

SHORT COMMUNICATION

Identification of a New MLST Subtype of *Cryptosporidium muris* from a Brown Rat (*Rattus norvegicus*) in China

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Abstract

Cryptosporidium muris is a zoonotic protozoan. Characterization of genetic differentiation can be exploited to trace the origins of the parasite. A *Cryptosporidium* isolate from brown rat was obtained by microscopy and genetically characterized. *C. muris* was identified with minor nucleotide differences. Further subtype analysis was performed using a multilocus sequence typing (MLST) tool. At each locus, the *C. muris* isolate was confirmed to be M12, M4, M1 and M2 subtype, representing as a new MLST subtype. Therefore, the results suggest that there was probably a new source of infection for *C. muris* in the study region.

Keywords: *Cryptosporidium muris*, 18S rRNA, MLST, Subtype, Brown rat

Çin'de Kahverengi Sıçanda (*Rattus norvegicus*) *Cryptosporidium muris*'in Yeni Bir MLST Alt Tipinin Tanımlanması

Öz

Cryptosporidium muris, zoonotik bir protozondur. Genetik farklılaşmanın karakterizasyonu parazitin kökenini izlemek için kullanılabilir. *Cryptosporidium* izolatu kahverengi bir sıçandan mikroskopi ile elde edildi ve genetik olarak karakterize edildi. Küçük nükleotid farklılıklarla *C. muris* olarak tanımlandı. İleri identifikasyonu, çoklu lokus dizilim analizi (MLST) ile gerçekleştirildi. Her lokusta, *C. muris* izolatının yeni bir MLST alt tipini temsil eden M12, M4, M1 ve M2 alt tipi tanımlandı. Bu nedenle bu sonuçlar, çalışma bölgesinde *C. muris* için muhtemelen yeni bir enfeksiyon kaynağı olduğunu göstermektedir.

Anahtar sözcükler: *Cryptosporidium muris*, 18S rRNA, MLST, Alt Tip, Kahverengi sıçan

INTRODUCTION

Cryptosporidium is a ubiquitous zoonotic protozoan parasite that can infect a variety of vertebrate animals. Depending on the immunological status of hosts, the clinical symptoms of cryptosporidiosis commonly range from acute to chronic diarrhea. Epidemiologic studies indicate that multiple routes of transmission are likely, including direct, zoonotic, foodborne and waterborne transmission may occur in different areas ^[1,2]. To date, at least 45 valid species and a similar number of genotypes have been described based on molecular characteristics, revealing an extensive genetic diversity ^[3]. Among those, *C. muris*

as gastric protozoa has a wide range of host-adaptations and is generally detected in various rodents ^[4,5]. Other animals including deer, Tasmanian devil, pig, camel, dog, cat, monkey, and bird can also be infected with *C. muris* at low frequency ^[6-10]. In addition, a few cases of human cryptosporidiosis have been attributed to *C. muris* in recent years, suggesting the potential for zoonotic transmission ^[11,12].

Numerous types of molecular diagnostic tools have been used to differentiate *Cryptosporidium* in host and environmental samples at the genotype and subtype levels to trace the likely source of infection ^[6]. Among those,

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a multilocus sequence typing (MLST) tool was developed for *C. muris* by employing microsatellite and minisatellite markers with simple tandem repeats, which showed the capacity to distinguish various subtypes^[13]. Currently, 14 MLST subtypes have been identified in diverse hosts and geographical regions^[13,14]. Additionally, the MLST tool is also helpful for molecular epidemiology and population genetic studies^[14].

Reports on *C. muris* infecting brown rats are limited, and subtyping *C. muris* was not conducted in those studies^[15]. To our knowledge, there are no reports on brown rats being infected with *C. muris* in China^[16]. In the present study, one *C. muris* isolate (BN01) from brown rat was genetically characterized by analysis of three genotyping and four MLST-subtyping genes for the purpose of further tracking of infection sources.

MATERIAL AND METHODS

Ethics Approval

This study was performed according to the recommendations of the Guide for the Care and Use of Laboratory Animals of the Ministry of Health, China. The research protocol was reviewed and approved by the Research Ethics Committee of the Sichuan Agricultural University.

Cryptosporidium muris Isolate

One brown rat was trapped in the suburbs of Ya'an city and kept in a sterilized mouse cage. Approximately 2-5 g of feces were collected for five consecutive days. The feces were dissolved into 50 mL double-distilled water via stirring. The Sheather's sugar flotation method was used for examining *Cryptosporidium* oocysts by microscopy at x400 magnification as previously described^[17]. The

specimen was found to be positive. Then, the concentrates containing oocysts were separated to 2.5% potassium dichromate solution and stored at 4°C.

DNA Extraction and PCR Amplification

Oocysts were rinsed twice in double-distilled water and pelleted at 5.000xg centrifugation to remove potassium dichromate. Genetic DNA was extracted using the E.Z.N.A.® Stool DNA Kit (OMEGA Biotek Inc D4015-02, USA) according to the manufacturer's protocol. The extracted DNA was stored at -20°C until the time of PCR analysis.

The nested-PCR was used to amplify the partial 18S rRNA, COWP and CpA135 genes to genotype identification. The primers and amplification conditions were adopted as previously described^[17,18]. Premix Ex Taq™ (Takara, Dalian, China) was used for PCR amplification. The secondary PCR products at each genotyping locus were resolved using 1% agarose gel electrophoresis with ethidium bromide staining, and were identified by DNA sequencing (Sangon Biotech, Shanghai, China). Sequence accuracy was confirmed using bidirectional sequencing, and a new PCR product was re-sequenced if necessary.

MLST Subtype

Analysis using the MLST incorporated MS1 (coding for hypothetical protein), MS2 (coding for 90 kDa heat shock protein), MS3 (coding for hypothetical protein), and MS16 (coding for leucine rich repeat family protein) in order to subtype the *C. muris* isolated from a brown rat. Primers and amplification conditions were in accordance with those developed previously (Table 1)^[13]. PrimeSTAR® DNA Polymerase (TaKaRa, Dalian, China) was used to ensure high sequence fidelity. To neutralize PCR inhibitors, 400 ng/mL of non-acetylated bovine serum albumin (TaKaRa, Dalian,

Table 1. PCR primers for MLST subtype used in this study

Genes	Primer	Sequence (5'-3')	Tm	Length	Tandem Repeat Sequence
MS1	F1	ACCATCTAGAGATAACGAGCGA	55°C	~550 bp	GAACGAGATAGG
	R1	GAATCAGAAGATGAGCGACAA			
	F2	CGTGATAGTGGGTATGAATTGGACA	55°C		
	R2	CGACTGCGATACTCACGTCCT			
MS2	F1	TTGCAACTGTACCTAAATTAGTA	52°C	~460 bp	CCATATCCC
	R1	GTGAGACTTCTGGGGTCCTGA			
	F2	TCATGACGCGTCATACCAACA	55°C		
	R2	ACTTAGACAGTTCTATGCTGA			
MS3	F1	AACCAAGTGAATCACGAACTT	55°C	~540 bp	TGTTGG /GCTGCA
	R1	TCAAGTACAGCAGTCTATTGCTT			
	F2	GCAATATCTTCGACGATCCCA	55°C		
	R2	ATGGGAATAATTCTTCATCATCAA			
MS16	F1	GAAGAGGTCTGAAGTTAAGCTA	50°C	~600 bp	CTTCTTCAT
	R1	GACAATCATCTAAATCGTGT			
	F2	AAGTTTCATCTAGGTACTACTAAGA	55°C		
	R2	CACTACCTAATCTCGTACTT			

F and R: the forward and reverse primers, respectively; 1 and 2: the primary and secondary PCRs, respectively

Table 2. Subtype of *C. muris* brown rat isolate in this study and other representative *C. muris* isolates at four MLST loci

No.	Specimen	Host	Source	MLST Subtype				Reference
				MS1	MS2	MS3	MS16	
1	RN66	Brown rat	Japan	M5	M4	M1	M5	[13]
2	6853	Human	Lima, Peru	M1	M2	M4	M5	[13]
3	7511	Bactrian camel	Czech Republic	M1	M1	M4	M5	[13]
4	14907	Rat snake	Czech Republic	M2	M2	M3	M5	[13]
5	13469	Camel via mice	Egypt	M5	M4	M2	M3	[13]
6	OH1	Ostrich	Henan, China	M5	M4	M6	M4	[14]
7	14714	Human	Nairobi, Kenya	M6	M4	M1	M2	[13]
8	14906	Lab mouse	Czech Republic	M6	M4	M2	M4	[13]
9	7379	Domestic mouse	Czech Republic	M7	M4	M2	M1	[13]
10	1666	Chipmunk	Czech Republic	M7	M4	M1	M5	[13]
11	7512	Mountain goat	Czech Republic	M8	M4	M2	M4	[13]
12	7380	Mara	Czech Republic	M9	M4	M1	M1	[13]
13	14243	SCID mice	Czech Republic	M10	M5	M5	M6	[13]
14	MC4	Hamster	Henan, China	M11	M4	M6	M1	[14]
15	BN01	Brown rat	Sichuan, China	M12	M4	M1	M2	This study

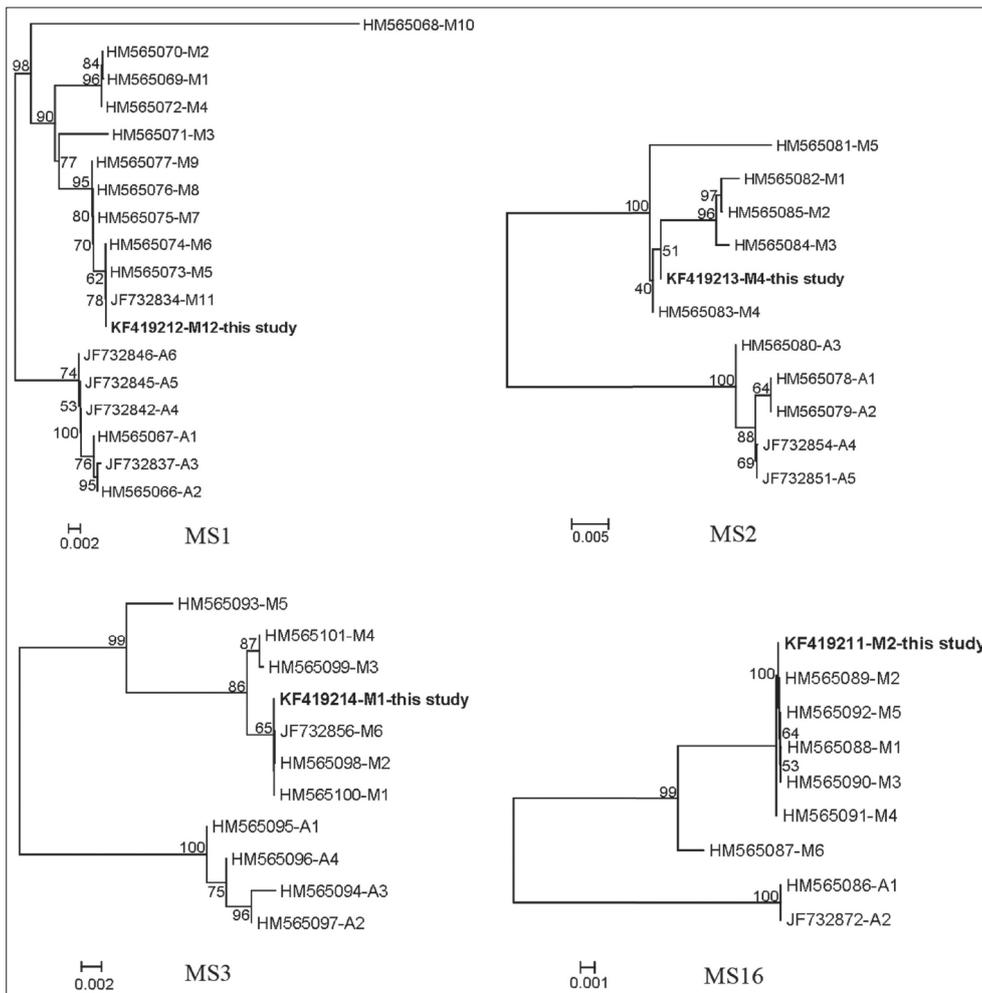


Fig 2. Phylogenetic relationships of *C. muris* and *C. andersoni* subtypes based on MS1, MS2, MS3, and MS16 sequences as assessed by a neighbor-joining analysis of the nucleotide sequences. Distances were calculated by the Kimura two-parameter model. Bootstrap values above 50% are shown using 1,000 pseudoreplicates. Bar = substitutions/site

sets [13]. The *C. muris* analyzed here was considered to be a new haplotype, M12, at MS1 locus and identified as MLST subtype M12, M4, M1 and M2. This characterization was supported by the phylogenetic relationship of each locus (Table 2, Fig. 2).

DISCUSSION

Previously, a total of 11, 5, 6 and 6 haplotypes comprising 14 MLST subtypes were identified for *C. muris* subtype loci (Table.1; Fig. 2). However, the current subtype was distinct

from previous *C. muris*, especially isolate RN66 (subtype M5, M4, M1 and M5). Thus, the *C. muris* isolate in this study was evaluated as a novel MLST subtype.

Subtyping tools have proven useful in the tracing of sources of infection. Of those, the 60 kDa glycoprotein (gp60) gene is a frequently used subtype marker, and is extensively used for distinguishing intestinal *Cryptosporidium* [6,19,20]. In addition, the MLST tool with a high resolution could also accurately describe genetic diversity and perform subtype identification for *Cryptosporidium*, including *C. muris* [6,13,19]. Previous data indicated that genetic differences of *Cryptosporidium* were likely to be related to geographical restriction [6,19]. The presence of a new MLST subtype suggests a new infection source of *C. muris*. Moreover, subtype differences of *C. muris* were observed in different animal hosts. For example, the MLST subtypes from ostriches and other hosts are unique and distinct from each other [13,14]. Thus, the new subtype in this study may be indicative of host-specificity.

Cryptosporidium muris infection in a brown rat was first reported in China, and a new MLST subtype was identified. However, limited information about *C. muris* infection in current areas has been reported. Therefore, there was a need of surveying cryptosporidiosis in extensive areas and hosts to trace the infection sources and characterize the transmission dynamics of *C. muris*.

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AUTHOR CONTRIBUTIONS

Conceived and designed the experiments: X. LIU, Z. ZHONG. Performed the experiments: L. DENG, F. YANG, Z. AN. Analyzed the data: X. LIU, Z. ZHONG, Z. ZHOU. Wrote the paper: X. LIU, L. DENG.

DECLARATION OF CONFLICT OF INTEREST

The authors declare that they have no competing interests.

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