

Effects of Pasteurization and Storage on Stability of Aflatoxin M₁ in Yogurt ^[1]

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Summary

In present study yogurt was produced from cow's milk contaminated artificially with aflatoxin M₁ (AFM₁) at two different levels, 1.5 and 3.5 µg kg⁻¹ (ppb), and the effects of pasteurization and storage on the AFM₁ contents were investigated. It was found statistically important that pasteurization at 95°C for 5 min caused losses of AFM₁ in milk contaminated with 1.5 and 3.5 µg kg⁻¹ AFM₁ about 18% and 16% , respectively (P<0.01). After yogurt production, total AFM₁ levels of yogurt samples produced by 1.5 and 3.5 µg kg⁻¹ AFM₁ contaminated milk samples were decreased to 36.5 and 34.6%, respectively. After the 2-week refrigerated storage at 4°C, AFM₁ content of yogurts produced from 1.5 and 3.5 µg kg⁻¹ AFM₁ contaminated milks decreased by 6.5 and 9.0%, respectively.

Keywords: Aflatoxin M₁, Milk, Stability, Yogurt, Storage

Yoğurttaki Aflatoxin M₁ Stabilitesi Üzerine Pastörizasyon ve Depolamanın Etkileri

Özet

Bu çalışmada aflatoxin M₁ (AFM₁) ile iki farklı düzeyde, 1.5 ve 3.5 µg kg⁻¹ (ppb), doğal olarak kontamine olmuş inek sütünden yoğurt üretilmiş ve AFM₁ içerikleri üzerine pastörizasyon ve depolamanın etkileri araştırılmıştır. 95°C'de 5 dakika pastörizasyonun, 1.5 ve 3.5 µg kg⁻¹ AFM₁ ile kontamine sütte AFM₁'de sırasıyla %18 ve %16 oranında istatistiksel açıdan önemli bulunan kayba neden olduğu bulunmuştur (P<0.01). Yoğurt üretiminden sonra, 1.5 ve 3.5 µg kg⁻¹ AFM₁ ile kontamine süttten üretilen yoğurt örneklerinin toplam AFM₁ düzeylerinde sırasıyla %36.5 ve %34.6 oranlarında düşüş olmuştur. Buzdolabında 4°C'de 2 haftalık depolamanın ardından 1.5 ve 3.5 µg kg⁻¹ AFM₁ ile kontamine süttten üretilen yoğurt örneklerinin toplam AFM₁ içerikleri sırasıyla %6.5 ve %9.0 oranında azalmıştır.

Anahtar sözcükler: Aflatoxin M₁, Süt, Stabilite, Yoğurt, Depolama

INTRODUCTION

Aflatoxins are highly toxic, mutagenic, teratogenic and carcinogenic compounds that have been implicated as causative agent in human hepatic and extrahepatic carcinogenesis ^{1,2}. AFM₁ found in animal milk is a major metabolite of aflatoxin B₁ (AFB₁) produced by *Aspergillus flavus* and *Aspergillus parasiticus* which contaminate agricultural commodities and feeds ³. A direct relationship has been observed between the amount of AFM₁ in milk and AFB₁ consumption via feedstuffs. The conservation rate of ingested AFB₁ into AFM₁ is highly variable, ranging from 0.3% to 6.2% ¹⁻³.

Agency for Research on Cancer, IARC), has now moved to Group 1 carcinogenic agent by IARC ⁴. Therefore, the presence of AFM₁ in milk and milk products is considered to be undesirable ^{5,6}. The occurrence of AFM₁ in milk and milk products is one of the most serious problems for food safety because of its importance as a foodstuff for adults and especially children ⁵⁻⁷.

It is of interest to know how the processes common to the dairy industry influence the AFM₁ content of milk and milk products. There are some contradictions in literature relating this issue. Some studies indicate that pasteurization

AFM₁, initially classified as a Group 2B agent (International



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and sterilization treatments are not reduce the content of AFM₁ in milk^{5,6,8}. However, some reports show that content of AFM₁ are degraded depending on time and temperature combination of heat treatment applied^{9,10}. Similarly, Fallah⁴ reported that AFM₁ is relatively stable during processing and storage of various dairy products.

The objective of the present work was to determine the stability of AFM₁ during production and storage of yogurt. For that purpose, the milk used for yogurt production was artificially contaminated with AFM₁ at the level of 1.5 and 3.5 µg kg⁻¹ and two batches of yogurt containing AFM₁ at two different levels were obtained. Effects of process stages such as heat treatment, incubation and cold storage on AFM₁ stability were investigated.

MATERIAL and METHODS

Production of Yogurt Samples Contaminated with AFM₁

Whole cow milk (6 kg) obtained from the Ankara University Agricultural Faculty dairy farm was equally divided into four portions. Two of 1.5 kg portions were spiked with standard AFM₁ (Sigma Chemical Co. St. Louis, USA) at the levels of 1.5 µg kg⁻¹ while other two 1.5 kg portions were spiked with 3.5 µg kg⁻¹ of AFM₁. Milk was sampled for AFM₁ analysis before and after the spiking procedure. Contaminated milks were pasteurized at 95°C for 5 min, cooled to 4°C and then 2% of mixed culture (1:1 ratio of *Streptococcus thermophilus* and *Lactobacillus delbrueckii* subsp. *bulgaricus*, YC 380, Direct Vat Set type, Chr. Hansen A/S. Horlom, Denmark) were added. After the shaking to distribute culture evenly each 1.5 kg milk portions were fermented at 43°C until 4.6 of pH. Following incubation process, yogurt samples were cooled to 4°C. With this production method four yogurt samples containing AFM₁ at two different concentrations (1.5 and 3.5 µg kg⁻¹) were produced. Two of them (contaminated with 1.5 and 3.5 µg kg⁻¹ of AFM₁) were used for analysis while other two samples were stored at 4°C for two weeks.

Analysis of Milk and Yogurt Samples for AFM₁

Milk samples (100 mL) were warmed to 37°C under gentle stirring for better fat dissolution. Samples were subsequently centrifuged at 4.000 rpm for 10 min and the upper thin fat layer was discarded (if any). The bottom skimmed milk layer was filtered through Whatman 4 paper (Brendford, UK) and 50 mL filtrate was collected for analysis.

Yogurt samples were prepared according to Mayes and MacDonalds¹². Briefly, yogurt (40 g), Celite-545 (10 g), chloroform (150 mL) and saturated sodium chloride solution (2 mL) were mixed by Ultra Turrax (Junke and Kunkel, GmbH, Germany) at medium speed for 2 to 3 min. The slurry was filtered through Whatman 4 filter paper. Filtrate was evaporated to dryness under vacuum at 60°C using rotary evaporator (Heidolph Rotary Evaporator vv-2000, Germany). Two milliliters of methanol and 60 mL of phosphate-buffered

saline (900 mL water, 8 g NaCl, 1.16 g Na₂HPO₄ mixed, and pH was adjusted to 7.4) were added to the residue, and the mixture was placed into the separating funnel (500 mL). Then, hexane (100 mL) was added, and mixture was shaken vigorously and the layers were allowed to separate and a portion of the bottom (aqueous) layer (50 mL) was collected for analysis.

An aliquot (50 mL) of the filtrate from each sample (milk and yogurt) was passed through an immunoaffinity column (Vicam AFLA M₁, USA) containing specific monoclonal antibodies bound to a solid support material. As the sample passed through the column, the antibodies selectively bound with AFM₁ present and formed an antibody-antigen complex. Other components of the sample matrix were washed off the column with 10 mL of water. The toxin was slowly eluted from the column with 3 mL of acetonitrile, and final extract was evaporated to 300 µL under the nitrogen stream and finally was redissolved in the HPLC (High performance liquid chromatography) mobile phase to 3 mL again. The volume of injection into HPLC was 100 µL. AFM₁ chromatogram of milk sample contaminated artificially with AFM₁ at the level of 1.5 µg kg⁻¹ was given in Fig. 1.

A liquid chromatographic system (Model 1100, Hewlett Packard, USA) equipped with a Octadodecil silicagel C-18 (5µm, 250 x 4.6 mm i.d.) column (Hicrom, Reading, UK) was used for AFM₁ determination. The mobile phase consisted of acetonitrile-water (25:75, v/v) was delivered to the column at a rate of 1 mL min⁻¹. Under these conditions AFM₁ was eluted from the column at 10.5 min. Fluorescence detector (Model G1321A, Hewlett Packard, USA) was used with excitation wavelength of 360 nm and an emission wavelength of 430 nm.

Preparation of Calibration Solutions for AFM₁

Pure, AFM₁ standard (Sigma-Aldrich, USA) was dissolved in chloroform to prepare a stock solution. A series of calibration solutions were prepared using stock solution in the concentrations of 0.5, 1.0, 2.5, 5.0 and 7.5 AFM₁ ng mL⁻¹. Calibration curves were constructed by plotting the peak area for each calibration solution against the mass of AFM₁ injected. The

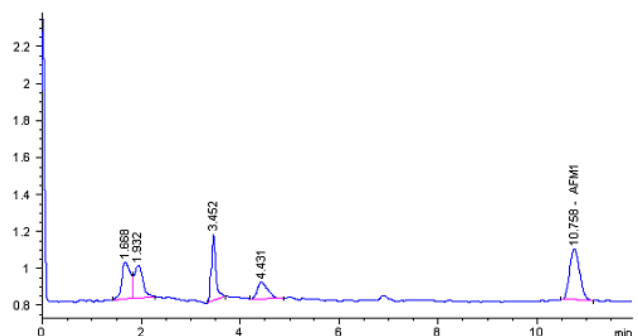


Fig 1. AFM₁ chromatogram of milk sample contaminated artificially with AFM₁ at the level of 1.5 µg kg⁻¹

Şekil 1. 1.5 µg kg⁻¹ düzeyinde AFM₁ ilave edilmiş süt örneğine ait kromatogram

detection limit was 0.01 $\mu\text{g kg}^{-1}$. Recoveries of AFM₁ from milk and yogurt were 99.60% and 94.42%, respectively. Analytical results were not corrected for recovery.

Statistical Analysis

Data were subjected to analysis of variance, whereas differences between means were tested for significance by Duncan's multiple range test in the general linear model of SPSS statistical package (SPSS version 10.0., SPSS Ltd. Working, UK). Differences between means were considered significant at $P < 0.01$.

RESULTS

Total AFM₁ content of raw milk contaminated with 1.5 and 3.5 $\mu\text{g kg}^{-1}$ of AFM₁ (RM-1.5 and RM-3.5), pasteurized milk and yogurt produced from those contaminated raw milks are shown in [Table 1](#). Total AFM₁ of RM-1.5 was 2.175 μg . After pasteurization at 95°C for 5 min, it decreased to 1.785 μg . In other words, heating of milk at 95°C for 5 min caused a significant ($P < 0.01$) loss of AFM₁ at the level of 17.93%. Similarly, a decrease in AFM₁ content was observed on subjecting the RM-3.5 to pasteurization at 95°C for 5 min. After pasteurization, the AFM₁ level of RM-3.5 decreased to 4.250 μg from those initially present in milk at the level of 5.063 μg . This 16.06% decrease was also found to be significant ($P < 0.01$).

Total AFM₁ of yogurt samples contaminated with 1.5 and 3.5 $\mu\text{g kg}^{-1}$ of AFM₁ (YOG-1.5 and YOG-3.5) produced from RM-1.5 and RM-3.5 were 1.381 $\mu\text{g kg}^{-1}$ and 3.312 $\mu\text{g kg}^{-1}$, respectively. This indicated a significant decrease ($P < 0.01$) from those initially presented in milk at the ratio of 36.51% for YOG-1.5 and 34.58% for YOG-3.5.

The concentration of AFM₁ in yogurt did not remain constant during the 2-week refrigerated storage period

Table 1. Total AFM₁ contents of samples contaminated with 1.5 and 3.5 $\mu\text{g kg}^{-1}$ AFM₁

Tablo 1. 1.5 ve 3.5 $\mu\text{g kg}^{-1}$ AFM₁ ile kontamine örneklerin toplam AFM₁ içerikleri

| Samples | Sample Amount | AFM ₁ ($\mu\text{g kg}^{-1}$) | Total AFM ₁ (μg) | Loss of Total AFM ₁ (%) ^c |
|---|---------------|--|--|---|
| Contaminated with 1.5 $\mu\text{g kg}^{-1}$ AFM₁ | | | | |
| RM | 1.500 kg | 1.450±0.010 | 2.175 ^a | - |
| PM | 1.500 kg | 1.190±0.027 | 1.785 ^b | 17.93 |
| YOG | 1.490 kg | 0.927±0.068 | 1.381 ^c | 36.51 |
| Contaminated with 3.5 $\mu\text{g kg}^{-1}$ AFM₁ | | | | |
| RM | 1.500 kg | 3.375±0.038 | 5.063 ^a | - |
| PM | 1.500 kg | 2.833±0.052 | 4.250 ^b | 16.06 |
| YOG | 1.490 kg | 2.223±0.018 | 3.312 ^c | 34.58 |

RM: raw milk, PM: pasteurized milk, YOG: yogurt, Sample weight was taken into consideration in the calculation of AFM₁ loss, ^{a,b}Differences between the averages of sample with different capital letters in same column were significant ($P < 0.01$), ^cLosses in percentage is calculated according to AFM₁ level of RM

Table 2. Effect of storage period on the AFM₁ levels of yogurts samples produced from 1.5 and 3.5 $\mu\text{g kg}^{-1}$ AFM₁ contaminated raw milk

Tablo 2. 1.5 ve 3.5 $\mu\text{g kg}^{-1}$ AFM₁ ile kontamine çiğ süttten üretilen yoğurt örneklerinin AFM₁ içerikleri üzerine depolamanın etkisi

| Samples | Sample Amount | AFM ₁ ($\mu\text{g kg}^{-1}$) | Total AFM ₁ (μg) | Loss of Total AFM ₁ (%) ^c |
|---|---------------|--|--|---|
| Contaminated with 1.5 $\mu\text{g kg}^{-1}$ AFM₁ | | | | |
| YOG | 1.490 kg | 0.927±0.068 | 1.381 ^a | 100 |
| YOG after 2-week of storage | 1.490 kg | 0.867±0.024 | 1.292 ^a | 6.45 |
| Contaminated with 3.5 $\mu\text{g kg}^{-1}$ AFM₁ | | | | |
| YOG | 1.490 kg | 2.223±0.018 | 3.312 ^b | 100 |
| YOG after 2-week of storage | 1.490 kg | 2.023±0.026 | 3.014 ^b | 9 |

YOG: Yogurt. Sample weight was taken into consideration in the calculation of AFM₁ loss, ^{a,b}Differences between the averages of sample with different capital letters in same column were significant ($P < 0.01$), ^cLosses in percentage is calculated according to AFM₁ level of RM

([Table 2](#)). Throughout the storage period 6.45% decrease was observed in the total AFM₁ content of YOG-1.5 which was found significant ($P > 0.01$). Similarly, when the total AFM₁ content of YOG-3.5 was considered as 100%, its AFM₁ level decreased to 91% after 2-week storage. This 9% decrease was not found significant ($P > 0.01$).

DISCUSSION

In this study, results showed that pasteurization process during yogurt production had an effect in the way of decrease of AFM₁. This decrease may be attributed to simultaneous interactions of whey proteins with casein, which may hinder the extraction of casein which is bounded with aflatoxin M₁ and consequently prevent heat destruction of the toxin. Wiseman et al.¹⁴ were also implied that changes in AFM₁ stability by heat treatment could be the result of changed extraction of the toxin after milk was heated and it might be showing an effect of extraction efficiency than a real decrease in AFM₁ content.

Results of milk samples are in agreement with those of El-Deep et al.⁹, who found 9.5% and 26% decreases in AFM₁ after heat treatment at 63°C for 30 min and at 121°C for 15 min, respectively, while Kiermeier and Mashaley¹³, reported that various heat-time treatments caused reductions in the AFM₁ concentrations of milk varying from 12 to 40%. Choudhary et al.¹⁰ have recently studied the effect of various heat treatments on AFM₁ stability of cow's milk and reported that sterilization of milk at 121°C for 15 min caused 12.21% degradation of AFM₁, whereas boiling decreased AFM₁ by 14.50%. They also concluded that destruction of AFM₁ depends on time and temperature combination of the heat treatment applied. However, some reports showed that aflatoxin was stable during heat-treatments such as pasteurization and sterilization^{8,14}.

Also, the results of this study were similar to obtained from Megalla and Hafez¹⁵ in a study on the effect of yogurt production on AFB₁. They reported complete removal of 800 µg kg⁻¹ of AFB₁ by its transformation into AFB_{2α} and concluded that the acidity of yogurt was responsible for this conversion. Rasic et al.¹⁶ reported that AFB₁ levels decreased from 90-97% after fermentation. Similarly, Hassanin¹⁷ mentioned a 17% decrease of AFM₁ in yogurt whereas Govaris et al.⁸ determined that AFM₁ levels in yogurt samples showed a significant decrease after fermentation. On the contrary, Blanco et al.¹⁸ reported that the yogurt production has no influence on AFM₁, while Munksgaard et al.¹⁹ showed a slight increase in concentration of AFM₁ in yogurt from those initially presented in raw milk.

The decrease of AFM₁ during yogurt production might be attributed to factors such as low pH, formation of organic acids or other fermentation by-products and presence of lactic acid bacteria²⁰. Changes in pH due to the acid development can affect the structure of milk proteins. Changes in casein structure may also affect the relationship between AFM₁ and casein²¹. In a study conducted by Govaris et al.²⁰, AFM₁ was significantly lower (P<0.01) in yogurts with pH 4.0 than in yogurts with pH 4.6. Production of larger amounts of lactic acid and other fermentation by-products along with the lower pH could cause this further decrease of AFM₁. Lactic acid and several other fermentation by-products such as volatile fatty acids, amino acids, peptides and aldehydes could also cause degradation of AFM₁ in yogurt²⁰⁻²². Rasic et al.¹⁶ showed a significant decrease of AFB₁ in milk acidified with lactic, citric or acetic acid. It was also found that lactic acid bacteria either viable or heat-killed could reduce the amount of AFB₁ and AFM₁²⁴. Govaris et al.²⁰ reported that the growth rate of *Streptococcus thermophilus* was affected by the higher level of AFM₁ but *Lactobacillus delbrueckii* subsp. *bulgaricus* was not affected by the presence of AFB₁ (600 - 1400 µg kg⁻¹) in milk during fermentation of yogurt.

Blanco et al.¹⁸ reported that AFM₁ in yogurt was stable until 7th day of storage but at the end of the 12th day, AFM₁ content decreased at the level of 9%. Hassanin¹⁷ also reported 40 and 59% of AFM₁ loss in yogurt samples after the 7 and 13 days of storage at 4°C, respectively. Whereas, Govaris et al.²¹ determined that storage of yogurt for 2 and 4 week resulted in 15.6 and 16.2% of AFM₁ lost, respectively.

Results indicated that the initial amount of AFM₁ in milk decreased after yogurt production. Some factors such as low pH, presence of lactic acid bacteria, organic acids and fermentation by products, could responsible for this decrease. Throughout the storage period of 2 weeks AFM₁ contents of yogurt samples were almost stable. Briefly, when the milk contaminated with AFM₁, toxin passes to milk products at different levels. Therefore, it should be mentioned that reduced recovery may not mean reduced toxicity. Avoiding contamination appears to be the only practical and economical way to ensure safety of milk products for human

consumption. Further studies are needed to understand the nature of the casein-AFM₁ association in order to completely explain the behavior of AFM₁ in dairy products.

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