoj HS:** Recent progress in host immunity to avian coccidiosis: IL-17 family cytokines as sentinels of the intestinal mucosa. *Dev Comp Immunol*, 41 (3): 418-428, 2013. DOI: 10.1016/j.dci.2013.04.003
- **36.** Facciolà A, Visalli G, Laganà A, Di Pietro A: An overview of vaccine adjuvants: Current evidence and future perspectives. *Vaccines*, 10 (5):819, 2022. DOI: 10.3390/vaccines10050819
- **37. Fan J, Jin S, Gilmartin L, Toth I, Hussein WM, Stephenson RJ:** Advances in infectious disease vaccine adjuvants. *Vaccines*, 10 (7):1120, 2022. DOI: 10.3390/vaccines10071120
- **38. Qiao N, Liu Q, Meng H, Zhao D:** Haemolytic activity and adjuvant effect of soyasaponins and some of their derivatives on the immune responses to ovalbumin in mice. *Int Immunopharmacol*, 18 (2): 333-339, 2014. DOI: 10.1016/j.intimp.2013.12.017
- **39. Sobral MC, Cabizzosu L, Kang SJ, Feng Z, Ijaz H, Mooney DJ:** Modulating adjuvant release kinetics from scaffold vaccines to tune adaptive immune responses. *Adv Healthc Mater,* 14 (5):2304574, 2024. DOI: 10.1002/adhm.202304574
- **40. Ren H, Jia W, Xie Y, Yu M, Chen Y:** Adjuvant physiochemistry and advanced nanotechnology for vaccine development. *Chem Soc Rev*, 52 (15): 5172-5254, 2023. DOI: 10.1039/d2cs00848c
- 41. Schijns V, Fernández-Tejada A, Barjaktarović Ž, Bouzalas I, Brimnes J, Chernysh S, Gizurarson S, Gursel I, Jakopin Ž, Lawrenz M, Nativi C, Paul S, Pedersen GK, Rosano C, Ruiz-de-Angulo A, Slütter B, Thakur A, Christensen D, Lavelle EC: Modulation of immune responses using adjuvants to facilitate therapeutic vaccination. *Immunol Rev*, 296 (1): 169-190, 2020. DOI: 10.1111/imr.12889
- **42. Francis MJ:** Recent advances in vaccine technologies. *Vet Clin North Am Small Anim Pract*, 48 (2): 231-241, 2018. DOI: 10.1016/j.cvsm.2017.10.002
- **43. Abbas RZ, Qureshi MA, Saeed Z:** Botanical compounds: A promising control strategy against *Trypanosoma cruzi. Bol Latinoam Caribe Plantas Med Aromat*, 24 (3): 308-327, 2025.
- **44. 1. Allenspach K:** Clinical immunology and immunopathology of the canine and feline intestine. *Vet Clin North Am Small Anim Pract*, 41 (2): 345-360, 2011. DOI: 10.1016/j.cvsm.2011.01.004
- **45. Felici M, Tugnoli B, Piva A, Grilli E**: *In vitro* assessment of anticoccidials: Methods and molecules. *Animals*, 11 (7):1962, 2021. DOI: 10.3390/ani11071962
- **46. Baz MM, Alfagham AT, Al-Shuraym LA, Moharam AF:** Efficacy and comparative toxicity of phytochemical compounds extracted from aromatic perennial trees and herbs against vector borne *Culex pipiens* (Diptera: Culicidae) and *Hyalomma dromedarii* (Acari: Ixodidae) as green insecticides. *Pak Vet J*, 44 (1): 55-62, 2024. DOI: 10.29261/pakvetj/2024.144
- **47. Zeng Z, Zhang S, Wang H, Piao X:** Essential oil and aromatic plants as feed additives in non-ruminant nutrition: A review. *J Anim Sci Biotechnol*, 6, 1-10, 2015. DOI: 10.1186/s40104-015-0004-5
- **48.** Ahmad R, Yu YH, Hua KF, Chen WJ, Zaborski D, Dybus A, Hsiao FSH, Cheng YH: Management and control of coccidiosis in poultry A review. *Anim Biosci*, 37 (1):1, 2024. DOI: 10.5713/ab.23.0189
- **49.** Choudhary S, Khan S, Rustagi S, Rajpal VR, Khan NS, Kumar N, Thomas G, Pandey A, Hamurcu M, Gezgin S, Zargar S, Khan MK: Immunomodulatory effect of phytoactive compounds on human health: A narrative review integrated with bioinformatics approach. *Curr Top Med Chem*, 24 (12): 1075-1100, 2024. DOI: 10.2174/0115680266274272240321 065039

- **50.** Fleck JD, Betti AH, Da Silva FP, Troian EA, Olivaro C, Ferreira F, Verza SG: Saponins from *Quillaja saponaria* and *Quillaja brasiliensis*: Particular chemical characteristics and biological activities. *Molecules*, 24 (1):171, 2019. DOI: 10.3390/molecules24010171
- **51.** Jolly A, Kim H, Moon JY, Mohan A, Lee YC: Exploring the imminent trends of saponins in personal care product development: A review. *Ind Crops Prod*, 205:117489, 2023. DOI: 10.1016/j.indcrop.2023.117489
- 52. Nguyen LT, FĂRcaŞ AC, Socaci SA, TofanĂ M, Diaconeasa ZM, Pop OL, SalanȚĂ LC: An overview of saponins A bioactive group. *Bull Univ Agric Sci Vet Med Cluj-Napoca Food Sci Technol*, 77 (1): 25-36, 2020. DOI: 10.15835/buasvmcn-fst:2019.0036
- **53.** Siddiqui M, Shah N, Dur-Re-Shahwar M, Ali SY, Muzammil A, Fatima N: The phytochemical analysis of some medicinal plants. *Liaquat Med Res J*, 3 (1): 1-7, 2021. DOI: 10.38106/lmri.2021.36
- **54.** Kumar A, P N, Kumar M, Jose A, Tomer V, Oz E, Proestos C, Zeng M, Elobeid T, Sneha K, Oz F: Major phytochemicals: Recent advances in health benefits and extraction method. *Molecules*, 28 (2):887, 2023. DOI: 10.3390/molecules 28020887
- **55.** Wang P: Natural and synthetic saponins as vaccine adjuvants. *Vaccines*, 9 (3):222, 2021. DOI: 10.3390/vaccines9030222
- **56. Hailat AM, Abdelqader AM, Gharaibeh MH:** Efficacy of phyto-genic products to control field coccidiosis in broiler chickens. *Int J Vet Sci*, 13 (3): 266-272, 2023. DOI: 10.47278/journal.ijvs/2023.099
- 57. Afzal MU, Pervaiz M, Ejaz A, Bajwa E, Naz S, Saeed Z, Ullah S, Gillani SS, Kan RRM, Younas U: A comprehensive study of the sources, extraction methods and structures of the saponin compounds for its antidiabetic activity. *Biocatal Agric Biotechnol*, 54:102913, 2023. DOI: 10.1016/j. bcab.2023.102913
- **58.** Zaynab M, Sharif Y, Abbas S, Afzal MZ, Qasim M, Khalofah A, Ansari MJ, Khan KA, Tao L, Li S: Saponin toxicity as key player in plant defense against pathogens. *Toxicon*, 193, 21-27, 2021. DOI: 10.1016/j. toxicon.2021.01.009
- 59. Ma Y, Zhao Y, Luo M, Jiang Q, Liu S, Jia Q, Bai Z, Wu F, Xie J: Advancements and challenges in pharmacokinetic and pharmacodynamic research on the traditional Chinese medicine saponins: A comprehensive review. *Front Pharmacol*, 15:1393409, 2024. DOI: 10.3389/fphar. 2024.1393409
- **60. Mieres-Castro D, Mora-Poblete F:** Saponins: Research progress and their potential role in the post-COVID-19 pandemic era. *Pharmaceutics*, 15 (2):348, 2023. DOI: 10.3390/pharmaceutics15020348
- **61.** Hayajneh FMF, Abdelqader A, Zakaria H, Abuajamieh M, Araj SA: Drug resistance and coccidiosis affects on immunity, performance, blood micronutrients, and Intestinal Integrity in broiler chickens. *Int J Vet Sci*, 13 (1): 34-41, 2024. DOI: 10.47278/journal.ijvs/2023.054
- **62.** Luo X, Song Z, Zeng X, Ye Y, Zheng H, Cai D, Yuan Q, Li H, Tong Y, Lu D: A promising self-nanoemulsifying adjuvant with plant-derived saponin D boosts immune response and exerts an anti-tumor effect. *Front Immunol*, 14:1154836, 2023. DOI: 10.3389/fimmu.2023.1154836
- **63.** Lee SH, Lillehoj HS, Jang SI, Lee KW, Kim DK, Lillehoj EP, Yancey RJ, Dominowski PJ: Evaluation of novel adjuvant *Eimeria* profilin complex on intestinal host immune responses against live *E. acervulina* challenge infection. *Avian Dis*, 56 (2): 402-405, 2012. DOI: 10.1637/9906-082411-resnote.1
- **64.** Kim DK, Lillehoj HS, Lee SH, Dominowski P, Yancey RJ, Lillehoj EP: Effects of novel vaccine/adjuvant complexes on the protective immunity against *Eimeria acervulina* and transcriptome profiles. *Avian Dis*, 56 (1): 97-109, 2012. DOI: 10.1637/9720-031711-reg.1
- **65.** Wilson-Welder JH, Torres MP, Kipper MJ, Mallapragada SK, Wannemuehler MJ, Narasimhan B: Vaccine adjuvants: Current challenges and future approaches. *J Pharm Sci*, 98 (4): 1278-1316, 2009. DOI: 10.1002/jps.21523
- **66.** Lee S-H, Lillehoj HS, Jang SI, Hong Y-H, Min W, Lillehoj EP, Yancey RJ, Dominowski P: Embryo vaccination of chickens using a novel adjuvant formulation stimulates protective immunity against *Eimeria maxima* infection. *Vaccine*, 28 (49): 7774-7778, 2010. DOI: 10.1016/j. vaccine.2010.09.051
- 67. Mad T, Sterk H, Mittelbach M, Rechberger GN: Tandem mass

- spectrometric analysis of a complex triterpene saponin mixture of *Chenopodium quinoa. J Am Soc Mass Spectrom*, 17, 795-806, 2006. DOI: 10.1016/j.jasms.2006.02.013
- **68.** Lee S-H, Lillehoj HS, Jang SI, Lee K-W, Yancey RJ, Dominowski P: The effects of a novel adjuvant complex/*Eimeria* profilin vaccine on the intestinal host immune response against live *E. acervulina* challenge infection. *Vaccine*, 28 (39): 6498-6504, 2010. DOI: 10.1016/j.vaccine.2010.06.116
- 69. Berezin VE, Bogoyavlenskyi AP, Khudiakova SS, Alexuk PG, Omirtaeva ES, Zaitceva IA, Tustikbaeva GB, Barfield RC, Fetterer RH: Immunostimulatory complexes containing *Eimeria tenella* antigens and low toxicity plant saponins induce antibody response and provide protection from challenge in broiler chickens. *Vet Parasitol*, 167 (1): 28-35, 2010. DOI: 10.1016/j.vetpar.2009.09.045
- 70. Garcia JL, da Silva Guimarães Jr J, Headley SA, Bogado ALG, Bugni FM, Ramalho DC, de Souza LM: *Eimeria tenella*: Utilization of a nasal vaccine with sporozoite antigens incorporated into Iscom as protection for broiler breeders against a homologous challenge. *Exp Parasitol*, 120 (2): 185-190, 2008. DOI: 10.1016/j.exppara.2008.07.007
- **71. Guo FC, Kwakkel RP, Williams BA, Suo X, Li WK, Verstegen MWA:** Coccidiosis immunization: Effects of mushroom and herb polysaccharides on immune responses of chickens infected with *Eimeria tenella. Avian Dis*, 49 (1): 70-73, 2005. DOI: 10.1637/7227-062504r1
- 72. Kadiyska T, Tourtourikov I, Dabchev K, Zlatarova A, Stoynev N, Hadjiolova R, Spandidos DA, Adamaki M, Zoumpourlis V: Herbs and plants in immunomodulation. *Int J Funct Nutr*, 4 (1): 1-11, 2023. DOI: 10.3892/iifn.2023.31
- 73. Chen K, Wang N, Zhang X, Wang M, Liu Y, Shi Y: Potentials of saponins-based adjuvants for nasal vaccines. *Front Immunol*, 14:1153042, 2023. DOI: 10.3389/fimmu.2023.1153042
- 74. Stertman L, Palm A-KE, Zarnegar B, Carow B, Lunderius Andersson C, Magnusson SE, Carnrot C, Shinde V, Smith G, Glenn G, Fries L, Lövgren Bengtsson K: The Matrix-M[™] adjuvant: A critical component of vaccines for the 21st century. *Hum Vaccin Immunother*, 19 (1):2189885, 2023. DOI: 10.1080/21645515.2023.2189885
- **75.** Eze CO, Berebon DP, Evurani SA, Asilebo IC, Gugu TH: Vaccine delivery using nanoparticles: A critical look at ISCOMs 4 decades but 2 and still counting. *Trop J Nat Prod Res*, 6 (5):680, 2022.
- **76.** Britez JD, Rodriguez AE, Di Ciaccio L, Marugán-Hernandez V, Tomazic ML: What do we know about surface proteins of chicken parasites *Eimeria? Life*, 13 (6):1295, 2023. DOI: 10.3390/life13061295
- **77. Soutter F, Werling D, Tomley FM, Blake DP:** Poultry coccidiosis: Design and interpretation of vaccine studies. *Front Vet Sci*, 7:101, 2020. DOI: 10.3389/fvets.2020.00101
- **78. Abd El-Ghany WA:** Intervention strategies for controlling poultry coccidiosis: Current knowledge. *J Worlds Poult Res*, 11 (4): 487-505, 2021. DOI: 10.36380/jwpr.2021.58
- **79. Dalsgaard K:** A study of the isolation and characterization of the saponin quil A. Evaluation of its adjuvant activity, with a special reference to the application in the vaccination of cattle against foot-and-mouth disease. *Acta Vet Scand.* **69**, 7-40, 1978.
- **80.** Joshi SS, Dice L, Ailavadi S, D'Souza DH: Antiviral effects of *Quillaja saponaria* extracts against human noroviral surrogates. *Food Environ Virol*, 15 (2): 167-175, 2023. DOI: 10.1007/s12560-023-09550-7
- 81. Lupi GA, Valtierra FXS, Cabrera G, Spinelli R, Siano ÁS, González V, Osuna A, Oresti GM, Marcipar I: Development of low-cost cage-like particles to formulate veterinary vaccines. *Vet Immunol Immunopathol*, 251:110460, 2022. DOI: 10.1016/j.vetimm.2022.110460
- 82. Hamerski L, Carbonezi CA, Cavalheiro AJ, Bolzani VdS, Young MCM: Saponinas triterpênicas de *Tocoyena brasiliensis* Mart. (Rubiaceae). *Quim Nova*, 28 (4): 601-604, 2005. DOI: 10.1590/s0100-40422005000400009
- **83.** Carnet F, Perrin-Cocon L, Paillot R, Lotteau V, Pronost S, Vidalain PO: An inventory of adjuvants used for vaccination in horses: The past, the present and the future. *Vet Res*, 54 (1): 18, 2023. DOI: 10.1186/s13567-023-01151-3
- 84. Facciolà A, Visalli G, Laganà A, Di Pietro A: An overview of vaccine adjuvants: Current evidence and future perspectives. *Vaccines*, 10 (5):819,

- 2022. DOI: 10.3390/vaccines10050819
- 85. Moni SS, Abdelwahab SI, Jabeen A, Elmobark ME, Aqaili D, Ghoal G, Oraibi B, Farasani AM, Jerah AA, Alnajai MMA: Advancements in vaccine adjuvants: The journey from alum to nano formulations. *Vaccines*, 11 (11):1704, 2023. DOI: 10.3390/vaccines11111704
- **86.** Rathogwa NM, Scott KA, Opperman P, Theron J, Maree FF: Efficacy of SAT2 foot-and-mouth disease vaccines formulated with Montanide ISA 206B and quil-A saponin adjuvants. *Vaccines*, 9 (9):996, 2021. DOI: 10.3390/vaccines9090996
- **87.** Zhou S, Song Y, Nilam A, Luo Y, Huang W-C, Long MD, Lovell JF: The predominant *Quillaja Saponaria* fraction, QS-18, is safe and effective when formulated in a liposomal murine cancer peptide vaccine. *J Control Release*, 369, 687-695, 2024. DOI: 10.1016/j.jconrel.2024.04.002
- **88.** Correa VA, Portilho AI, De Gaspari E: Vaccines, adjuvants and key factors for mucosal immune response. *Immunology*, 167 (2): 124-138, 2022. DOI: 10.1111/imm.13526
- **89.** Morein B, Hu K-F, Abusugra I: Current status and potential application of ISCOMs in veterinary medicine. *Adv Drug Deliv Rev*, 56 (10): 1367-1382, 2004. DOI: 10.1016/j.addr.2004.02.004
- **90. Hook S, Rades** T: Immune stimulating complexes (ISCOMs) and quil-A containing particulate formulations as vaccine delivery systems. **In,** Immunomic Discovery of Adjuvants and Candidate Subunit Vaccines. 233-261. Springer, New York, 2012.
- 91. Sánchez-Hernández C, Castañeda-Gómez del Campo JA, Trejo-Castro L, Mendoza-Martínez GD, Gloria-Trujillo A: Evaluation of a feed plant additive for coocidiosis control in broilers herbals for coccidiosis control. *Braz J Poult Sci*, 21:eRBCA-2019, 2019. DOI: 10.1590/1806-9061-2018-0846
- **92. Su JL, Shi BL, Zhang PF, Sun DS, Li TY, Yan SM:** Effects of yucca extract on feed efficiency, immune and antioxidative functions in broilers. *Braz Arch Biol Technol*, 59:e16150035, 2016. DOI: 10.1590/1678-4324-2016150035
- **93. Bafundo KW, Johnson AB, Mathis GF:** The effects of a combination of *Quillaja saponaria* and *Yucca schidigera* on *Eimeria* spp. in broiler chickens. *Avian Dis*, 64 (3): 300-304, 2020. DOI: 10.1637/aviandiseases-d-20-00016
- 94. El-Shall NA, Abd El-Hack ME, Albaqami NM, Khafaga AF, Taha AE, Swelum AA, El-Saadony MT, Salem HM, El-Tahan AM, AbuQamar SF, El-Tarabily KA, Elbestawy AR: Phytochemical control of poultry coccidiosis: A review. *Poult Sci*, 101 (1): 101542-101542, 2022. DOI: 10.1016/j.psj.2021.101542
- **95.** Chung ELT, Alghirani MM, Kassim NA, Ong YL, Jesse FFA, Sazili AQ. Loh TC: Impact of *Brachiaria decumbens* leaf meal supplementation on broiler chickens raised in tropical environments in terms of growth performance, blood biochemistry, and stress biomarkers. *Bra J Poult Sci*, 26 (1): 1-8, 2024. DOI: 10.1590/1806-9061-2023-1878
- 96. Behl T, Kumar K, Brisc C, Rus M, Nistor-Cseppento DC, Bustea C, Aron RAC, Pantis C, Zengin G, Sehgal A, Kaur R, Kumar A, Arora S, Setia D, Chandel D, Bungau S: Exploring the multifocal role of phytochemicals as immunomodulators. *Biomed Pharmacother*, 133:110959, 2021. DOI: 10.1016/j.biopha.2020.110959
- 97. Pop LM, Varga E, Coroian M, Nedişan ME, Mircean V, Dumitrache MO, Farczádi L, Fülöp I, Croitoru MD, Fazakas M, Györke M: Efficacy of a commercial herbal formula in chicken experimental coccidiosis. *Parasit Vectors*, 12, 1-9, 2019. DOI: 10.1186/s13071-019-3595-4
- **98.** Chen Y, Liu L, Yu L, Li S, Zhu N, You J: Curcumin supplementation improves growth performance and anticoccidial index by improving the antioxidant capacity, inhibiting inflammatory responses, and maintaining intestinal barrier function in *Eimeria tenella*-infected broilers. *Animals*, 14 (8):1223, 2024. DOI: 10.3390/ani14081223
- **99. Ahmad Bhat B, Aadil S:** Adjuvants used in animal vaccines-their formulations and modes of action: An overview. *Osmaniye Korkut Ata Üniv Fen Bil Derg,* **4** (3): 492-506, 2021. DOI: 10.47495/okufbed.852809
- **100.** Reyes C, Patarroyo MA: Adjuvants approved for human use: What do we know and what do we need to know for designing good adjuvants? *Eur J Pharmacol*, 945:175632, 2023. DOI: 10.1016/j.ejphar.2023.175632
- 101. Ahmad S, Humak F, Ahmad M, Altaf H, Qamar W, Hussain A,

- **Ashraf U, Abbas RZ, Siddique A, Ashraf T:** Phytochemicals as alternative anthelmintics against poultry parasites: A review. *Agrobiol Rec*, 12, 34-45, 2023. DOI: 10.47278/journal.abr/2023.015
- 102. Basiouni S, Tellez-Isaias G, Latorre JD, Graham BD, Petrone-Garcia VM, El-Seedi HR, Yalçın S, El-Wahab AA, Visscher C, May-Simera HL, Huber C, Eisenreich W, Shehata AA: Anti-inflammatory and antioxidative phytogenic substances against secret killers in poultry: Current status and prospects. *Vet Sci*, 10 (1):55, 2023. DOI: 10.3390/vetsci10010055
- **103.** Jiang L, Zhang G, Li Y, Shi G, Li M: Potential application of plant-based functional foods in the development of immune boosters. *Front Pharmacol*, 12:637782, 2021. DOI: 10.3389/fphar.2021.637782
- **104.** Díaz-Dinamarca DA, Salazar ML, Castillo BN, Manubens A, Vasquez AE, Salazar F, Becker MI: Protein-based adjuvants for vaccines as immunomodulators of the innate and adaptive immune response: Current knowledge, challenges, and future opportunities. *Pharmaceutics*, 14 (8):1671, 2022. DOI: 10.3390/pharmaceutics14081671
- 105. Batool S, Munir F, Sindhu ZD, Abbas RZ, Aslam B, Khan MK, Imran M, Aslam MA, Ahmad M, Chaudhary MK: *In vitro* anthelmintic activity of *azadirachta indica* (neem) and *Melia azedarach* (BAKAIN) essential oils and their silver nanoparticles against haemonchus contortus. *Agrobiol Rec*, 11, 6-12, 2023. DOI: 10.47278/journal.abr/2023.002
- **106.** Manohar MM, Campbell BE, Walduck AK, Moore RJ: Enhancement of live vaccines by co-delivery of immune modulating proteins. *Vaccine*, 40 (40): 5769-5780, 2022. DOI: 10.1016/j.vaccine.2022.08.059
- **107.** Duffy CF, Mathis GF, Power RF: Effects of Natustat[™] supplementation on performance, feed efficiency and intestinal lesion scores in broiler chickens challenged with *Eimeria acervulina*, *Eimeria maxima* and *Eimeria tenella*. *Vet Parasitol*, 130 (3-4): 185-190, 2005. DOI: 10.1016/j. vetpar.2005.03.041

- **108. Broom LJ:** Evidence-based consideration of dietary 'alternatives' to anticoccidial drugs to help control poultry coccidial infections. *Worlds Poult Sci J,* 77 (1): 43-54, 2021. DOI: 10.1080/00439339.2021.1873713
- **109.** Rashid S, Ashraf FH, Shoukat A, Nawaz A, Hassan K: Phytomedicine efficacy and prospects in poultry: A new insight to old anthelmintic resistance. *Continent Vet J*, 4 (1): 62-75, 2024. DOI: 10.71081/cvj/2024.009
- 110. Arif M, Baty RS, Althubaiti EH, Ijaz MT, Fayyaz M, Shafi ME, Albaqami NM, Alagawany M, Abd El-Hack ME, Taha AE, Salem HM, El-Tahan AM, Elnesr MM: The impact of betaine supplementation in quail diet on growth performance, blood chemistry, and carcass traits. *Saudi J Biol Sci*, 29 (3): 1604-1610, 2022. DOI: 10.1016/j.sjbs.2021.11.002
- **111. Gakuubi MM, Wanzala W:** A survey of plants and plant products traditionally used in livestock health management in Buuri district, Meru County, Kenya. *J Ethnobiol Ethnomed*, 8, 1-20, 2012. DOI: 10.1186/1746-4269-8-39
- 112. Jambwa P, Katsande S, Matope G, McGaw LJ: Ethnoveterinary remedies used in avian complementary medicine in selected communal areas in Zimbabwe. *Planta Med*, 88 (03/04): 313-323, 2022. DOI: 10.1055/a-1529-8618
- **113. Singh VK, Arora D, Ansari MI, Sharma PK:** Phytochemicals based chemopreventive and chemotherapeutic strategies and modern technologies to overcome limitations for better clinical applications. *Phytother Res,* 33 (12): 3064-3089, 2019. DOI: 10.1002/ptr.6508
- **114. Abad MREH, Ghaniei A:** Effects of herbal medicine in the treatment of poultry coccidiosis. *J Worlds Poult Sci*, 2 (1): 1-7, 2023. DOI: 10.58803/jwps.v2i1.10
- **115. Saleem M, Rahman HU, Abbas J:** Rapid recovery of *Salmonella* from chicken meat and poultry fecal samples by selective pre-enrichment. *Continent Vet J*, 3 (1): 49-53, 2023. DOI: 10.71081/cvj/2023.007