

## REVIEW ARTICLE

## Epidemiology and Public Health Importance of Bovine Salmonellosis

Abdulaziz M. ALMUZAINI <sup>1</sup>(\*)  Ahmed I. ALAJAJI <sup>1</sup> <sup>1</sup>Department of Veterinary Preventive Medicine, College of Veterinary Medicine, Qassim University, 51452 Buraydah, SAUDI ARABIA

## (\*) Corresponding author:

Abdulaziz M. ALMUZAINI

Phone: +966-553360084

E-mail: ammzieny@qu.edu.sa

How to cite this article?

Almuzaini AM, Alajaji AI: Epidemiology and public health importance of bovine salmonellosis. *Kafkas Univ Vet Fak Derg*, 31 (3): 341-350, 2025.

DOI: 10.9775/kvfd.2025.34126

Article ID: KVFD-2025-34126

Received: 19.03.2025

Accepted: 25.04.2025

Published Online: 28.04.2025

## Abstract

Bovine salmonellosis caused by *Salmonella enterica* subsp. *enterica* serovar Dublin (S. Dublin) is a significant public health and economic concern globally. It leads to severe health issues in cattle, including enteritis, septicaemia, and abortion, with high mortality rates, especially in newborn calves. The disease not only impacts the well-being of the animals but also results in substantial economic losses through treatment costs, reduced milk production, and potential outbreaks. The transmission of S. Dublin is primarily through contaminated food, water, and environmental exposure, with the faecal route being the most significant mode of spread. The pathogenesis of S. Dublin involves complex interactions between the bacteria and the host immune system, with the bacteria capable of persisting in the herd as carriers, further complicating control measures. Effective control strategies are critical to minimizing its spread, and understanding the epidemiology, clinical signs, and diagnostic methods is key. This review demonstrates the public health importance, clinical manifestations, economic importance and diagnostic techniques of bovine salmonellosis.

**Keywords:** Bovine, Epidemiology, Food, Public health, Salmonellosis, S. Dublin

## INTRODUCTION

Salmonellosis is a serious public health concern which affects multiple animal species and humans <sup>[1,2]</sup>. It is a foodborne bacterial infection that is caused by *Salmonella enterica* subsp. *enterica* serovar Dublin (S. Dublin) in bovines <sup>[3,4]</sup>. S. Dublin is a Gram-negative, non-spore-forming, oxidase-negative, motile and rod-shaped bacteria that belongs to family *Enterobacteriaceae* <sup>[5,6]</sup>. The survivability of S. Dublin depends on the environmental conditions, temperature, pH and other microflora <sup>[7,8]</sup>. It can survive for years in dried faecal matter and for months in different organic matters such as soil, cattle manure, and slurry <sup>[9,10]</sup>. However, S. Dublin does not resist antibiotics, sunlight, and disinfectants <sup>[11,12]</sup>. Multidrug-resistant strains can be isolated from dairy and beef sources <sup>[13,14]</sup>. The bacteria have the ability to reproduce in moist and warm conditions outside the host cell <sup>[15,16]</sup>. Salmonellosis cause severe health problems in bovines, which include 2 major syndromes: enteritis (inflammation of small intestine) and septicaemia (blood poisoning) <sup>[17,18]</sup>. However, other clinical signs include pyrexia, dysentery, and abortions in pregnant animals <sup>[19,20]</sup>. The severity of clinical signs of Salmonellosis depends on the age

of the animal, infection dose, immune response, and physiological state of the host <sup>[1,17,21]</sup>. In newborn calves, septicaemia with enteritis is most commonly seen <sup>[18,22]</sup>. However, pneumonia and neurological signs may also occur <sup>[23,24]</sup>. In case of young ones or animals with age more than one-week, acute enteritis mostly occurs without systemic involvement <sup>[25,26]</sup>. The infection starts with pyrexia (40.5-41.5°C [105-107°F]) followed by dysentery and sometimes with tenesmus <sup>[27,28]</sup>. Mortality rates in both newborn calves and young ones may increase to 100%, depending on the virulence and infection load <sup>[29,30]</sup>. Milk production may drop in lactating animals <sup>[13,31]</sup>. Salmonellosis is a serious concern for economic losses and public health that needs critical attention <sup>[32-34]</sup>. However, different alternative therapeutics, mainly plant-based compounds, are under study to control multiple diseases <sup>[35,36]</sup>. To minimize the salmonellosis around the globe, we must understand the pathogenesis and epidemiology as drug resistance is increasing dramatically <sup>[37-39]</sup>.

S. Dublin most commonly causes infection after direct transmission through contaminated food, water, and environment <sup>[40,41]</sup>. However, the severity of the disease depends upon the infectious dose of the pathogen <sup>[15,42]</sup>. *Salmonella* bacteria colonize the gut of the host, followed



by invasion in columnar enterocytes through the lymphatic system <sup>[43-45]</sup>. *Salmonella* can also enter the macrophages, which is a critical barrier <sup>[46,47]</sup>. The bacteria replicate in the macrophages, and during the replication they can easily enter the blood, lymph, lungs, liver, spleen, tonsils and lymph nodes <sup>[48,49]</sup>. *S. Dublin* can become the latent carrier, which leads to the persistence of the infection in the herd <sup>[50-52]</sup>. However, *S. Dublin* has been found in the internal organs of the animals while they show no signs of the infection <sup>[53-55]</sup>. Different stages of the *S. Dublin* infection have been reported, including peracute, acute, chronic, passive carrier, active carrier and latent carrier stage <sup>[56-58]</sup>. The infected animal may or may not shed the bacteria in these different stages <sup>[59]</sup>. However, the *S. Dublin* may shed through urine, saliva, milk, faeces and vaginal discharge <sup>[60,61]</sup>. The duration and amount of bacterial excretion vary greatly among infected animals <sup>[62,63]</sup>. This is because the faecal material of the animal contains the highest number of bacteria, and they are produced in the large quantities <sup>[64,65]</sup>. So, the faecal route of the transmission is the most important route to cause the infection <sup>[66,67]</sup>. Humoral and cellular components of the immune system work together to fight pathogenic bacteria <sup>[68,69]</sup>. The first line of defence against the *S. Dublin* consists of neutrophils, polymorphonuclear leukocytes, macrophages, natural killer cells and their secreted cytokines <sup>[70,71]</sup>. This non-specific immune system activates the adaptive immune response <sup>[72,73]</sup>. IgG and IgM titres begin to increase, and IgG attains maximum titre between 6-11 weeks after the inoculation <sup>[74,75]</sup>. However, *S. Dublin* is host adopted to cattle, but there is a lack of agreement on the mechanism of the host adoption <sup>[76]</sup>.

To prevent the *S. Dublin* infection, the mechanism of host adoption is not much important <sup>[77,78]</sup>. However, this feasibility of the host interaction with *S. Dublin* initiates the effective control programs without involving the other livestock sectors <sup>[4,79]</sup>. These effective control programs aid in preventing bovine salmonellosis and other important foodborne and public health important diseases from spreading <sup>[80,81]</sup>. This review emphasises the pathogenesis, public health importance and epidemiology of *S. Dublin* to adopt better preventive measures and control strategies. We will briefly discuss the economic importance, clinical signs, diagnosis, pathogenesis and public health aspect of bovine salmonellosis of *S. Dublin*.

## ECONOMIC IMPORTANCE OF BOVINE SALMONELLOSIS

The high cost of treating clinical salmonellosis in farm animals leads to significant economic losses <sup>[82,83]</sup>. This includes the cost of diagnosis, treatments, cleaning and

disinfectants, laboratory tests, cost of prevention and control, and death of the infected animal <sup>[84,85]</sup>. However, other related economic losses include a drop in the milk production in the lactating animals, poor growth and pregnancy loss in some severe cases of salmonellosis <sup>[86]</sup>. If one animal is diagnosed positive in a large herd of the animals, it would be difficult to diagnose all animals if they are infected with *S. Dublin* or not <sup>[87,88]</sup>. This will increase the cost of diagnostic tools and prevention strategies. The annual estimated loss due to bovine salmonellosis in the United States is billions of dollars, millions of pounds in United Kingdom and \$160 million in Canada <sup>[89]</sup>. However, in North America, economic loss due to 5 outbreaks was \$36.400-\$62 million <sup>[90]</sup>. It is strictly suggested that every £1 spent on the investigation and control strategies can save £5 <sup>[91,92]</sup>.

## CLINICAL MANIFESTATION OF BOVINE SALMONELLOSIS

*S. Dublin* is usually endemic in bovine herds and bovines are the most important carrier of *Salmonella* infection <sup>[90,93]</sup>. They can carry *S. Dublin* for a longer period or sometimes it may be for a lifetime <sup>[94,95]</sup>. In calves, signs and symptoms of clinical ailment are present at the age of 2-6 weeks <sup>[96]</sup>. Signs and symptoms may vary with the infectious dose of the pathogen <sup>[97]</sup>. However, enteric form of salmonellosis is present in young calves which is characterized by dullness, pyrexia, and anorexia that is followed by severe diarrhoea <sup>[98]</sup>. Blood can also be present in the faecal material and faeces may become stringy because of the presence of necrotic mucosa of intestine <sup>[99]</sup>. While in adult animals, subacute or acute salmonellosis mostly occurs with abortion in the pregnant animals during the early stage of acute enteric disease <sup>[100]</sup>. Animals with severe infection show signs of pyrexia, depression and anorexia <sup>[101]</sup>. However, other important clinical manifestations include drop in the milk production, fowl smelling diarrhoea, bloody and mucoid faeces, with shreds of necrotic mucosa of intestine, dehydration, and congestion of mucous membrane <sup>[102]</sup>. Another most common clinical manifestation of bovine salmonellosis is retained placenta, which occurs in approximately 70% of the cases <sup>[103]</sup>. The acute phase of the bovine salmonellosis may only last for 1 week. However, the postmortem findings may vary <sup>[104,105]</sup>.

The animal died in per-acute stage of the infection may have no gross lesions in the postmortem findings <sup>[106]</sup>. But extensive petechial haemorrhages are usually present in subserosal and submucosal layer <sup>[107]</sup>. In young calves, the mesenteric lymph nodes are enlarged in size, congested and oedematous <sup>[108]</sup>. The small intestine shows muco-haemorrhagic or diffused mucoid enteritis <sup>[109]</sup>. Necrotic enteritis, particularly affecting the ileum and large

intestine, occurs in adults <sup>[110]</sup>. Spleen and mesenteric lymph nodes are enlarged and swollen <sup>[111]</sup>. However, the wall of the intestine becomes thickened and covered grey-yellow necrotic material overlying granular, red surface <sup>[108]</sup>.

## INFECTIOUS STAGES AND PATHOGENESIS OF *S. DUBLIN*

There are multiple infection stages of *S. Dublin* that include per-acute, acute, chronic, passive carrier, active carrier and latent carrier <sup>[112,113]</sup>. In per-acute stage of the infection, animals die within a very short period of time usually in 1-2 days <sup>[114]</sup>. Death of animals occurs even before they start to shed the bacteria <sup>[115]</sup>. While the acute phase may remain from 1 to 2 weeks to 5-9 weeks <sup>[116]</sup>. In this stage, animals shed bacteria in a large amount continuously or intermittently through faeces, urine, milk and vaginal discharge <sup>[101]</sup>. The amount of the bacteria in the acute stage of the infection varies from 1 to  $10^8$  CFU/g <sup>[51]</sup>. The chronic infection may remain for months, and the infected animal may or may not shed bacteria <sup>[117]</sup>. However, other carrier stages may remain from weeks to years and infected ones may or may not shed bacteria <sup>[118]</sup>. When the animals shed bacteria in the carrier stage, it may be in a continuous pattern or intermittent <sup>[119,120]</sup>. The amount of the bacteria may be low ( $10^1$ - $10^4$  CFU/g faeces), moderate ( $10^4$ - $10^5$  CFU/g faeces) and high ( $>10^5$  CFU/g faeces). These amounts depend on the infection dose and pathogenesis of *S. Dublin* <sup>[121]</sup>.

*S. Dublin* most commonly enters by ingestion of contaminated feed or water in cattle <sup>[122]</sup>. After entering the body of host, it directly goes to the rumen or stomach where it faces harsh conditions <sup>[123]</sup>. *S. Dublin* adopt to survive against the gastric acid and normal flora of the stomach/rumen <sup>[124]</sup>. After successfully adopting the conditions, the bacteria enter the epithelium of the

intestine for colonization purpose <sup>[125]</sup>. Intestinal motility and mucus work as defence of the host in the intestine and *S. Dublin* have fimbriae lipopolysaccharide to overcome the host defence <sup>[126]</sup>. Host secretes neutrophils and macrophages when the bacteria evade the host defences <sup>[127]</sup>. *S. Dublin* have the ability to secrete the effector proteins by TTSS-1 & 2 (Type III secretion system) that combat the macrophages and neutrophils <sup>[25]</sup>. When the bacteria overcome the host defence system, the bacteria spread systemically, and enteritis has been induced in the host <sup>[128,129]</sup>. In response to systemic salmonellosis, pyrexia has been developed <sup>[130]</sup>. Detailed pathogenesis is explained in Fig. 1.

## PUBLIC HEALTH IMPORTANCE OF BOVINE SALMONELLOSIS

Salmonellosis is an important public health concern around the globe that causes a large amount of mortality and morbidity <sup>[131]</sup>. Although most of the cases are self-limiting and moderate, the serious cases may lead to death <sup>[132]</sup>. Over 3 million deaths of the humans are because of the salmonellosis annually <sup>[133,134]</sup>. In US, 500-1000 deaths occur out of 2-4 million cases annually <sup>[135]</sup>. Regardless of the hygiene, education and food processing, salmonellosis still remains an important public health concern that needs attention <sup>[136]</sup>. *S. Dublin* cause infection in average 10 humans per year in Ireland <sup>[137]</sup>. In Ireland, bovine salmonellosis is considered as the most important public health disease as it is a major threat to the livestock of the Ireland and because of the genetic evolutions of *S. Dublin* <sup>[138]</sup>. It is believed to have evolved recently due to the consistency in its multi-locus enzyme genotype and fliC flagellin DNA sequence analysis <sup>[139]</sup>. *S. Dublin* is considered being one step away from *S. enteritidis* (a common *Salmonella* serotype in humans and poultry) <sup>[140,141]</sup>. However, the genetic distinction between these two serovars is insignificant <sup>[18]</sup>. If *S. Dublin* have been diagnosed in the herd of cattle, there is a significant risk of persistent infection in the carrier cows for as long as the cow remains in the herd <sup>[52,142]</sup>. To diagnose the bovine salmonellosis in the herd, there are multiple options which are given in detail as follows.

## DIAGNOSIS

Diagnosis of the disease in the herd of the animal is the most important key to prevent the economic loss and maintaining health of the animals <sup>[143,144]</sup>. Pathological, postmortem findings and clinical signs are not adequate for diagnosing salmonellosis <sup>[145]</sup>. For definitive diagnosis, there are multiple techniques including cultivation of bacteria, clinical examinations and counting the antibodies against *S. Dublin* <sup>[146]</sup>. Detection of bacteria in fluids, body organs, faeces, and environmental samples can be done by

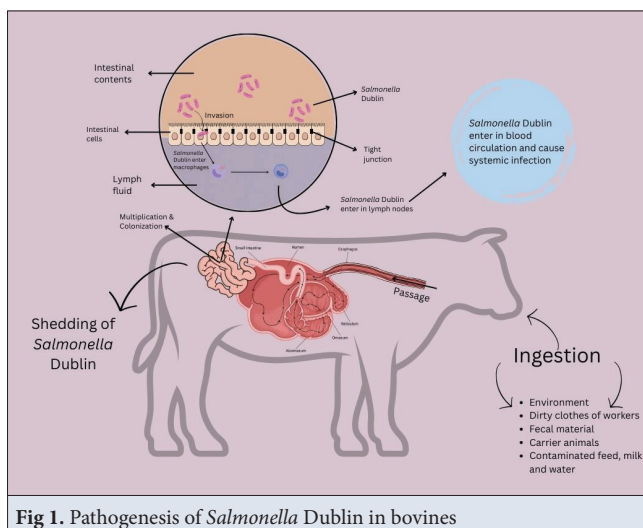


Fig 1. Pathogenesis of *Salmonella Dublin* in bovines



conventional culture methods of bacteria <sup>[147]</sup>. The main advantage of detecting bacteria is tracing infection in large groups of animals during the investigation of an outbreak. This method has a disadvantage of low sensitivity <sup>[148]</sup>. The new techniques for the detection of the bacteria are based on the genetic material, i.e., Polymerase chain reaction (PCR) techniques <sup>[149,150]</sup>. The PCR techniques are considered being the more sensitive techniques, but subsequent typing is not always possible <sup>[151,152]</sup>. The detailed diagnostic techniques are given as follows:

### Culture Techniques

Culturing techniques are stepwise procedure in order to isolate the live bacteria in the sample <sup>[153]</sup>. The steps to isolate the bacteria in the sample include pre-enrichment, selective enrichment, plating and confirmation <sup>[154]</sup>. For positive results, bacteria must have the ability to grow in the enrichment steps <sup>[155]</sup>. This method should be able to detect the minimum CFU in the sample; up to 1 CFU <sup>[156]</sup>. However, this may not be always true in every case. For faecal samples of *S. Dublin*, the specificity is assumed to be 100% while the sensitivity in faecal samples is less than ideal <sup>[157]</sup>. For more accurate sensitivity test, faecal cultures should be used repeatedly to detect the active carriers <sup>[158]</sup>.

### PCR Techniques

PCR-based techniques are considered being more precise and accurate to detect the bacteria in the sample material <sup>[159,160]</sup>. These techniques are based on the detection of the genetic material of the bacteria (*S. Dublin*) <sup>[141]</sup>. However, there are 2 basic principles of PCR technique: real-time PCR and the traditional PCR <sup>[161,162]</sup>. The traditional PCR technique only gives qualitative results (whether the bacteria is present or not) while in real-time PCR, the exact amount of the DNA copied after every cycle is counted by the computer <sup>[149,163]</sup>. The performance of the test depends on the proper functioning of probes, internal control systems, and primers <sup>[164]</sup>. However, the PCR tests do not provide the information of the serotype of the *Salmonella* <sup>[165]</sup>. To detect the serotype *S. Dublin* in the tested sample, a follow-up culture test must be conducted on the positive samples <sup>[166]</sup>. Multiple studies show that the specificity and sensitivity of the PCR tests is not accurate <sup>[167,168]</sup>. In a case of naturally infected animals with *S. Dublin*, the sensitivity results of rt-PCR found poorer than compared to the conventional culture method <sup>[169]</sup>.

### Antibodies Detection

Antibodies detection has been proven very effective in checking the previous or current infection <sup>[170,171]</sup>. Enzyme-linked immunosorbent assays (ELISA) can be used to detect the O-antigens from *S. Dublin* in milk and blood samples <sup>[172]</sup>. This measures the humoral

immune response that specifies the infection (previous or early) <sup>[173]</sup>. IgG antibody titre increases up to measurable amount in the animals after one too few weeks of the infection <sup>[174]</sup>. This method of diagnosis has multiple benefits of low cost and sampling feasibility. ELISA is used widely for the surveillance purpose, evaluation of control strategies and supporting decision <sup>[175]</sup>. In Denmark, bulk milk is collected regularly or alternative days for milk testing <sup>[176]</sup>. The samples have been tested for the surveillance of *S. Dublin*, Infectious Bovine Rhinotracheitis (IBR) and Bovine Viral Diarrhoea virus (BVDv) <sup>[177]</sup>. This routine testing helps in effective control strategies. Serum ELISA can also be performed to check the antibodies titre as its sensitivity is close to the individual milk ELISA testing <sup>[178]</sup>. However, other genomic and molecular methods are available for the differentiation of *S. Dublin* strain <sup>[179]</sup>. The details of these methods are beyond the scope of this review.

## CONCLUSION

Bovine salmonellosis caused by *Salmonella enterica* subsp. *enterica* serovar *Dublin* (*S. Dublin*) is a significant public health and economic concern globally. It leads to severe health issues in cattle, including enteritis, septicaemia, and abortion, with high mortality rates, especially in newborn calves. The disease not only impacts the well-being of the animals but also results in substantial economic losses through treatment costs, reduced milk production, and potential outbreaks. The transmission of *S. Dublin* is primarily through contaminated food, water, and environmental exposure, with the faecal route being the most significant mode of spread. The pathogenesis of *S. Dublin* involves complex interactions between the bacteria and the host immune system, with the bacteria capable of persisting in the herd as carriers, further complicating control measures. Effective control strategies are critical to minimizing its spread, and understanding the epidemiology, clinical signs, and diagnostic methods is key. Current diagnostic tools, such as culture techniques, PCR, and antibody detection methods, while valuable, have limitations and require further refinement to enhance sensitivity and specificity. From a public health perspective, *S. Dublin* represents a major risk, especially due to its potential to evolve and cross-infect humans. As such, continued vigilance and research into prevention, control strategies, and early detection are essential for mitigating the public health risk posed by bovine salmonellosis. The development of more efficient surveillance systems and control measures will be vital in preventing the spread of this zoonotic pathogen, ensuring the health of both livestock and humans.

## DECLARATIONS

**Acknowledgements:** The researchers would like to thank the Deanship of Graduate Studies and Scientific Research at Qassim University for financial support (QU-APC-2025).

**Conflict of Interest:** The authors declared that there is no conflict of interest.

**Generative Artificial Intelligence:** No Generative Artificial Intelligence was used in this research

**Author Contributions:** Idea/concept: AMA, Design: AMA, Data collection and processing: AIA, Analysis: AIA, Writing: AMA & AIA

## REFERENCES

- Galán-Relaño Á, Valero Díaz A, Huerta Lorenzo B, Gómez-Gascón L, Mena Rodríguez MÁ, Carrasco Jiménez E, Pérez Rodríguez F, Astorga Márquez RJ: *Salmonella* and salmonellosis: An update on public health implications and control strategies. *Animals*, 13 (23):3666, 2023. DOI: 10.3390/ani13233666
- Eng SK, Pusparajah P, Ab Mutalib NS, Ser HL, Chan KG, Lee LH: *Salmonella*: A review on pathogenesis, epidemiology and antibiotic resistance. *Front Life Sci*, 8 (3): 284-293, 2015. DOI: 10.1080/21553769.2015.1051243
- García-Soto S, Tomaso H, Linde J, Methner U: Epidemiological analysis of *Salmonella enterica* subsp. *enterica* serovar Dublin in German cattle herds using whole-genome sequencing. *Microbiol Spectr*, 9 (2): e00332-00321, 2021. DOI: 10.1128/Spectrum.00332-21
- Velasquez-Munoz A, Castro-Vargas R, Cullens-Nobis FM, Mani R, Abuelo A: *Salmonella* Dublin in dairy cattle. *Front Vet Sci*, 10:1331767, 2024. DOI: 10.3389/fvets.2023.1331767
- Boyle EC, Bishop JL, Grassl GA, Finlay BB: *Salmonella*: From pathogenesis to therapeutics. *J Bacteriol*, 189 (5): 1489-1495, 2007. DOI: 10.1128/JB.01730-06
- Powell J, Daly M, O'Connell NH, Dunne CP: Seek and you shall find: *Yersinia enterocolitica* in Ireland's drinking water. *Ir J Med Sci*, 193 (4): 1885-1890, 2024. DOI: 10.1007/s11845-024-03641-5
- Russell L, Whyte P, Zintl A, Gordon SV, Markey B, de Waal T, Nolan S, O'Flaherty V, Abram F, Richards K, Fenton O, Bolton D: The survival of *Salmonella* Senftenberg, *Escherichia coli* O157: H7, *Listeria monocytogenes*, *Enterococcus faecalis* and *Clostridium sporogenes* in sandy and clay loam textured soils when applied in bovine slurry or unpasteurised digestate and the run-off rate for a test bacterium, *Listeria innocua*, when applied to grass in slurry and digestate. *Front Sustain Food Syst*, 6:806920, 2022. DOI: 10.3389/fsufs.2022.806920
- Petrin S, Mancin M, Losasso C, Deotto S, Olsen JE, Barco L: Effect of pH and salinity on the ability of *Salmonella* serotypes to form biofilm. *Front Microbiol*, 13:821679, 2022. DOI: 10.3389/fmicb.2022.821679
- Polley S, Biswas S, Kesh SS, Maity A, Batabyal S: The link between animal manure and zoonotic disease. In: Mahajan S, Varma A (Eds): *Animal Manure: Agricultural and Biotechnological Applications*, 297-333, Springer, 2022.
- Pedersen L, Houe H, Rattenborg E, Nielsen LR: Semi-quantitative biosecurity assessment framework targeting prevention of the introduction and establishment of *Salmonella* Dublin in dairy cattle herds. *Animals*, 13 (16):2649, 2023. DOI: 10.3390/ani13162649
- Adil M, Rani Z, Ahmad S, Tariq M, Akram MS, Ajmal M, Murtaza G, Khan AMA, Qamar W, Zafar U, Gil S, Saeed M: A review on methicillin resistance in *Staphylococcus aureus* in dairy cows and its consequences. *Continental Vet J*, 3 (2): 32-42, 2023. DOI: 10.71081/cvj/2023.018
- Zhao Y, Bhavya ML, Patange A, Sun DW, Tiwari BK: Plasma-activated liquids for mitigating biofilms on food and food contact surfaces. *Compr Rev Food Sci Food Saf*, 22 (3): 1654-1685, 2023. DOI: 10.1111/1541-4337.13126
- do Amarante VS, de Castro Pereira JK, Serafini MF, Ramos CP, Zanon IP, de Souza TGV, Moreira TF, de Carvalho AU, Meneses RM, Aburjaile FF: Dynamics of *Salmonella* Dublin infection and antimicrobial resistance in a dairy herd endemic to salmonellosis. *PLoS One*, 20 (1):e0318007, 2025. DOI: 10.1371/journal.pone.0318007
- Srednik ME, Lantz K, Hicks JA, Morningstar-Shaw BR, Mackie TA, Schlater LK: Antimicrobial resistance and genomic characterization of *Salmonella* Dublin isolates in cattle from the States. *PLoS One*, 16 (9):e0249617, 2021. DOI: 10.1371/journal.pone.0249617
- Sia CM, Ambrose RL, Valcanis M, Andersson P, Ballard SA, Howden BP, Williamson DA, Pearson JS, Ingle DJ: Distinct adaptation and epidemiological success of different genotypes within *Salmonella enterica* serovar Dublin. *Biorxiv*, 2024-2007, 2024. DOI: 10.1101/2024.07.30.605691
- Sia CM, Ambrose RL, Valcanis M, Andersson P, Ballard SA, Howden BP, Williamson DA, Pearson JS, Ingle DJ: Dynamics of antimicrobial resistance and virulence of *Salmonella enterica* serovar Dublin. *eLife*, 13:RP102253, 2024. DOI: 10.7554/eLife.102253.1
- Robi DT, Mossie T, Tememe S: A comprehensive review of the common bacterial infections in dairy calves and advanced strategies for health management. *Vet Med (Auckl)*, 1-14, 2024. DOI: 10.2147/VMRR.S452925
- Casaux ML, Neto WS, Schild CO, Costa RA, Macías-Rioseco M, Caffarena RD, Silveira CS, Araújo V, Díaz BD, Giannitti F: Epidemiological and clinicopathological findings in 15 fatal outbreaks of salmonellosis in dairy calves and virulence genes in the causative *Salmonella enterica* Typhimurium and Dublin strains. *Braz J Microbiol*, 54 (1): 475-490, 2023. DOI: 10.1007/s42770-022-00898-9
- Saleem M, Rahman HU, Abbas J: Rapid recovery of *Salmonella* from chicken meat and poultry fecal samples by selective pre-enrichment. *Continental Vet J*, 3 (1): 49-53, 2023. DOI: 10.71081/cvj/2023.007
- Yanmaz B, Özgen EK: Identification and phylogenetic positioning of *Salmonella* Dublin from aborted cattle materials. *Kafkas Univ Vet Fak Derg*, 27 (6): 781-786, 2021. DOI: 10.9775/kvfd.2021.26315
- Mkangara M: Prevention and control of human *Salmonella enterica* infections: An implication in food safety. *Int J Food Sci*, 2023 (1):8899596, 2023. DOI: 10.1155/2023/8899596
- Dall Agnol AM, Lorenzetti E, Leme RA, Ladeia WA, Mainardi RM, Bernardi A, Headley SA, Freire RL, Pereira UP, Alfieri AF: Severe outbreak of bovine neonatal diarrhea in a dairy calf rearing unit with multifactorial etiology. *Braz J Microbiol*, 52 (4): 2547-2553, 2021. DOI: 10.1007/s42770-021-00565-5
- Brown SE, Bycroft KA, Adam K, Collett MG: Acute fibrinous pleuropneumonia and septicemia caused by *Bibersteinia trehalosi* in neonatal calves in New Zealand. *N Z Vet J*, 69 (1): 51-57, 2021. DOI: 10.1080/00480169.2020.1792372
- Boyd E, Dick J, Millar C, Ghosh K, Arya G, Himsworth C: A retrospective analysis of postmortem *Salmonella* Dublin cases in dairy cattle in British Columbia. *Transbound Emerg Dis*, 2024 (1):9461144, 2024. DOI: 10.1155/2024/9461144
- Lamichhane B, Mawad AMM, Saleh M, Kelley WG, Harrington PJ, Lovestad CW, Amezcua J, Sarhan MM, El Zowalaty ME, Ramadan H: Salmonellosis: An overview of epidemiology, pathogenesis, and innovative approaches to mitigate the antimicrobial resistant infections. *Antibiotics*, 13 (1):76, 2024. DOI: 10.3390/antibiotics13010076
- Salavatiha Z, Tavakoli A, Kiani SJ, Rezvani MR, Mokarinejad R, Monavari SH: Investigation the prevalence of norovirus, rotavirus, human bocavirus, and adenovirus in inpatient children with gastroenteritis in Tehran, Iran, during 2021-2022. *Iran J Med Microbiol*, 18 (4): 230-237, 2024. DOI: 10.30699/ijmm.18.1.25
- Hugho EA, Mmbaga BT, Lukumbagire A-HS, Kinabo GD, Thomas KM, Kumburu HH, Hald T: Risk factors for *Salmonella* infection in children under five years: A hospital-based study in Kilimanjaro Region, Tanzania. *Pathogens*, 13 (9):798, 2024. DOI: 10.3390/pathogens13090798
- Eddy RG: *Alimentary conditions*. In: *Bovine Medicine Diseases and Husbandry of Cattle*. 2<sup>nd</sup> ed., 821-859, Blackwell, 2004.
- Mohamed HE, Ibrahim HN, Ibrahim GA: Marbofloxacin influence on haemato-biochemical alterations in diarrheic calves infected with *Salmonella* spp. *J Adv Vet Res*, 13 (6): 1027-1036, 2023.
- Nagati SF, Hammad HO, Abou-Khadra SH, Farhan HE, Afify AE, Hassanien RT, Elnady AM, Mansour SS, Shahein MA: Longitudinal

study of some bacterial, parasitic, and viral enteric pathogens isolated from diarrheic calves from dairy herd in Egypt. *J Adv Vet Res*, 13 (6): 1214-1226, 2023.

31. Boyd E, Cuthbert E, Dick J, Ghosh K, Leung D, Renaud DL, Himsworth C: Understanding *Salmonella* Dublin in British Columbia through bulk tank milk surveillance. *J Dairy Sci*, 108 (3): 2749-2755, 2025. DOI: 10.3168/jds.2024-25710
32. Raut R, Maharjan P, Fouladkhah AC: Practical preventive considerations for reducing the public health burden of poultry-related salmonellosis. *Int J Environ Res Public Health*, 20 (17):6654, 2023. DOI: 10.3390/ijerph20176654
33. Alhumaidan OS: Comprehensive review of salmonellosis: Current status of the disease and future perspectives. *Ital J Food Saf*, 13 (4): 12904, 2024. DOI: 10.4081/ijfs.2024.12904
34. Ayuti SR, Khairullah AR, Al-Arif MA, Lamid M, Warsito SH, Moses IB, Hermawan IP, Silaen OSM, Lokapirnasari WP, Aryaloka S: Tackling salmonellosis: A comprehensive exploration of risks factors, impacts, and solutions. *Open Vet J*, 14 (6): 1313, 2024. DOI: 10.5455/OVJ.2024.v14.i6.1
35. Almuzaini AM: Phytochemicals: potential alternative strategy to fight *Salmonella enterica* serovar Typhimurium. *Front Vet Sci*, 10, 1188752-1188752, 2023. DOI: 10.3389/fvets.2023.1188752
36. Abbas RZ, Qureshi MA, Saeed Z: Botanical compounds: A promising control strategy against *Trypanosoma cruzi*. *Bol Latinoam Caribe Plantas Med Aromat*, 24 (3): 308-327, 2025. DOI: 10.37360/blacpma.25.24.3.23
37. Durrani RH, Sheikh AA, Humza M, Ashraf S, Kokab A, Mahmood T, Khan MUZ: Evaluation of Antibiotic resistance profile and multiple antibiotic resistance index in avian adapted *Salmonella enterica* serovar Gallinarum isolates. *Pak Vet J*, 44 (4): 1349-1352, 2024. DOI: 10.29261/pakvetj/2024.253
38. Wu X, Wang Y, Wang Q, Wang Y, Wang H, Luo X: Acinetobacter of pigs reveals high multiple drug resistance through genomics and antimicrobial resistance monitoring. *Pak Vet J*, 44 (4): 1284-1290, 2024. DOI: 10.29261/pakvetj/2024.259
39. Bui MTL, Nguyen TT, Nguyen HC, Ly KLT, Nguyen TK: Antibiotic resistance and pathogenicity of *Escherichia coli* isolated from cattle raised in households in the Mekong Delta, Vietnam. *Int J Vet Sci*, 13 (5): 730-736, 2024. DOI: 10.47278/journal.ijvs/2024.166
40. Oludoun OY, Adeniyi MO, Ogunlaran OM, Akinola EI, Abiodun OE: Transmission analysis of *Salmonella* Dublin of dairy calves. In, *IOP Conference Series: Earth and Environmental Science*, 1219 012019. IOP Publishing, 2023.
41. Ali S, Alsayeqh AF: Review of major meat-borne zoonotic bacterial pathogens. *Front Public Health*, 10:1045599, 2022. DOI: 10.3389/fpubh.2022.1045599
42. Fritz HM, Pereira RV, Toohey-Kurth K, Marshall E, Tucker J, Clothier KA: *Salmonella enterica* serovar Dublin from cattle in California from 1993-2019: Antimicrobial resistance trends of clinical relevance. *Antibiotics*, 11 (8):1110, 2022. DOI: 10.3390/antibiotics11081110
43. Ménard S, Lacroix-Lamandé S, Ehrhardt K, Yan J, Grassl GA, Wiedemann A: Cross-talk between the intestinal epithelium and *Salmonella* Typhimurium. *Front Microbiol*, 13:906238, 2022. DOI: 10.3389/fmicb.2022.906238
44. Ijaz A, Veldhuizen EJA, Broere F, Rutten VPMG, Jansen CA: The interplay between *Salmonella* and intestinal innate immune cells in chickens. *Pathogens*, 10 (11):1512, 2021. DOI: 10.3390/pathogens10111512
45. Richards AF, Torres-Velez FJ, Mantis NJ: *Salmonella* uptake into gut-associated lymphoid tissues: Implications for Targeted mucosal vaccine design and delivery. *Methods Mol Biol*, 2410, 305-324, 2022. DOI: 10.1007/978-1-0716-1884-4\_15
46. Dai Y, Zhang M, Liu X, Sun T, Qi W, Ding W, Chen Z, Zhang P, Liu R, Chen H: *Salmonella* manipulates macrophage migration via SteC-mediated myosin light chain activation to penetrate the gut-vascular barrier. *EMBO J*, 43 (8): 1499-1518, 2024. DOI: 10.1038/s44318-024-00076-7
47. Li W, Ren Q, Ni T, Zhao Y, Sang Z, Luo R, Li Z, Li S: Strategies adopted by *Salmonella* to survive in host: A review. *Arch Microbiol*, 205 (12):362, 2023. DOI: 10.1007/s00203-023-03702-w
48. Huang H, Naushad S: *Salmonella*: Perspectives for Low-Cost Prevention, Control and Treatment. In, BoD-Books on Demand, 2024.
49. Paul J: Blood and lymphatic infections. In, Huang H, Naushad S (Eds): *Disease Causing Microbes*. 247-314, Springer, 2024.
50. Nielsen LR, Schukken YH, Gröhn YT, Ersbøll AK: *Salmonella* Dublin infection in dairy cattle: Risk factors for becoming a carrier. *Prev Vet Med*, 65 (1-2): 47-62, 2004. DOI: 10.1016/j.prevetmed.2004.06.010
51. Nielsen LR: Review of pathogenesis and diagnostic methods of immediate relevance for epidemiology and control of *Salmonella* Dublin in cattle. *Vet Microbiol*, 162 (1): 1-9, 2013. DOI: 10.1016/j.vetmic.2012.08.003
52. Castro-Vargas RE, Cullens-Nobis FM, Mani R, Roberts JN, Abuelo A: Effect of dry period immunization of *Salmonella* Dublin latent carriers with a commercial live culture vaccine on intrauterine transmission based on the presence of precolostral antibodies in offspring. *J Dairy Sci*, 107 (12): 11436-11445, 2024. DOI: 10.3168/jds.2024-24945
53. Lawson GHK, McPherson EA, Laing AH, Wooding P: The epidemiology of *Salmonella* Dublin infection in a dairy herd: I. Excretion and persistence of the organism. *J Hyg*, 72 (3): 311-328, 1974. DOI: 10.1017/s0022172400023548
54. Wang F, Wang L, Ge H, Wang X, Guo Y, Xu Z, Geng S, Jiao X, Chen X: Safety of the *Salmonella enterica* serotype Dublin strain Sdu189-derived live attenuated vaccine - A pilot study. *Front Vet Sci*, 9:986332, 2022. DOI: 10.3389/fvets.2022.986332
55. Foster N, Tang Y, Berchieri A, Geng S, Jiao X, Barrow P: Revisiting persistent *Salmonella* infection and the carrier state: What do we know? *Pathogens*, 10 (10):1299, 2021. DOI: 10.3390/pathogens10101299
56. Harvey RR, Friedman CR, Crim SM, Judd M, Barrett KA, Tolar B, Folster JP, Griffin PM, Brown AC: Epidemiology of *Salmonella enterica* serotype Dublin infections among humans, United States, 1968-2013. *Emerg Infect Dis*, 23 (9):1493, 2017. DOI: 10.3201/eid2309.170136
57. Sullivan DJ, Moran GP, Pinjon E, Al-Mosaid A, Stokes C, Vaughan C, Coleman DC: Comparison of the epidemiology, drug resistance mechanisms, and virulence of *Candida dubliniensis* and *Candida albicans*. *FEMS Yeast Res*, 4 (4-5): 369-376, 2004. DOI: 10.1016/s1567-1356(03)00240-x
58. Martinez-Sanguiné AY, D'alessandro B, Langleib M, Traglia GM, Mónaco A, Durán R, Chabalgoity JA, Betancor L, Yim L: *Salmonella enterica* serovars Dublin and enteritidis comparative proteomics reveals differential expression of proteins involved in stress resistance, virulence, and anaerobic metabolism. *Infect Immun*, 89 (3):e00606-20, 2021. DOI: 10.1128/IAI.00606-20
59. Townsend L, Fogarty H, Dyer A, Martin-Loeches I, Bannan C, Nadarajan P, Bergin C, O'Farrelly C, Conlon N, Bourke NM: Prolonged elevation of D-dimer levels in convalescent COVID-19 patients is independent of the acute phase response. *J Thromb Haemost*, 19 (4): 1064-1070, 2021. DOI: 10.1111/jth.15267
60. Holschbach CL, Breuer RM, Pohly AE, Crawford C, Aulik NA: Multi-drug resistant *Salmonella* ser. Dublin cultured from cryopreserved Holstein semen. *Vet Rec Case Rep*, 12 (1):e791, 2024. DOI: 10.1002/vrc.2.791
61. Qureshi MA, Fatima Z, Muqadas SML, Najaf DE, Husnain M, Moeed HA, Ijaz U: Zoonotic diseases caused by mastitic milk. In, Altaf S, Khan A, Abbas RZ (Eds): *Zoonosis*. Vol. 4, 557-572, Unique Scientific Publishers, Faisalabad, Pakistan, 2023.
62. Tommasoni C, Schiavon E, Lisuzzo A, Giancesella M, Merenda M, Coin P, Patreggiani T, Tola S, Catania S, Barberio A: *Salmonella enterica* serovar Dublin infection in dairy cattle: a case study on the management of an outbreak in Italy. *Large Anim Rev*, 29 (2): 99-103, 2023.
63. Perry KV, Kelton DF, Dufour S, Miltenburg C, Sedo SGU, Renaud DL: Risk factors for *Salmonella* Dublin on dairy farms in Ontario, Canada. *J Dairy Sci*, 106 (12): 9426-9439, 2023. DOI: 10.3168/jds.2023-23517
64. Lourenco JM, Welch CB: Using microbiome information to understand and improve animal performance. *Ital J Anim Sci*, 21 (1): 899-913, 2022. DOI: 10.1080/1828051x.2022.2077147
65. Ovuru KE, Izah SC, Ogidi OI, Imarhiagbe O, Ogbu MC: Slaughterhouse facilities in developing nations: Sanitation and hygiene practices, microbial contaminants and sustainable management system. *Food Sci Biotechnol*, 33 (3): 519-537, 2024. DOI: 10.1007/s10068-023-01406-x



66. Guo M, Tao W, Flavell RA, Zhu S: Potential intestinal infection and faecal-oral transmission of SARS-CoV-2. *Nat Rev Gastroenterol Hepatol*, 18 (4): 269-283, 2021. DOI: 10.1038/s41575-021-00416-6
67. Godijk NG, Bootsma MCJ, Bonten MJM: Transmission routes of antibiotic resistant bacteria: A systematic review. *BMC Infect Dis*, 22 (1):482, 2022. DOI: 10.1186/s12879-022-07360-z
68. Li M, Zhou Y, Cheng J, Wang Y, Lan C, Shen Y: Response of the mosquito immune system and symbiotic bacteria to pathogen infection. *Parasit Vectors*, 17 (1):69, 2024. DOI: 10.1186/s13071-024-06161-4
69. Nooraei S, Sarkar Lotfabadi A, Akbarzadehmoallemkolaei M, Rezaei N: Immunogenicity of different types of adjuvants and nano-adjuvants in veterinary vaccines: A comprehensive review. *Vaccines*, 11 (2):453, 2023. DOI: 10.3390/vaccines11020453
70. Pan T, Sun S, Chen Y, Tian R, Chen E, Tan R, Wang X, Liu Z, Liu J, Qu H: Immune effects of PI3K/Akt/HIF-1 $\alpha$ -regulated glycolysis in polymorphonuclear neutrophils during sepsis. *Crit Care*, 26 (1):29, 2022. DOI: 10.1186/s13054-022-03893-6
71. Miles MA, Luong R, To EE, Erlich JR, Liong S, Liong F, Logan JM, O'Leary J, Brooks DA, Selemidis S: TLR9 monotherapy in immune-competent mice suppresses orthotopic prostate tumor development. *Cells*, 13 (1):97, 2024. DOI: 10.3390/cells13010097
72. Blanco FC, y Garcia JS, Bigi F: Recent advances in non-specific immune memory against bovine tuberculosis. *Comp Immunol Microbiol Infect Dis*, 75:101615, 2021. DOI: 10.1016/j.cimid.2021.101615
73. Rajme-Manzur D, Gollas-Galván T, Vargas-Albores F, Martínez-Porchas M, Hernández-Oñate MÁ, Hernández-López J: Granulomatous bacterial diseases in fish: An overview of the host's immune response. *Comp Biochem Physiol A Mol Integr Physiol*, 261:111058, 2021. DOI: 10.1016/j.cbpa.2021.111058
74. Johnston PI, Bogue P, Chirambo AC, Mbewe M, Prakash R, Kandoole-Kabwere V, Lester R, Darton T, Baker S, Gordon MA: Bacterial shedding and serologic responses following an outbreak of *Salmonella* Typhi in an endemic cohort. *BMC Infect Dis*, 23 (1):416, 2023. DOI: 10.1186/s12879-023-08385-8
75. Verma S, Singh K, Bansal A: Multi-epitope DnaK peptide vaccine accords protection against lethal *S. typhimurium* challenge: elicits both cell mediated immunity and long-lasting serum-neutralizing antibody titers. *Pharmacol Res*, 169:105652, 2021. DOI: 10.1016/j.phrs.2021.105652
76. Sekhwal MK, Li L, Pierre T, Matthews T, Luley E, Tewari D, Kuchipudi SV, Jayarao B, Byukusenge M: Molecular epidemiology of *Salmonella enterica* serotype Dublin isolated from 2011 to 2022 from veal and dairy cattle in Pennsylvania. *Microorganisms*, 13 (2):400, 2025. DOI: 10.3390/microorganisms13020400
77. Sarkar A, McInroy CJA, Harty S, Raulo A, Ibata NGO, Valles-Colomer M, Johnson KVA, Brito IL, Henrich J, Archie EA: Microbial transmission in the social microbiome and host health and disease. *Cell*, 187 (1): 17-43, 2024. DOI: 10.1016/j.cell.2023.12.014
78. Sheedy FJ, Divangahi M: Targeting immunometabolism in host defence against *Mycobacterium tuberculosis*. *Immunology*, 162 (2): 145-159, 2021. DOI: 10.1111/imm.13276
79. Nielsen LR, Houe H, Nielsen SS: Narrative review comparing principles and instruments used in three active surveillance and control programmes for Non-EU-regulated diseases in the Danish cattle population. *Front Vet Sci*, 8:685857, 2021. DOI: 10.3389/fvets.2021.685857
80. Elbehiry A, Marzouk E, Aldubaib M, Moussa I, Abalkhail A, Ibrahim M, Hamada M, Sindi W, Alzaben F, Almuzaini AM, Algamal AM, Rawway M: *Pseudomonas* species prevalence, protein analysis, and antibiotic resistance: An evolving public health challenge. *AMB Express*, 12 (1): 53-53, 2022. DOI: 10.1186/s13568-022-01390-1
81. Almuzaini AM, Aljohani ASM, Alajaji AI, Elbehiry A, Abalkhail A, Almujaidel A, Aljarallah SN, Sherif HR, Marzouk E, Draz AA: Seroprevalence of brucellosis in camels and humans in the Al-Qassim region of Saudi Arabia and its implications for public health. *AMB Express*, 15 (1): 22-22, 2025. DOI: 10.1186/s13568-025-01822-8
82. Papoula-Pereira R, Alvseike O, Cenci-Goga BT, Grisoldi L, Nagel-Alne GE, Ros-Lis JV, Thomas L: Economic evidence for the control of *Salmonella* in animal-derived food systems: A scoping review. *Food Control*, 175:111275, 2025. DOI: 10.1016/j.foodcont.2025.111275
83. Whatford L, van Winden S, Häslér B: A systematic literature review on the economic impact of endemic disease in UK sheep and cattle using a One Health conceptualisation. *Prev Vet Med*, 209:105756, 2022. DOI: 10.1016/j.prevetmed.2022.105756
84. Verma S, Malik YS, Singh G, Dhar P, Singla AK: Key concepts and principles of epidemiology, prevention, and control of animal diseases. In, *Core Competencies of a Veterinary Graduate*. Springer, Singapore, 2024.
85. Clemmons EA, Alfson KJ, Dutton III JW: Transboundary animal diseases, an overview of 17 diseases with potential for global spread and serious consequences. *Animals*, 11 (7):2039, 2021. DOI: 10.3390/ani11072039
86. Szelényi Z, Szenci O, Bodó S, Kovács L: Noninfectious causes of pregnancy loss at the late embryonic/early fetal stage in dairy cattle. *Animals*, 13 (21):3390, 2023. DOI: 10.3390/ani13213390
87. Hofer K, Trockenbacher B, Sodoma E, Khol JL, Dünser M, Wittek T: Establishing a surveillance programme for *Salmonella* Dublin in Austrian dairy herds by comparing herd-level vs. individual animal detection methods. *Prev Vet Med*, 230:106277, 2024. DOI: 10.1016/j.prevetmed.2024.106277
88. Um MM, Dufour S, Bergeron L, Gauthier M-L, Paradis M-È, Roy J-P, Falcon M, Molgat E, Ravel A: Development of a decision support tool to compare diagnostic strategies for establishing the herd status for infectious diseases: An example with *Salmonella* Dublin infection in dairies. *Prev Vet Med*, 228:106234, 2024. DOI: 10.1016/j.prevetmed.2024.106234
89. Sharif MK, Sarwar K, Abid N, Bashir MA: Food security, food safety, and sanitation. In, *Bashir MK, Schilizzi SGM, Ali G (Eds): Food Security in the Developing World*. 191-225, Wiley, 2024.
90. Kemal J: A review on the public health importance of bovine salmonellosis. *J Vet Sci Technol*, 5 (2): 1-10, 2014. DOI: 10.4172/2157-7579.1000175
91. Owusu-Apenten R, Vieira E: Microbial foodborne disease outbreaks. In, *Elementary Food Science*. 171-196, Springer, 2022.
92. Pal M, Teashal BM, Gizaw F, Alemayehu G, Kandi V: Animals and food of animal origin as a potential source of Salmonellosis: A review of the epidemiology, laboratory diagnosis, economic impact and public health significance. *Am J Microbiol Res*, 8 (2): 48-56, 2020. DOI: 10.12691/ajeid-5-2-2
93. Kudirkienė E, Sørensen G, Torpdahl M, de Knecht LV, Nielsen LR, Rattenborg E, Ahmed S, Olsen JE: Epidemiology of *Salmonella enterica* serovar Dublin in cattle and humans in Denmark, 1996 to 2016: A retrospective whole-genome-based study. *Appl Environ Microbiol*, 86 (3): e01894-01819, 2020. DOI: 10.1128/AEM.01894-19
94. Nielsen LR, Kudahl AB, Østergaard S: Age-structured dynamic, stochastic and mechanistic simulation model of *Salmonella* Dublin infection within dairy herds. *Prev Vet Med*, 105 (1-2): 59-74, 2012. DOI: 10.1016/j.prevetmed.2012.02.005
95. Mõtus K, Rilanto T, Viidu D-A, Orro T, Viltrop A: Seroprevalence of selected endemic infectious diseases in large-scale Estonian dairy herds and their associations with cow longevity and culling rates. *Prev Vet Med*, 192:105389, 2021. DOI: 10.1016/j.prevetmed.2021.105389
96. Cuevas-Gómez I, McGee M, Sánchez JM, O'Riordan E, Byrne N, McDaniel T, Earley B: Association between clinical respiratory signs, lung lesions detected by thoracic ultrasonography and growth performance in pre-weaned dairy calves. *Ir Vet J*, 74:7, 2021. DOI: 10.1186/s13620-021-00187-1
97. Janik E, Ceremuga M, Niemcewicz M, Bijak M: Dangerous pathogens as a potential problem for public health. *Medicina*, 56 (11):591, 2020. DOI: 10.3390/medicina56110591
98. Konieczny K, Pomorska-Mól M: A literature review of selected bacterial diseases in Alpacas and Llamas - Epidemiology, Clinical signs and Diagnostics. *Animals*, 14 (1):45, 2023. DOI: 10.3390/ani14010045
99. Singh K, Aulakh NS, Prakash B: Strategic detection of food contaminants using nanoparticle-based paper sensors. *J Food Saf*, 43 (6):e13089, 2023. DOI: 10.1111/jfs.13089

100. Siepkner CL, Schwartz KJ, Feldhacker TJ, Magstadt DR, Sahin O, Almeida M, Li G, Hayman KP, Gorden PJ: *Salmonella enterica* serovar Brandenburg abortions in dairy cattle. *J Vet Diagn Invest*, 34 (5): 864-869, 2022. DOI: 10.1177/10406387221105890
101. Salman M, Steneroden K: Important zoonotic diseases of cattle and their prevention measures. In, Sing A (Ed): *Zoonoses: Infections Affecting Humans and Animals*. 1-22, Springer, 2022.
102. Boulianne M, Blackall PJ, Hofacre CL, Ruiz JA, Sandhu TS, Hafez HM, Chin RP, Register KB, Jackwood MW: Pasteurellosis and other respiratory bacterial infections. In, Swayne DE, Boulianne M, Logue CM, McDougald LR, Nair V, Suarez DL, de Wit S, Grimes T, Johnson D, Kromm M, Prajito TY, Rubinoff I, Zavala G (Eds): *Diseases of Poultry*. 831-889, Wiley, 2020.
103. Hecker YP, González-Ortega S, Cano S, Ortega-Mora LM, Horcajo P: Bovine infectious abortion: A systematic review and meta-analysis. *Front Vet Sci*, 10:1249410, 2023. DOI: 10.3389/fvets.2023.1249410
104. Azaldegui I, Fiorentino MA, Morrell E, Odriozola E, García JA, Cantón G: Salmonellosis in adult cattle in Central Argentina: Case series. *Braz J Microbiol*, 55 (3): 2991-2996, 2024. DOI: 10.1007/s42770-024-01419-6
105. Mee JF: Investigation of bovine abortion and stillbirth/perinatal mortality-similar diagnostic challenges, different approaches. *Ir Vet J*, 73 (1):20, 2020. DOI: 10.1186/s13620-020-00172-0
106. Abd-Elrahman AH, Khafaga AE, Abas OM: The first identification of contagious caprine pleuropneumonia (CCPP) in sheep and goats in Egypt: Molecular and pathological characterization. *Trop Anim Health Prod*, 52, 1179-1186, 2020. DOI: 10.1007/s11250-019-02116-5
107. Poitras P, Ghia JE, Sawadogo A, Deslandres C, Wassef R, Dapoigny M, Bernstein C: The colon. In, Poitras P, Bilodeau M, Bouin M, Ghia JE (Eds): *The Digestive System: From Basic Sciences to Clinical Practice*. 125-171, Springer, 2022.
108. Mahmoud MAM, Megahed G, Yousef MS, Ali FAZ, Zaki RS, Abdelhafeez HH: *Salmonella* typhimurium triggered unilateral epididymorchitis and splenomegaly in a Holstein bull in Assiut, Egypt: A case report. *Pathogens*, 9 (4):314, 2020. DOI: 10.3390/pathogens9040314
109. Wulcan JM, Ketzis JK, Dennis MM: Typhlitis associated with natural *Trichuris* sp. infection in cats. *Vet Pathol*, 57 (2): 266-271, 2020. DOI: 10.1177/0300985819898894
110. Wilson DJ, Kelly EJ, Gucwa S: Causes of mortality of dairy cattle diagnosed by complete necropsy. *Animals*, 12 (21):3001, 2022. DOI: 10.3390/ani12213001
111. Guo P, Zhang K, Ma X, He P: *Clostridium* species as probiotics: Potentials and challenges. *J Anim Sci Biotechnol*, 11:24, 2020. DOI: 10.1186/s40104-019-0402-1
112. de Knecht LV, Kudirkiene E, Rattenborg E, Sorensen G, Denwood MJ, Olsen JE, Nielsen LR: Combining *Salmonella* Dublin genome information and contact-tracing to substantiate a new approach for improved detection of infectious transmission routes in cattle populations. *Prev Vet Med*, 181:104531, 2020. DOI: 10.1016/j.prevetmed.2018.09.005
113. Pharo F, Serrenho RC, Greer AL, Oremush R, Habing G, Gillies M, Keunen A, Renaud DL: Exploring the impact and transmission of *Salmonella* Dublin in crossbred dairy calves. *J Dairy Sci*, 108 (4): 4225-4233, 2025. DOI: 10.3168/jds.2024-25875
114. Goonewardene KB, Onyilagha C, Goolia M, Le VP, Blome S, Ambagala A: Superficial inguinal lymph nodes for screening dead pigs for African swine fever. *Viruses*, 14 (1):83, 2022. DOI: 10.3390/v14010083
115. Abebe E, Gugsa G, Ahmed M: Review on major food-borne zoonotic bacterial pathogens. *J Trop Med*, 2020;4674235, 2020. DOI: 10.1155/2020/4674235
116. Huang K, Fresno AH, Skov S, Olsen JE: Dynamics and outcome of macrophage interaction between *Salmonella* Gallinarum, *Salmonella* Typhimurium, and *Salmonella* Dublin and macrophages from chicken and cattle. *Front Cell Infect Microbiol*, 9:420, 2020. DOI: 10.3389/fcimb.2019.00420
117. Moreira MAS, Júnior AS, Lima MC, da Costa SL: Infectious diseases in dairy cattle. In, Nero LA, De Carvalho AF (Eds): *Raw Milked*. 235-258, Elsevier, 2019.
118. Menanteau P, Kempf F, Trottereau J, Virlogeux-Payant I, Gitten E, Dalifard J, Gabriel I, Rychlik I, Velge P: Role of systemic infection, cross contaminations and super-shedders in *Salmonella* carrier state in chicken. *Environ Microbiol*, 20 (9): 3246-3260, 2018. DOI: 10.1111/1462-2920.14294
119. Kitchens SR, Wang C, Price SB: Bridging classical methodologies in *Salmonella* investigation with modern technologies: A comprehensive review. *Microorganisms*, 12 (11):2249, 2024. DOI: 10.3390/microorganisms12112249
120. Gopinath S, Carden S, Monack D: Shedding light on *Salmonella* carriers. *Trends Microbiol*, 20 (7): 320-327, 2012. DOI: 10.1016/j.tim.2012.04.004
121. Dumontet S, Scopa A, Kerje S, Krovacek K: The importance of pathogenic organisms in sewage and sewage sludge. *J Air Waste Manag Assoc*, 51 (6): 848-860, 2001. DOI: 10.1080/10473289.2001.10464313
122. Vohra P, Vrettou C, Hope JC, Hopkins J, Stevens MP: Nature and consequences of interactions between *Salmonella enterica* serovar Dublin and host cells in cattle. *Vet Res*, 50 (1):99, 2019. DOI: 10.1186/s13567-019-0720-5
123. Meli G, Guerrini A, Tedesco DEA, Savoini G, Invernizzi G: Micro-cooling interventions improved the resilience to heat stress of Italian Holstein heifers. *J Dairy Sci*, 107 (Suppl. 1): 60-61, 2024.
124. Stevens MP, Kingsley RA: *Salmonella* pathogenesis and host-adaptation in farmed animals. *Curr Opin Microbiol*, 63, 52-58, 2021. DOI: 10.1016/j.mib.2021.05.013
125. Rogers AP, Mileto SJ, Lyras D: Impact of enteric bacterial infections at and beyond the epithelial barrier. *Nat Rev Microbiol*, 21 (4): 260-274, 2023. DOI: 10.1038/s41579-022-00794-x
126. Zhou G, Zhao Y, Ma Q, Li Q, Wang S, Shi H: Manipulation of host immune defenses by effector proteins delivered from multiple secretion systems of *Salmonella* and its application in vaccine research. *Front Immunol*, 14:1152017, 2023. DOI: 10.3389/fimmu.2023.1152017
127. Cacciottolo C, Alberti A: Eating the enemy: Mycoplasma strategies to evade neutrophil extracellular traps (NETs) promoting bacterial nucleotides uptake and inflammatory damage. *Int J Mol Sci*, 23 (23):15030, 2022. DOI: 10.3390/ijms232315030
128. Khan I, Bai Y, Zha L, Ullah N, Ullah H, Shah SRH, Sun H, Zhang C: Mechanism of the gut microbiota colonization resistance and enteric pathogen infection. *Front Cell Infect Microbiol*, 11:716299, 2021. DOI: 10.3389/fcimb.2021.716299
129. Lian S, Liu J, Wu Y, Xia P, Zhu G: Bacterial and viral co-infection in the intestine: competition scenario and their effect on host immunity. *Int J Mol Sci*, 23 (4):2311, 2022. DOI: 10.3390/ijms23042311
130. Boroujeni BM, Ghandali MV, Saki N, Ekrami A, Dezfili AAZ, Yousefi-Avarvand A: Mini review *Salmonella*: A problem in patients with sickle cell anemia. *Gene Rep*, 23:101118, 2021. DOI: 10.1016/j.genrep.2021.101118
131. Marchello CS, Birkhold M, Crump JA, Martin LB, Ansah MO, Breggi G, Canals R, Fiorino F, Gordon MA, Kim JH: Complications and mortality of non-typhoidal *Salmonella* invasive disease: A global systematic review and meta-analysis. *Lancet Infect Dis*, 22 (5): 692-705, 2022. DOI: 10.1016/S1473-3099(21)00615-0
132. Griffith RW, Carlson SA, Krull AC: Salmonellosis. In, *Diseases of Swine*. 912-925, Wiley, 2019.
133. Sanni AO, Onyango J, Rota AF, Mikecz O, Usman A, PicaCiamarra U, Fasina FO: Underestimated economic and social burdens of non-typhoidal *Salmonella* infections: The One Health perspective from Nigeria. *One Health*, 16:100546, 2023. DOI: 10.1016/j.onehlt.2023.100546
134. Teklemariam AD, Al-Hindi RR, Albiheyri RS, Alharbi MG, Alghamdi MA, Filimban AAR, Al Mutiri AS, Al-Alyani AM, Alseghayer MS, Almaneaa AM: Human salmonellosis: A continuous global threat in the farm-to-fork food safety continuum. *Foods*, 12 (9):1756, 2023. DOI: 10.3390/foods12091756
135. Huang WTK, Masselot P, Bou-Zeid E, Fatichi S, Paschalis A, Sun T, Gasparrini A, Manoli G: Economic valuation of temperature-related mortality attributed to urban heat islands in European cities. *Nat Commun*, 14 (1):7438, 2023. DOI: 10.1038/s41467-023-43135-z



136. Zizza A, Fallucca A, Guido M, Restivo V, Roveta M, Trucchi C: Foodborne infections and Salmonella: Current primary prevention tools and future perspectives. *Vaccines*, 13 (1):29, 2024. DOI: 10.3390/vaccines13010029
137. O'Connor L, McKeown P, Barrasa A, Garvey P: Epidemiology of *Campylobacter* infections in Ireland 2004-2016: What has changed? *Zoonoses Public Health*, 67 (4): 362-369, 2020. DOI: 10.1111/zph.12695
138. Namang BM, Joshua BI, Maryam M, Milton SN, Nyam LS, Ojonugwa AG, Davou GM, Abiola RM, Junaidu K, Antonnia L: Prevalence of *Salmonella* isolated in cattle, inva gene detection and antimicrobial susceptibility patterns of isolates. *J Anim Husband Dairy Sci*, 5 (1): 23-31, 2021. DOI: 10.22259/2637-5354.0501004
139. Ingle DJ, Ambrose RL, Baines SL, Duchene S, Gonçalves da Silva A, Lee DYJ, Jones M, Valcanis M, Taiaroa G, Ballard SA: Evolutionary dynamics of multidrug resistant *Salmonella enterica* serovar 4,[5], 12: i-in Australia. *Nat Commun*, 12 (1):4786, 2021. DOI: 10.1038/s41467-021-25073-w
140. Wigley P: *Salmonella* and the chicken: Reflections on salmonellosis and its control in the United Kingdom. *Poult Sci Manag*, 1 (1):1, 2024. DOI: 10.1186/s44364-024-00001-y
141. Farhat M, Khayi S, Berrada J, Mouahid M, Ameer N, El-Adawy H, Fellahi S: *Salmonella enterica* serovar Gallinarum biovars Pullorum and Gallinarum in poultry: Review of pathogenesis, antibiotic resistance, diagnosis and control in the genomic era. *Antibiotics*, 13 (1):23, 2023. DOI: 10.3390/antibiotics13010023
142. Nielsen TD, Kudahl AB, Østergaard S, Nielsen LR: Gross margin losses due to *Salmonella* Dublin infection in Danish dairy cattle herds estimated by simulation modelling. *Prev Vet Med*, 111 (1-2): 51-62, 2013. DOI: 10.1016/j.prevetmed.2013.03.011
143. Elbehiry A, Abalkhail A, Marzouk E, Elmanssury AE, Almuzaini AM, Alfheaid H, Alshahrani MT, Huraysh N, Ibrahim M, Alzaben F, Alanazi F, Alzaben M, Anagreyah SA, Bayameen AM, Draz A, Abu-Okail A: An overview of the public health challenges in diagnosing and controlling human foodborne pathogens. *Vaccines*, 11 (4):725, 2023. DOI: 10.3390/vaccines11040725
144. Medeiros I, Fernandez-Novo A, Astiz S, Simões J: Historical evolution of cattle management and herd health of dairy farms in OECD countries. *Vet Sci*, 9 (3):125, 2022. DOI: 10.3390/vetsci9030125
145. Falleti J, Orabona P, Municinò M, Castellaro G, Fusco G, Mansueto G: An update on myocarditis in forensic pathology. *Diagnostics*, 14 (7):760, 2024. DOI: 10.3390/diagnostics14070760
146. An K, Wu Z, Zhong C, Li S: Case report: Uncommon presentation of *Salmonella* Dublin infection as a large paravertebral abscess. *Front Med*, 10:1276360, 2023. DOI: 10.3389/fmed.2023.1276360
147. Canciu A, Tertis M, Hosu O, Cernat A, Cristea C, Graur F: Modern analytical techniques for detection of bacteria in surface and wastewaters. *Sustainability*, 13 (13):7229, 2021. DOI: 10.3390/su13137229
148. Vashisht V, Vashisht A, Mondal AK, Farmaha J, Alptekin A, Singh H, Ahluwalia P, Srinivas A, Kolhe R: Genomics for emerging pathogen identification and monitoring: Prospects and obstacles. *BioMedInformatics*, 3 (4): 1145-1177, 2023. DOI: 10.3390/biomedinformatics3040069
149. Artika IM, Dewi YP, Nainggolan IM, Siregar JE, Antonjaya U: Real-time polymerase chain reaction: Current techniques, applications, and role in COVID-19 diagnosis. *Genes*, 13 (12):2387, 2022. DOI: 10.3390/genes13122387
150. Elhalem Mohamed AA, Mady WH, Omar SE, Atteya LAF, Alkhateeb MA, Al-Doaiss AA, Saleh O, Alhazmi N, Al-Nazawi AM, Said D: Development of tagman real-time fluorescent quantitative PCR for rapid detection and differentiation between DHAV-1 and DHAV-3 in duck farming. *Pak Vet J*, 44 (2): 490-498, 2024. DOI: 10.29261/pakvetj/2024.181
151. Tozzo P, Delicati A, Zambello R, Caenazzo L: Chimerism monitoring techniques after hematopoietic stem cell transplantation: An overview of the last 15 years of innovations. *Diagnostics*, 11 (4):621, 2021. DOI: 10.3390/diagnostics11040621
152. Akçakavak G, Karataş Ö, Tuzcu N, Tuzcu M: Determination of apoptosis, necroptosis and autophagy markers by real-time PCR in naturally infected pneumonic pasteurellosis caused by *Pasteurella multocida* and *Mannheimia haemolytica* in cattle. *Pak Vet J*, 44 (2): 483-489, 2024. DOI: 10.29261/pakvetj/2024.177
153. Lee TCH, Chan PL, Tam NFY, Xu SJL, Lee FWF: Establish axenic cultures of armored and unarmored marine dinoflagellate species using density separation, antibacterial treatments and stepwise dilution selection. *Sci Rep*, 11 (1):202, 2021. DOI: 10.1038/s41598-020-80638-x
154. Costa-Ribeiro A, Lamas A, Prado M, Garrido-Maestu A: Evaluation of the novel mTA10 selective broth, MSB, for the co-enrichment and detection of *Salmonella* spp., *Escherichia coli* O157 and *Listeria monocytogenes* in ready-to-eat salad samples. *Foods*, 13 (1):63, 2023. DOI: 10.3390/foods13010063
155. Neyaz LA, Alghamdi HS, Alghashmari RM, Alswat SS, Almaghrabi RO, Bazaid FS, Albarakaty FM, Elbanna K, Abulreesh HH: A comprehensive review on the current status of culture media for routine standardized isolation of *Salmonella* and *Shigella* spp. from contaminated food. *J Umm Al-Qura Univ Appl Sci*, 2024:1-14, 2024. DOI: 10.1007/s43994-024-00205-2
156. Mutlaq S, Albiss B, Al-Nabulsi AA, Jaradat ZW, Olaimat AN, Khalifeh MS, Osaili T, Ayyash MM, Holley RA: Conductometric immunosensor for *Escherichia coli* O157: H7 detection based on polyaniline/ zinc oxide (PANI/ZnO) nanocomposite. *Polymers*, 13 (19):3288, 2021. DOI: 10.3390/polym13193288
157. Elghryani N, McAloon C, Mincher C, McOwan T, Waal TD: Comparison of the automated OvaCyte telenostic faecal analyser versus the McMaster and Mini-FLOTAC techniques in the estimation of helminth faecal gg counts in equine. *Animals (Basel)*, 2023 13 (24):3874, 2023. DOI: 10.3390/ani13243874
158. Neupane DP, Dulal HP, Song J: Enteric fever diagnosis: current challenges and future directions. *Pathogens*, 10 (4):410, 2021. DOI: 10.3390/pathogens10040410
159. Loderstädt U, Hagen RM, Hahn A, Frickmann H: New developments in pcr-based diagnostics for bacterial pathogens causing gastrointestinal infections - A narrative mini-review on challenges in the tropics. *Trop Med Infect Dis*, 6 (2):96, 2021. DOI: 10.3390/tropicalmed6020096
160. Alberfkani MI, Swar SO, Almutairi LA, Hasan HK, Ahmed AE, Khalid HM, Mero W: Molecular characterization and phylogenetic analysis of 18S rRNA, gp60 and HSP70 genes of *Cryptosporidium parvum* isolated from cattle owners and cattle using nested PCR. *Pak Vet J*, 44 (4): 1237-1242, 2024. DOI: 10.29261/pakvetj/2024.281
161. Cook N, D'Agostino M, Wood A, Scobie L: Real-time PCR-based methods for detection of hepatitis E virus in pork products: A critical review. *Microorganisms*, 10 (2):428, 2022. DOI: 10.3390/microorganisms10020428
162. de Barros Moura AC, Silva Filho E, Machado Barbosa E, Assunção Pereira WL: Comparative analysis of PCR, real-time PCR and LAMP techniques in the diagnosis of *Trypanosoma vivax* infection in naturally infected buffaloes and cattle in the Brazilian Amazon. *Pak Vet J*, 44 (1): 123-128, 2024. DOI: 10.29261/pakvetj/2024.128
163. Zhang G, Jiang H, Zhang G, Li P, Feng Y, Shen X, Ding J: Strain-level Identification of *Brucella melitensis* reference strain 63/9 using multiplex PCR method by targeting BMEA\_B0162 and BMEA\_A1238. *Pak Vet J*, 44 (1): 183-189, 2024. DOI: 10.29261/pakvetj/2024.135
164. Anumbe N, Saidy C, Harik R: A primer on the factories of the future. *Sensors*, 22 (15):5834, 2022. DOI: 10.3390/s22155834
165. Kong-Ngoen T, Santajit S, Tunyong W, Pumirat P, Sookrung N, Chaicumpa W, Indrawattana N: Antimicrobial resistance and virulence of non-typhoidal *Salmonella* from retail foods marketed in Bangkok, Thailand. *Foods*, 11 (5):661, 2022. DOI: 10.3390/foods11050661
166. Torreggiani C, Paladini C, Cannistrà M, Botti B, Prosperi A, Chiapponi C, Soliani L, Mescoli A, Luppi A: Managing a *Salmonella* Bredeney outbreak on an Italian dairy farm. *Animals*, 14 (19):2775, 2024. DOI: 10.3390/ani14192775
167. Ban E, Song EJ: Considerations and suggestions for the reliable analysis of miRNA in plasma using qRT-PCR. *Genes*, 13 (2):328, 2022. DOI: 10.3390/genes13020328
168. Mistry DA, Wang JY, Moeser M-E, Starkey T, Lee LYW: A systematic review of the sensitivity and specificity of lateral flow devices in the detection

of SARS-CoV-2. *BMC Infect Dis*, 21:828, 2021. DOI: 10.1186/s12879-021-06528-3

**169. Faustini G, Tucciarone CM, Franzo G, Donneschi A, Boniotti MB, Alborali GL, Drigo M:** Molecular survey on Porcine Parvoviruses (PPV1-7) and their association with major pathogens in reproductive failure outbreaks in Northern Italy. *Viruses*, 16 (1):157, 2024. DOI: 10.3390/v16010157

**170. Hanson KE, Caliendo AM, Arias CA, Englund JA, Hayden MK, Lee MJ, Loeb M, Patel R, Altayar O, El Alayli A:** Infectious Diseases Society of America guidelines on the diagnosis of COVID-19: Serologic testing (September 2020). *Clin Infect Dis*, 78 (7): e150-e169, 2024. DOI: 10.1093/cid/ciaa1343

**171. Tursunov K, Tokhtarova L, Kanayev D, Mustafina R, Tarlykov P, Mukantayev K:** Evaluation of an in-house ELISA for detection of antibodies against the Lumpy Skin Disease Virus in vaccinated cattle. *Int J Vet Sci*, 13 (2): 248-253, 2024. DOI: 10.47278/journal.ijvs/2023.089

**172. Nazir J, Manzoor T, Saleem A, Gani U, Bhat SS, Khan S, Haq Z, Jha P, Ahmad SM:** Combatting *Salmonella*: A focus on antimicrobial resistance and the need for effective vaccination. *BMC Infect Dis*, 25 (1):84, 2025. DOI: 10.1186/s12879-025-10478-5

**173. Williamson KM, Faddy H, Nicholson S, Stambos V, Hoad V, Butler M, Housen T, Merritt T, Durrheim DN:** A cross-sectional study of measles-specific antibody levels in Australian blood donors - Implications for measles post-elimination countries. *Vaccines*, 12 (7):818, 2024. DOI: 10.3390/vaccines12070818

**174. Bauer BU, Schwecht KM, Jahnke R, Matthiesen S, Ganter M, Knittler**

**MR:** Humoral and cellular immune responses in sheep following administration of different doses of an inactivated phase I vaccine against *Coxiella burnetii*. *Vaccine*, 41 (33): 4798-4807, 2023. DOI: 10.1016/j.vaccine.2023.06.061

**175. Haselbeck AH, Im J, Prifti K, Marks F, Holm M, Zellweger RM:** Serology as a tool to assess infectious disease landscapes and guide public health policy. *Pathogens*, 11 (7):732, 2022. DOI: 10.3390/pathogens11070732

**176. Um MM, Castonguay M-H, Arsenault J, Bergeron L, Fecteau G, Francoz D, Dufour S:** Accuracy of testing strategies using antibody-ELISA tests on repeated bulk tank milk samples and/or sera of individual animals for predicting herd status for *Salmonella* Dublin in dairy cattle. *Prev Vet Med*, 220:106048, 2023. DOI: 10.1016/j.prevetmed.2023.106048

**177. Pautienius A, Dudas G, Simkute E, Grigas J, Zakienė I, Paulauskas A, Armonaitė A, Zienius D, Slyzius E, Stankevicius A:** Bulk milk tank samples are suitable to assess circulation of tick-borne encephalitis virus in high endemic areas. *Viruses*, 13 (9):1772, 2021. DOI: 10.3390/v13091772

**178. Righi C, Iscaro C, Ferroni L, Rosati S, Pellegrini C, Nogarol C, Rossi E, Dettori A, Feliziani F, Petrini S:** Validation of a commercial indirect ELISA kit for the detection of Bovine alphaherpesvirus 1 (BoHV-1)-specific glycoprotein E antibodies in bulk milk samples of Dairy cows. *Vet Sci*, 9 (7):311, 2022. DOI: 10.3390/vetsci9070311

**179. García-Soto S, Linde J, Methner U:** Epidemiological analysis on the occurrence of *Salmonella enterica* subspecies enterica serovar Dublin in the German federal state Schleswig-Holstein using whole-genome sequencing. *Microorganisms*, 11 (1): 122, 2023. DOI: 10.3390/microorganisms11010122