


# Effects of Antioxidants Resveratrol, Catechin and Lipoic Acid and Carcinogen $KBrO_3$ on Lipophylic Vitamins and Cholesterol in Lung, Liver and Kidney of Wistar Rats <sup>[1]</sup>

Serhat KESER \*   
Orhan ERMAN \*\*

Okkes YILMAZ \*\*  
Sevgi IRTEGUN \*\*

Mehmet TUZCU \*\*

[1] This research was supported by the DPT-2002K120240 and DPT-2003K120440 and FUBAP 1357

\* Firat University, Faculty of Science, Chemistry Department, TR-23119 Elazig - TURKEY

\*\* Firat University, Faculty of Science, Biology Department, TR-23119 Elazig - TURKEY

Makale Kodu (Article Code): KVFD-2011-5022

## Summary

The aim of this research is to examine effects of antioxidants resveratrol, lipoic acid, catechin and carcinogen potassium bromate on the level of lipophylic vitamins and cholesterol in liver, lung and kidney tissue of Wistar albino rats. In this study, total 50 Wistar rats were randomly divided into five groups: 1. Control (C), 2.  $KBrO_3$  (K) (40 mg/kg two times per week), 3. Catechin (30 mg/kg four times per week) +  $KBrO_3$  (40 mg/kg two times per week) (Cat), 4. Lipoic Acid (30 mg/kg four times per week) +  $KBrO_3$  (40 mg/kg two times per week) (LA) and 5. Resveratrol (30 mg/kg four times per week) +  $KBrO_3$  (40 mg/kg two times per week) (R). All treatments were continued for 5 weeks, after which time each experimental rat was decapitated and tissues were collected and stored in  $-85^{\circ}C$  prior to biochemical analysis. In lung, liver and kidney, lipophylic vitamins and cholesterol were measured by HPLC. According to our results, while the cholesterol level was decreased in the K, Cat, LA and R groups, the  $\alpha$ -tocopherol level was decreased in K and R groups of the lung tissue.  $\alpha$ -tocopherol, cholesterol and retinol levels were increased in K, Cat, LA and R groups of the liver tissue. While the  $\delta$ -tocopherol, vitamin  $D_2$ , vitamin  $D_3$  and  $\alpha$ -tocopherol levels were decreased in K, Cat, LA and R groups, the cholesterol level was increased in Cat, LA and R groups of kidney tissue. In conclusion, our results indicated that the applications of resveratrol, lipoic acid, catechin and potassium bromate influenced cholesterol and lipophylic vitamins levels and these applications can be affected cholesterol biosynthesis in Wistar albino rats.

**Keywords:** Resveratrol, Potassium bromate, Catechin, Lipoic acid, Liver, Lung, Kidney

## Wistar Sıçanların Akciğer, Karaciğer ve Böbrek Dokularında Lipofilik Vitaminler ve Kolesterol Üzerinde Antioksidanlar Resveratrol, Kateşin ve Lipoik Asit ile Kanserojen $KBrO_3$ 'ün Etkileri

### Özet

Bu çalışmanın amacı kanserojen  $KBrO_3$ 'e karşı resveratrol, lipoik asit ve kateşin gibi antioksidanların Wistar ratların karaciğer, akciğer ve böbrek dokularında lipofilik vitaminler ve kolesterol üzerindeki etkilerini incelemektir. Bu çalışmada, toplam 50 Wistar sıçan rasgele beş gruba dağıtıldı: 1. Kontrol (C), 2.  $KBrO_3$  (K), (40 mg/kg dozunda haftada iki kez), 3. Kateşin (30 mg/kg dozunda haftada dört kez) +  $KBrO_3$  (40 mg/kg dozunda haftada iki kez) (Cat), 4. Lipoik Asit (30 mg/kg dozunda haftada dört kez) +  $KBrO_3$  (40 mg/kg dozunda haftada iki kez) (LA) and 5. Resveratrol (30 mg/kg dozunda haftada dört kez) +  $KBrO_3$  (40 mg/kg dozunda haftada iki kez) (R). Bütün uygulamalar 5 hafta boyunca sürdürüldü ve sonra her bir deney sıçanı dekapite edildi ve dokuları alınarak biyokimyasal analizlere kadar  $-85^{\circ}C$ 'de saklandı. Akciğer, karaciğer ve böbrek dokularında lipofilik vitaminler ve kolesterol seviyeleri HPLC cihazıyla belirlendi. Sonuçlarımıza göre, akciğer dokusunda kolesterol seviyesi K, Cat, LA ve R gruplarında,  $\alpha$ -tokoferol seviyesi ise K ve R gruplarında azalmıştır. Karaciğer dokusunda,  $\alpha$ -tokoferol, retinol ve kolesterol seviyeleri K, Cat, LA ve R gruplarında artmıştır. Böbrek dokusunda,  $\delta$ -tokoferol, vitamin  $D_2$ , vitamin  $D_3$  ve  $\alpha$ -tokoferol seviyeleri K, Cat, LA ve R gruplarında azalırken, kolesterol seviyesi Cat, LA ve R gruplarında artmıştır. Sonuç olarak, sonuçlarımız göstermiştir ki, resveratrol, lipoik asit, kateşin ve potasyum bromat uygulaması Wistar sıçanlarda lipofilik vitaminler ve kolesterol seviyelerini etkilemiştir ve bu uygulamalar sıçanlarda kolesterol biyosentezini etkilemiş olabilir.

**Anahtar sözcükler:** Resveratrol, Potasyum bromat, Kateşin, Lipoik asit, Karaciğer, Akciğer, Böbrek



İletişim (Correspondence)



+90 424 2370000/3734



serhatkeser@gmail.com

## INTRODUCTION

Resveratrol is a phytoalexin, first isolated from roots of *Veratrum grandiflorum* O. Loes (white hellebore) and then from *Polygonum cuspidatum*, but remained in obscurity for almost 50 years<sup>1,2</sup>. It got into prominence in early nineties in the context of "French paradox"; the phenomena where in certain population of France (and Greece), in spite of regular consumption of high fat diet, gets much less heart diseases<sup>3</sup>. The apparent cardioprotection was attributed to the regular consumption of moderate doses of red wine rich in resveratrol<sup>4</sup>. Initially resveratrol was characterized by its anti-platelet aggregation properties<sup>5</sup> and thereafter other beneficial effects such as vasorelaxation, antioxidant functions, etc., became apparent<sup>6,7</sup>. In experimental animals, resveratrol is rapidly metabolized by the liver and its plasma half-life remains quite low with a concomitant decline in its concentrations in tissues like brain, lung, liver and kidney<sup>8</sup>.

$\alpha$ -Lipoic acid is a disulfide compound that functions as a coenzyme in pyruvate dehydrogenase and  $\alpha$ -ketoglutarate dehydrogenase mitochondrial reactions, leading to the production of cellular energy (ATP).  $\alpha$ -Lipoic acid and its reduced form, dihydrolipoic acid, reduce oxidative stress by scavenging a number of free radicals in both membrane and aqueous domains, by chelating transition metals in biological systems, by preventing membrane lipid peroxidation and protein damage through the redox regeneration of other antioxidants such as vitamins C and E, and by increasing intracellular glutathione<sup>9-12</sup>.

Catechin, commonly known as the primary polyphenolic compounds in green tea, has been of great interest for its multiple health-promoting properties, such as antioxidant, anti-obesity, hypolipidemic, and anti-carcinogenic activities<sup>13-18</sup>.

KBrO<sub>3</sub> has been classified as a genotoxic carcinogen based on positive results in the Ames test<sup>19</sup>, and chromosome aberration<sup>20</sup> and micronucleus tests<sup>21</sup>. It has the potential to induce 8-hydroxy-2'-deoxyguanosine (8-OHdG) formation both *in vitro* and *in vivo*<sup>22-25</sup>. 8-OHdG is the most abundant oxidative DNA adduct, can induce mutations such as GC to TA transversions upon replication by DNA polymerases<sup>26</sup>.

The aim of this study was to examine effects of antioxidants resveratrol, lipoic acid, catechin and carcinogen potassium bromate on the level of lipophylic vitamins and cholesterol in liver, lung and kidney tissue of Wistar albino rats.

## MATERIAL and METHODS

### Chemicals

Resveratrol, lipoic acid, catechin, methanol and acetonitrile were obtained from Sigma Chemical Co. (USA).

Isopropyl alcohol was obtained from Fluka BioChemica (Switzerland). Potassium bromate was obtained Merck (Germany).

### Animals and Treatment

The following experiments were approved by the Ethical Committee of Firat University for the care and use of laboratory animals. In this study, total 50 old female Wistar rats were used. The animals were housed in cages where they had *ad libitum* rat chow and water in an air-conditioned room with a 12-h light/12-h dark cycle, and were randomly divided into five groups; each group containing ten rats. The first group was used as a Control (C), the second group KBrO<sub>3</sub> (K), the third group Catechin + KBrO<sub>3</sub> (Cat), the fourth group Lipoic Acid + KBrO<sub>3</sub> (LA), and fifth group Resveratrol + KBrO<sub>3</sub> (R). Rats in the K, Cat, LA and R groups were injected intraperitoneally potassium bromate 40 mg/kg in the physiologic saline (0.9% NaCl) two times per week. Rats in the Cat group was injected intraperitoneally catechin 30 mg/kg, rats in the LA group was injected intraperitoneally lipoic acid 30 mg/kg, rats in the R group was injected intraperitoneally resveratrol 30 mg/kg in the physiologic saline four times per week. In addition, physiological saline was injected to C group rats. These treatments were continued for five weeks, after which time each experimental rat was decapitated and tissue samples were collected and stored in -85°C prior to biochemical analysis<sup>27</sup>.

### Determination of Lipid Soluble Vitamins in Tissue Samples

500 mg lung, liver and kidney tissue sample was homogenized in 3 mL acetonitrile/methanol/isopropyl alcohol (2:1:1, v/v/v) containing tubes and the samples were vortexed for 30 s and centrifuged at 6.000×g for 10 min at 4°C. Supernatants were transferred to autosampler vials of the HPLC instrument. For lipophylic vitamins, the mixture of acetonitrile/methanol (3:1, v/v) was used as the mobile phase and the elution was performed at a flow-rate of 1 mL/min. The temperature of column was kept at 40°C. Supelcosil™ LC 18 DB column (250 x 4.6 mm, 5  $\mu$ m; Sigma, USA) was used as the HPLC column and detection was performed at 320 nm for retinol (vitamin A), and 215 nm for  $\delta$ -tocopherol,  $\alpha$ -tocopherol,  $\alpha$ -tocopherol acetate, vitamin D<sub>2</sub>, D<sub>3</sub>, K<sub>1</sub>. Identification of the individual vitamins was performed by frequent comparison with authentic external standard mixtures analyzed under the same conditions. Quantification was carried out by external standardization using Class VP software. The results of analysis were expressed as  $\mu$ g/g<sup>28,29</sup>.

### Total Cholesterol Analysis in Tissue Samples

500 mg lung, liver and kidney tissue sample in 3 mL acetonitrile/isopropyl alcohol (70:30, v/v) containing tubes and the mixture were vortexed for 30 s and centrifuged at

6.000 g for 10 min at 4°C. Supernatants were transferred to autosampler vials of the HPLC instrument. Acetonitrile-isopropyl alcohol (70:30 v/v) was used as mobile phase at a flow rate of 1 mL/min<sup>30</sup>. Supelcosil LC 18™ DB column (250 x 4.6 mm, 5 µm) was used as the HPLC column. Detection was performed by UV at 202 nm and 40°C column oven<sup>31</sup>. Quantification was carried out by external standardization using Class VP software. The results were expressed as µg/g wet weight tissue.

### Statistical Analysis

The experimental results were reported as mean ± S.E. Statistical analysis was performed using SPSS 15.0 Soft-ware. Analysis of variance (ANOVA) and an LSD test were used to compare the experimental groups with the controls<sup>32-35</sup>.

## RESULTS

### In Lung Tissue

Retinol level was decreased ( $P < 0.01$ ) in Cat group, but its level was not different ( $P > 0.05$ ) between C group and the other groups.  $\delta$ -tocopherol (K, Cat and R groups),  $\alpha$ -tocopherol (all groups) levels were significantly decreased

( $P < 0.001$ ). Vitamin D<sub>2</sub> and  $\alpha$ -tocopherol acetate (K, Cat and LA groups) levels were significantly increased ( $P < 0.001$ ). Vitamin D<sub>3</sub> level was increased ( $P < 0.05$ ) in K and R groups, but its level was significantly decreased ( $P < 0.01$ ,  $P < 0.001$ , respectively) in Cat and LA groups. Vitamin K<sub>1</sub> level was significantly increased ( $P < 0.001$ ) in K group, but its level was significantly decreased in LA and R groups ( $P < 0.001$ ). Cholesterol level was significantly decreased ( $P < 0.05$ ,  $P < 0.001$ , respectively) in K and R groups, but its level was not different ( $P > 0.05$ ) between C and other groups (Table 1).

### In Liver Tissue

Retinol, cholesterol (all groups),  $\delta$ -tocopherol (K, LA and R groups),  $\alpha$ -tocopherol (K, Cat, LA and R groups),  $\alpha$ -tocopherol acetate (Cat, LA and R groups) levels were significantly increased ( $P < 0.001$ ). Vitamin D<sub>2</sub> (all groups), Vitamin D<sub>3</sub> (K, Cat and R groups) levels were significantly decreased ( $P < 0.001$ ). Vitamin K<sub>1</sub> level was significantly decreased ( $P < 0.001$ ) in R group, but its level was significantly increased ( $P < 0.001$ ) in K, LA and R groups (Table 2).

### In Kidney Tissue

Retinol level was decreased ( $P < 0.05$ ) in K and Cat groups, but its level was significantly increased ( $P < 0.001$ ,  $P < 0.01$ ,

**Table 1.** Effects of KBrO<sub>3</sub> and antioxidants on lipophylic vitamins and cholesterol levels in lung

**Tablo 1.** Akciğer dokusunda lipofilik vitaminler ve kolesterol seviyeleri üzerinde KBrO<sub>3</sub> ve antioksidanların etkileri

Biochemical Parameters (µg/g)	Control (C)	KBrO <sub>3</sub> (K)	Catechin+ KBrO <sub>3</sub> (Cat)	Lipoic Acid+ KBrO <sub>3</sub> (LA)	Resveratrol+ KBrO <sub>3</sub> (R)
Retinol	1.44±0.04 <sup>a</sup>	1.47±0.10 <sup>a</sup>	0.94±0.04 <sup>c</sup>	1.16±0.07 <sup>a</sup>	1.20±0.13 <sup>a</sup>
$\delta$ -tocopherol	21.25±1.17 <sup>a</sup>	13.31±0.70 <sup>d</sup>	4.46±0.09 <sup>d</sup>	10.47±0.05 <sup>d</sup>	19.49±0.73 <sup>a</sup>
Vitamin D <sub>2</sub>	1.52±0.12 <sup>a</sup>	3.61±0.12 <sup>d</sup>	3.74±0.15 <sup>d</sup>	3.15±0.15 <sup>d</sup>	1.94±0.13 <sup>a</sup>
Vitamin D <sub>3</sub>	13.47±0.43 <sup>a</sup>	15.08±0.64 <sup>b</sup>	10.84±0.37 <sup>c</sup>	10.54±0.06 <sup>d</sup>	15.40±0.47 <sup>b</sup>
$\alpha$ -tocopherol	18.81±0.60 <sup>a</sup>	16.69±0.54 <sup>c</sup>	9.97±0.23 <sup>d</sup>	6.00±0.40 <sup>d</sup>	11.53±0.47 <sup>d</sup>
$\alpha$ -tocopherol acetate	2.12±0.29 <sup>a</sup>	3.65±0.06 <sup>c</sup>	21.06±0.42 <sup>d</sup>	25.63±0.26 <sup>d</sup>	1.73±0.12 <sup>a</sup>
Vitamin K <sub>1</sub>	3.48±0.27 <sup>a</sup>	9.75±0.23 <sup>d</sup>	3.49±0.10 <sup>a</sup>	1.15±0.08 <sup>d</sup>	1.02±0.08 <sup>d</sup>
Cholesterol	975.38±20.83 <sup>a</sup>	913.36±24.67 <sup>b</sup>	936.58±7.64 <sup>a</sup>	976.85±20.69 <sup>a</sup>	761.00±9.07 <sup>d</sup>

a, b, c, and d letters indicate statistical significant between groups. a:  $P > 0.05$ , b:  $P < 0.05$ , c:  $P < 0.01$ , d:  $P < 0.001$

**Table 2.** Effects of KBrO<sub>3</sub> and antioxidants on lipophylic vitamins and cholesterol levels in liver

**Tablo 2.** Karaciğer dokusunda lipofilik vitaminler ve kolesterol seviyeleri üzerinde KBrO<sub>3</sub> ve antioksidanların etkileri

Biochemical Parameters (µg/g)	Control (C)	KBrO <sub>3</sub> (K)	Catechin+ KBrO <sub>3</sub> (Cat)	Lipoic Acid+ KBrO <sub>3</sub> (LA)	Resveratrol+ KBrO <sub>3</sub> (R)
Retinol	18.74±1.36 <sup>a</sup>	30.98±0.30 <sup>d</sup>	26.36±0.85 <sup>c</sup>	43.86±2.65 <sup>d</sup>	24.03±0.34 <sup>b</sup>
$\delta$ -tocopherol	15.40±0.21 <sup>a</sup>	24.33±0.55 <sup>d</sup>	14.23±0.47 <sup>a</sup>	45.80±0.54 <sup>d</sup>	19.45±0.15 <sup>d</sup>
Vitamin D <sub>2</sub>	10.27±0.21 <sup>a</sup>	7.68±0.29 <sup>d</sup>	4.91±0.16 <sup>d</sup>	8.44±0.12 <sup>d</sup>	6.88±0.23 <sup>d</sup>
Vitamin D <sub>3</sub>	99.28±1.22 <sup>a</sup>	84.09±2.17 <sup>d</sup>	78.93±1.47 <sup>d</sup>	98.52±0.57 <sup>a</sup>	73.49±0.41 <sup>d</sup>
$\alpha$ -tocopherol	168.23±8.13 <sup>a</sup>	567.73±12.50 <sup>d</sup>	480.48±16.51 <sup>d</sup>	466.16±0.75 <sup>d</sup>	498.79±0.56 <sup>d</sup>
$\alpha$ -tocopherol acetate	20.83±0.83 <sup>a</sup>	21.76±1.20 <sup>a</sup>	25.62±0.89 <sup>c</sup>	35.36±0.38 <sup>d</sup>	29.56±0.17 <sup>d</sup>
Vitamin K <sub>1</sub>	20.47±1.21 <sup>a</sup>	120.81±1.64 <sup>d</sup>	56.86±1.34 <sup>d</sup>	213.08±0.57 <sup>d</sup>	3.98±0.25 <sup>d</sup>
Cholesterol	45.00±1.44 <sup>a</sup>	300.42±8.16 <sup>d</sup>	232.54±7.64 <sup>d</sup>	141.87±0.42 <sup>d</sup>	284.42±0.72 <sup>d</sup>

a, b, c, and d letters indicate statistical significant between groups. a:  $P > 0.05$ , b:  $P < 0.05$ , c:  $P < 0.01$ , d:  $P < 0.001$

**Table 3.** Effects of KBrO<sub>3</sub> and antioxidants on lipophilic vitamins and cholesterol levels in kidney**Tablo 3.** Böbrek dokusunda lipofilik vitaminler ve kolesterol seviyeleri üzerinde KBrO<sub>3</sub> ve antioksidanların etkileri

Biochemical Parameters (µg/g)	Control (C)	KBrO <sub>3</sub> (K)	Catechin+ KBrO <sub>3</sub> (Cat)	Lipoic Acid+ KBrO <sub>3</sub> (LA)	Resveratrol+ KBrO <sub>3</sub> (R)
Retinol	20.87±0.05 <sup>a</sup>	30.47±0.05 <sup>a</sup>	40.90±1.00 <sup>d</sup>	106.89±0.91 <sup>d</sup>	63.42±0.83 <sup>d</sup>
δ-tocopherol	27.10±0.64 <sup>a</sup>	34.98±0.89 <sup>d</sup>	4.90±0.08 <sup>d</sup>	20.56±0.18 <sup>d</sup>	3.79±0.13 <sup>d</sup>
Vitamin D <sub>2</sub>	7.93±0.36 <sup>a</sup>	4.03±0.31 <sup>d</sup>	2.52±0.12 <sup>d</sup>	6.44±0.07 <sup>d</sup>	4.36±0.10 <sup>d</sup>
Vitamin D <sub>3</sub>	102.27±0.81 <sup>a</sup>	39.71±0.69 <sup>d</sup>	27.47±0.58 <sup>d</sup>	88.25±0.57 <sup>d</sup>	46.13±0.53 <sup>d</sup>
α-tocopherol	39.91±1.13 <sup>a</sup>	15.23±0.38 <sup>d</sup>	15.20±0.08 <sup>d</sup>	12.26±0.84 <sup>d</sup>	19.62±0.10 <sup>d</sup>
Vitamin K <sub>1</sub>	29.46±2.04 <sup>a</sup>	13.75±0.39 <sup>d</sup>	9.80±0.11 <sup>d</sup>	31.84±1.03 <sup>b</sup>	7.90±0.14 <sup>d</sup>
Cholesterol	634.32±18.25 <sup>a</sup>	637.07±23.07 <sup>a</sup>	750.42±7.34 <sup>d</sup>	844.14±8.10 <sup>d</sup>	771.95±8.86 <sup>d</sup>

a, b, c, and d letters indicate statistical significant between groups. a: P>0.05, b: P<0.05, c: P<0.01, d: P<0.001

respectively) in LA and R groups. Vitamin D<sub>2</sub> and D<sub>3</sub> (all groups), α-tocopherol (all groups) levels were significantly decreased (P<0.001). Vitamin K<sub>1</sub> level was significantly decreased (P<0.001) in K, Cat and R groups, but its level was increased (P<0.01) in LA group. δ-tocopherol level was significantly increased (P<0.001) in K group, but its level was decreased (P<0.001) in the other groups. Cholesterol level was significantly increased (P<0.001) in Cat, LA and R groups, but its level was not different (P>0.05) between the C and K groups (Table 3).

## DISCUSSION

In lung tissue, α-tocopherol and cholesterol levels were significantly decreased in the K and R groups. In the R group, the reduction of cholesterol levels can be caused by the cholesterol-lowering properties of resveratrol. Keser et al.<sup>36</sup> showed that resveratrol administration was decreased the cholesterol level in muscle of Wistar rats. Keser et al.<sup>37</sup> has showed that cholesterol level was decreased in the heart and brain tissue of rats which administered resveratrol and potassium bromate. In addition these studies, we think that a molecular relationship cholesterol and α-tocopherol reduction. Laden and Porter<sup>38</sup> reported that the possibility that the protective effect of resveratrol on the development of cardiovascular disease may be explained in part by the inhibition of endogenous cholesterol biosynthesis. Decreasing of cholesterol level in the R group may be explained by a decline the squalene monooxygenase enzyme activity. Squalene monooxygenase is an enzyme in the endoplasmic reticulum of eukaryotic cells, catalyzes the epoxidation of squalene across a C=C double bond to yield 2,3-oxidosqualene in the first oxidative step of cholesterol biosynthesis<sup>38</sup>. Regulation of sterol receptors occurs at the level of transcription, suggesting that α-tocopherol acts through specific receptors or tocopherol-responsive transcription factors<sup>39</sup>. α-tocopherol similarly up-regulates the expression of α-tocopherol transfer protein (α-TTP), and thus plays a role in its own intracellular processing<sup>40,41</sup>. These findings provide a link between vitamin E and the regulation of cholesterol synthesis that is independent of the

antioxidant effects of vitamin E. Supernatant protein factor (SPF) is a recently cloned member of a family of cytosolic lipid-binding proteins that includes Sec14p, α-tocopherol transfer protein, and cellular retinal-binding protein. SPF stimulates the conversion of squalene to lanosterol in the downstream pathway for cholesterol biosynthesis, and overexpression of cloned SPF in hepatoma cells increases cholesterol synthesis. The recent identification of the SPF as α-tocopherol-associated protein (TAP) has called into question its long-standing association with the cholesterol biosynthesis. Unexpectedly, the sequence of TAP is identical to that SPF. TAP binds α-tocopherol, but not other isomers of tocopherol, with high affinity; in the presence of α-tocopherol TAP translocates to the nucleus and activates reporter gene transcription<sup>42</sup>. TAP is a recently identified cytosolic protein thought to be involved in the intracellular distribution of α-tocopherol<sup>43</sup>.

Miura et al.<sup>44</sup> reported that hypocholesterolemic action of resveratrol is attributed, at least in part, to an increased excretion of neutral sterols and bile acids into feces. They have suggested that dietary resveratrol is hypolipidemic with a tendency for anti-tumor-growth and anti-metastasis effects in hepatoma-bearing rats.

In the liver and kidney tissues, the cholesterol level was significantly increased in the K, Cat, LA and R groups. Increasing of cholesterol level may be caused application of KBrO<sub>3</sub> in the all groups. Yilmaz et al.<sup>28</sup> showed that application of KBrO<sub>3</sub> was increased the cholesterol level in serum of Wistar rats. Retinol and α-tocopherol levels were significantly increased in the all groups in liver tissue. We think that KBrO<sub>3</sub> administration was caused increasing of these vitamin levels in the liver tissue. Because, KBrO<sub>3</sub> administration was performed in all groups which increasing of retinol and α-tocopherol levels.

It was observed that α-tocopherol level was significantly decreased in the all groups in the kidney tissue. This may be due to the application of potassium bromate. Because it was demonstrated to induce renal cell tumors in male and female F344 rats after oral administration for 2 years in the drinking water and usage of KBrO<sub>3</sub> as a food additive

is now limited, so that exposure of humans via food is very low <sup>45</sup>.

Retinol level was increased in all groups of liver and kidney tissues. Vitamin D<sub>2</sub> and D<sub>3</sub> levels were decreased in all groups of liver and kidney tissues. The reason of these vitamin levels different may be application of KBrO<sub>3</sub> in these tissues when compared to control group.

In conclusion, present results confirm that there can be a relationship between the decreasing of the cholesterol and  $\alpha$ -tocopherol levels in the lung tissue. It can be speculated that resveratrol affected cholesterol biosynthesis in Wistar albino rats. And it was observed that the formation of lipid peroxidation in the kidney of old Wistar rats by induced a prooxidant and carcinogen chemical (KBrO<sub>3</sub>) administration.

## REFERENCES

- Nonomura S, Kanagawa H, Makimoto A:** Chemical constituents of polygonaceous plants. I. Studies on the components of Ko-jo-kon (*Polygonum cuspidatum* Sieb. et Zucc.). *Yaku Zass*, 83, 988-990, 1963.
- Takaoka MJ:** Of the phenolic substances of white hellebore (*Veratrum grandiflorum* Loes. fil.). *J Fac Sci Hokk Imper Univ*, 7, 8-9, 1999.
- Richard JL:** Coronary risk factors The French paradox. *Arch Mal Coeur Vaiss*, 80, 17-21, 1987.
- Kopp P:** Resveratrol a phytoestrogen found in red wine a possible explanation for the conundrum of the 'French paradox'. *Eur J Endocrinol*, 138, 619-620, 1998.
- Pace-Asciak CR, Hahn S, Diamandis EP, Soleas G Goldberg DM:** The red wine phenolics trans-resveratrol and quercetin block human platelet aggregation and eicosanoid synthesis: Implications for protection against coronary heart disease. *Clin Chim Acta*, 235, 207-219, 1995.
- Fitzpatrick DF, Hirschfield SL, Coffey RG:** Endothelium-dependent vasorelaxing activity of wine and other grape products. *Am J Physiol*, 265, H774-778, 1993.
- Frankel EN, Waterhouse AL, Kinsella JE:** Inhibition of human LDL oxidation by resveratrol. *Lancet*, 341, 1103-1104, 1993.
- Asensi M, Medina I, Ortega A, Carretero J, Baño MC, Obrador E, Estrela JM:** Inhibition of cancer growth by resveratrol is related to its low bioavailability. *Free Radic Biol Med*, 33, 387-398, 2002.
- Packer L:**  $\alpha$ -Lipoic acid: A metabolic antioxidant which regulates NF-kappa B signal transduction and protects against oxidative injury. *Drug Metab Rev*, 30, 245-275, 1998.
- Evans JL, Goldfine ID:**  $\alpha$ -Lipoic acid: A multifunctional antioxidant that improves insulin sensitivity in patients with type 2 diabetes. *Diab Technol Therapeut*, 2, 401-413, 2000.
- Packer L, Witt EH, Tritschler HJ:**  $\alpha$ -Lipoic acid as a biological antioxidant. *Free Radic Biol Med*, 19, 227-250, 1995.
- Çiftçi H, Özkaya A, Dayangaç A, Ölçücü A, Çelik S, Şahin Z, Ateş S:** Effect of lipoic acid on the some elements in brain tissue of DMBA-induced Guinea Pigs. *Kafkas Univ Vet Fak Derg*, 15 (4): 569-573, 2009.
- Crespy V, Williamson G:** A review of the health effects of green tea catechins in *in vivo* animal models. *J Nutr*, 134, 3431S-3440S, 2004.
- Koo MWL, Cho CH:** Pharmacological effects of green tea on the gastrointestinal system. *Eur J Pharmacol*, 500, 177-185, 2004.
- Lin YL, Cheng CY, Lin YP, Lau YW, Juan IM, Lin JK:** Hypolipidemic effect of green tea leaves through induction of antioxidant and phase II enzymes including superoxide dismutase, catalase, and glutathione S-transferase in rats. *J Agric Food Chem*, 46, 1893-1899, 1998.
- Seeram NP, Henning SM, Niu Y, Lee R, Scheuller HS, Heber D:** Catechin and caffeine content of green tea dietary supplements and correlation with antioxidant capacity. *J Agric Food Chem*, 54, 1599-1603, 2006.
- Weisburger JH:** Eat to live, not live to eat. *Nutr*, 16, 767-773, 2000.
- Yokozawa T, Nakagawa T, Kitani K:** Antioxidative activity of green tea polyphenol in cholesterol-fed rats. *J Agric Food Chem*, 50, 3549-3552, 2002.
- Ishidate M, Sofuni T, Yoshikawa K, Hayashi M, Nohmi T, Sawada M, Matsuoka A:** Primary mutagenicity screening of food additives currently used in Japan. *Food Chem Toxicol*, 22, 623-636, 1984.
- Ishidate M, Yoshikawa K:** Chromosome aberration tests with Chinese hamster cells *in vitro* with and without metabolic activation-a comparative study on mutagens and carcinogens. *Arch Toxicol Suppl*, 4, 41-44, 1980.
- Hayashi M, Kishi M, Sofuni T, Ishidate M:** Micronucleus tests in mice on 39 food additives and eight miscellaneous chemicals. *Food Chem Toxicol*, 26, 487-500, 1988.
- Parsons JL, Chipman JK:** The role of glutathione in DNA damage by potassium bromate *in vitro*. *Mutagenesis*, 15, 311-316, 2000.
- Kasai H, Nishimura S, Kurokawa Y, Hayashi Y:** Oral administration of the renal carcinogen, potassium bromate, specifically produces 8-hydroxydeoxyguanosine in rat target organ DNA. *Carcinogenesis*, 8, 1959-1961, 1987.
- Cadenas S, Barja G:** Resveratrol, melatonin, vitamin E, and PBN protect against renal oxidative DNA damage induced by the kidney carcinogen KBrO<sub>3</sub>. *Free Radic Biol Med*, 26, 1531-1537, 1999.
- Ballmaier D, Epe B:** Oxidative DNA damage induced by potassium bromate under cell-free conditions and in mammalian cells. *Carcinogenesis*, 16, 335-342, 1995.
- Cheng KC, Cahill DS, Kasai H, Nishimura S, Loeb LA:** 8-Hydroxyguanine, an abundant form of oxidative DNA damage, causes G-T and A-C substitutions. *J Biol Chem*, 267, 166-172, 1992.
- Keser S, Yilmaz O, Tuzcu M:** Effects of resveratrol on fatty acid levels in serum and erythrocytes of rats administered potassium bromate. *Asian J Chem*, 22 (10): 7841-7849, 2010.
- Yilmaz O, Keser S, Tuzcu M, Cetintas B:** Resveratrol (trans-3,4',5-trihydroxystilbene) decreases lipid peroxidation level and protects antioxidant capacity in sera and erythrocytes of old female Wistar rats induced by the kidney carcinogen potassium bromate. *ETAP*, 24 (2): 79-85, 2007.
- Özkaya A, Çelik S, Yüce A, Şahin Z, Yilmaz Ö:** The effects of ellagic acid on some biochemical parameters in the liver of rats against oxidative stress induced by aluminum. *Kafkas Univ Vet Fak Derg*, 16 (2): 263-268, 2010.
- Bragagnolo N, Rodriguez-Amaya DB:** Comparison of the cholesterol content of Brazilian chicken and quail eggs. *J Food Comp Anal*, 16 (2): 147-153, 2003.
- Katsanidis E, Addis PB:** Novel HPLC analysis of tocopherols, and cholesterol in tissue. *Free Radic Biol Med*, 27 (11-12): 1137-1140, 1999.
- Ulkey MB, Aktaş A, Bozkurt HH:** The effect of *Trifolium pratense* L. (red clover) on rat testes. *Kafkas Univ Vet Fak Derg*, 14 (2): 151-155, 2008.
- Kısmalı G:** Effects of Coenzyme Q10 on blood biochemistry in rats. *Kafkas Univ Vet Fak Derg*, 15 (2): 191-194, 2009.
- Ergun G, Aktas S:** ANOVA modellerinde kareler toplamı yöntemlerinin karşılaştırılması. *Kafkas Univ Vet Fak Derg*, 15 (3): 481-484, 2009.
- Akbay TT, Yarat A, Sacan O, Yanardag R:** The effect of glurenorm (gliquidone) on Aorta in STZ induced diabetic rats. *Kafkas Univ Vet Fak Derg*, 17 (2): 235-238, 2011.
- Keser S, Yilmaz O, Tuzcu M, Erman O, Irtegun S:** The effects of catechin, lipoic acid, resveratrol and potassium bromate on fatty acid, lipophylic vitamins and cholesterol levels in muscle of Wistar rats. *J Chem Soc Pak*, 34 (1): 89-93, 2012.
- Keser S, Yilmaz O, Tuzcu M:** The effects of potassium bromate and resveratrol on cholesterol and vitamin E levels in heart, muscle and brain of Wistar rats. *JABS*, 4 (2): 40-45, 2010.

- 38. Laden BP, Porter TD:** Resveratrol inhibits human squalene mono-oxygenase, *Nutr Res*, 21, 747-753, 2001.
- 39. Azzi A, Breyer I, Feher M, Ricciarelli R, Stocker A, Zimmer S, Zingg J:** Nonantioxidant functions of alpha-tocopherol in smooth muscle cells. *J Nutrition*, 131, 378-381, 2001.
- 40. Fechner H, Schlame M, Guthmann F, Stevens PA, Rustow B:** alpha and delta-tocopherol induce expression of hepatic alpha-tocopherol transfer-protein mRNA. *Biochem J*, 331 (Pt 2), 577-581, 1998.
- 41. Kim HS, Arai H, Arita M, Sato Y, Ogihara T, Inoue K, Mino M, Tamai H:** Effect of alpha-tocopherol status on alpha-tocopherol transfer protein expression and its messenger RNA level in rat liver. *Free Rad Res*, 28, 87-92, 1998.
- 42. Porter TD:** Supernatant protein factor and tocopherol-associated protein: An unexpected link between cholesterol synthesis and vitamin E. *J Nutr Biochem*, 14, 3-6, 2003.
- 43. Zimmer S, Stocker A, Sarbolouki MN, Spycher SE, Sassoon J, Azzi A:** A novel human tocopherol-associated protein: Cloning, *in vitro* expression, and characterization. *J Biol Chem*, 275, 25672-25680, 2000.
- 44. Miura D, Miura Y, Yagasaki K:** Hypolipidemic action of dietary resveratrol, a phytoalexin in grapes and red wine, in hepatoma-bearing rats. *Life Sci*, 73 (11): 1393-1400, 2003.
- 45. Kurokawa Y, Hayashi Y, Maekawa A, Takahashi M, Kokubo T:** Induction of renal cell tumors in F-344 rats by oral administration of potassium bromate, a food additive. *Gann*, 73, 335-338, 1982.