

## The Molecular Characterization of *Arcanobacterium pyogenes* Strains Isolated from Samples of Sheep and Cattle

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### Summary

The purpose of this study was to determine the presence of virulence genes, perform typing for the characterization of *Arcanobacterium pyogenes* field strains, and investigate the correlation between clonal types, virulence genes and occurrence of disease. The isolates (n=51) used in this study were isolated from different sources (liver (n=4) of sheep; liver (n=25), lungs (n=5), broncho-alveolar lavage (BAL) fluid (n=3), milk (n=12) and suppurative tissue (n=1) of dairy cows and synovial fluid (n=1) of a calf). The presence of haemolytic activity in *A. pyogenes* isolates was determined using rabbit, sheep, cattle, chicken, dog and human erythrocytes. Also, the presence of any cytotoxic effect was investigated by growth in Vero cell cultures. Genomic DNA fingerprinting for clonal analysis was generated by BOX-PCR typing. Conventional PCR was used for the determination of the presence of eight *A. pyogenes* virulence factor genes, namely, *nanH* (encoding neuraminidase H), *nanP* (encoding neuraminidase P), *plo* (encoding pyolysin-PLO), *cbpA* (encoding collagen-binding protein A) and *fimA*, *fimC*, *fimE* and *fimG* (encoding the major fimbrial subunit of four different fimbriae). Furthermore, the correlation between clonal types, virulence factors and the occurrence of disease was investigated. The haemolysins of all strains had haemolytic effect on rabbit, sheep, cattle, chicken, dog and human erythrocytes. In addition, all strains were found to be cytotoxic to Vero cells. According to clonal analysis results, the *A. pyogenes* isolates were determined to belong to 12 different types. While all *A. pyogenes* strains were positive for the *plo* gene, the positivity rate was 62% for the *nanH* gene, 84% for the *nanP* gene, 58% for the *cbp* gene, 96% for the *fimA* gene, 66% for the *fimC* gene, 42% for the *fimE* gene and 10% for the *fimG* gene. It was determined that no correlation existed between the clonal types and virulence factors of *A. pyogenes* isolates and occurrence of disease.

**Keywords:** *Arcanobacterium pyogenes*, Clonal typing, Virulence genes, Molecular characterization

## Koyun ve Sığırlardan İzole Edilen *Arcanobacterium pyogenes* Suşlarının Moleküler Karakterizasyonu

### Özet

Bu çalışmanın amacı, *Arcanobacterium pyogenes* saha izolatlarının karakterizasyonu için virülens genlerinin varlığını belirlemek, tiplendirmek ve klonal tipleri, virülens genleri ve hastalık oluşumu arasındaki ilişkiyi incelemektir. Bu çalışmada, farklı örneklerden izole edilen *A. pyogenes* izolatu (51) kullanıldı: koyun (n=4) ve sığır (n=25) apselli karaciğerlerinden, süt ineklerine ait akciğerler (n=5), bronko-alveolar yıkantılardan (n=3), mastitisli sütlerden (n=12), suppuratif doku örneğinden (n=1) ve buzağı eklem sıvısından (n=1). Bütün *A. pyogenes* izolatlarında, tavşan, koyun, sığır, tavuk, köpek ve insan eritrositleri ile hemolitik aktivite varlığı belirlendi. Aynı zamanda, vero hücre kültüründe sitotoksik etkileri incelendi. Klonal analiz için genomik DNA parmak izi BOX-PCR tiplendirme ile yapıldı. Sekiz virülens faktör genlerinin varlığının değerlendirilmesi için klasik PCR kullanıldı: *nanH* (nöraminidaz H), *nanP* (nöraminidaz P), *plo* (pyolysin-PLO), *cbpA* (kollejen bağlayan protein A) ve *fimA*, *fimC*, *fimE*, *fimG* (4 farklı fimribanın önemli fimbrial yapıları). Aynı zamanda, klonal tipleri, virülens faktörleri ve hastalık varlığı arasında bir ilişki incelendi. Bütün suşların hemolizini, tavşan, koyun, sığır, tavuk, köpek ve insan eritrositlerine hemolitik etkisine sahipti. Ayrıca, bütün suşların vero hücresine sitotoksik olduğu bulundu. Klonal analiz sonuçlarına göre; *A. pyogenes* izolatlarının 12 farklı tipte olduğu belirlendi. Bütün suşlar *plo* geni pozitif iken, *nanH* geni %62, *nanP* geni %84, *cbp* geni %58, *fimA* geni %96, *fimC* geni %66, *fimE* geni %42 ve *fimG* geni %10'du. Klonal tipler, virülens faktörleri ve hastalık varlığı için *A. pyogenes* izolatları arasında bir ilişki bulunmadı.

**Anahtar sözcükler:** *Arcanobacterium pyogenes*, Klonal tiplendirme, Virülens gen, Moleküler karakterizasyon



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## INTRODUCTION

*Arcanobacterium pyogenes* is an important opportunistic pathogen of mucosal surfaces in livestock, including cattle, sheep and goats <sup>1</sup>, and is also responsible for suppurative infectious disease <sup>2</sup>. *A. pyogenes* causes mastitis, abortion, pyometra, arthritis, and orchitis in livestock and foot abscesses in poultry, and can be recovered as either pure or mixed cultures <sup>3-5</sup>. It is also a secondary pathogen frequently isolated from liver and kidney abscesses in cattle with or without *Fusobacterium necrophorum* <sup>6-8</sup>. As it has also been isolated from the rumen of cattle and stomach of pigs, it is considered that *A. pyogenes* could be a common bacterium of the digestive system in animals <sup>2</sup>. In addition to animals, it has been reported that *A. pyogenes* has also been isolated from cases of arthritis and subcutaneous abscesses in humans dealing with animals <sup>9,10</sup>.

*A. pyogenes* expresses several known and putative virulence factors, including pyolysin (*ply*), neuraminidase (*nanH* and *nanP*) and collagen-binding protein (*cbpA*), which may contribute to its pathogenicity <sup>11-16</sup>. Pyolysin (PLO), a haemolytic exotoxin expressed by *A. pyogenes*, is a member of the thiol-activated cytolysin family of bacterial toxins. These toxins can play a primary role in the pathogenesis of infections caused by various Gram-positive pathogens. In this context, it has been demonstrated that PLO acts as the most important virulence determinant in *A. pyogenes* infections <sup>15,17</sup>. PLO is produced by all strains of *A. pyogenes* <sup>17</sup>. The collagen-binding protein (*cbpA*) is required by the infectious agent to adhere to collagen-rich tissue <sup>15,16</sup>. In addition, fimbriae can be involved in the adhesion of the agent to host cells <sup>1</sup>.

In our previous study <sup>18</sup>, we isolated, biochemically identified and confirmed with PCR the pyolysin gene in *A. pyogenes* strains. Furthermore, the antibiotic susceptibility of *A. pyogenes* isolates was determined. Silva et al. <sup>19</sup> described the genomic characterization of *A. pyogenes* isolates based on the screening of eight known and putative virulence factors using conventional PCR and BOX-PCR typing. The present study was aimed at establishing a research model similar to that described by Silva et al. <sup>19</sup>.

The purpose of this study was to determine the virulence genes, describe the genomic characterization of *Arcanobacterium pyogenes* field strains by typing, and investigate the correlation between clonal types, virulence genes and persistence of disease.

## MATERIAL and METHODS

### Bacterial Isolates

The isolates (n=51) used in this study were recovered from different sources (liver of sheep; liver, lungs, broncho-alveolar lavage fluid, milk and suppurative tissue of dairy

cows; and synovial fluid of a calf). Twenty-nine of the isolates were obtained from liver abscesses of slaughtered feed-lot cattle (n=25) and sheep (n=4) at a slaughterhouse in Konya province. Twelve strains were isolated from the milk of dairy cows with clinical mastitis. The lung (n=5) and broncho-alveolar lavage fluid (n=3) isolates were recovered from dairy cows suffering from respiratory problems. One strain was isolated from the suppurative tissue of a dead dairy cow. One other isolate was recovered from the synovial fluid of a calf.

### Determination of the Presence of Haemolysin and Cytotoxic Effect

Each isolate was grown in brain-heart infusion medium at 37°C for 48 h and the supernatants were harvested by centrifugation at 6.000 rpm. In order to determine the presence of haemolytic activity, rabbit, sheep, cattle, chicken, dog and human erythrocytes were added to each supernatant and the samples were incubated at 37°C for 18-20 h. Furthermore, the supernatant of each isolate was added to Vero cell cultures, which were observed for a period of 48-72 h for the development of cytopathological effect.

### Genomic DNA Isolation

Genomic DNA was extracted from 48 h-old-cultures of *A. pyogenes* using a commercial DNA purification kit (Wizard<sup>®</sup> Promega, USA). The procedure was performed according to the manufacturer's instructions. Firstly, isolates of *A. pyogenes* were grown at 37°C for 48 h and harvested by centrifugation at 12.000g for 3 min. After washed 3 times in PBS, the pelleted bacterial cells were first treated with 60 µl of lysozyme (10 mg ml<sup>-1</sup>; Merck, Germany) for 1 h at 37°C. The preparations were analysed on 0.7% agarose gel and the quantity and quality of DNA were determined spectrophotometrically using a ND 2000 spectrophotometer (Nanodrop, Germany). The amount of DNA for each isolate was adjusted to the required concentration for the detection of virulence factor genes.

### Clonal Analysis

Genomic DNA fingerprinting was generated by PCR using the BOX-A1R (5'-CTACGGCAAGGCGACGCTGACG-3') primer <sup>19</sup>. Amplification reactions occurred in a Thermal Cycler (Eppendorf, Germany) in a 25 µl volume of reaction mixture containing a primer concentration of 2 mM, 100 ng of genomic DNA and 1 U of DNA Taq polymerase (Fermentas, EU). An initial denaturation step (95°C, 2 min) was followed by 34 cycles of denaturation (95°C, 1 min), annealing (53°C, 1 min) and extension (72°C, 5 min), with a single final extension cycle at 72°C for 10 min <sup>19</sup>. A PCR mixture without DNA was used as a negative control.

PCR products (5 µl) were electrophoresed in 1.5% agarose gel at 90 V for 3 h and stained with ethidium bromide. Band sizes were determined by comparison with

a standard DNA ladder (Fermentas, EU). DNA banding profiles were imaged using Infinity Capture Version 12.4 for Windows (Vilbert Lourmat, France). Intra-species relationships on the basis of the similarity of DNA band profiles were calculated using the Bio1D++ program (Vilbert Lourmat, France), according to the NEI and LI (1979) similarity coefficient, i.e.  $a=2nxy/(nx + ny)$  where  $nx$  and  $ny$  are the number of bands in lane "x" and "y", respectively, and  $nxy$  the number of shared bands between the two lanes. Dendrograms were produced from the similarity values in the matrix using the UPGMA (unweighted pair group match average) algorithm.

### Screening of Genes Encoding Known and Putative Virulence Factors

The PCR protocols were performed as described by Silva et al.<sup>19</sup>. Conventional PCR was used for the determination of the presence of eight *A. pyogenes* virulence factor genes: *nanH* (encoding neuraminidase H), *nanP* (encoding neuraminidase P), *plo* (encoding pyolysin-PLO), *cbpA* (encoding CbpA) (Jost and Billington<sup>1</sup>) and four fimbrial genes, *fimA*, *fimC*, *fimE* and *fimG* (encoding the major fimbrial subunit of four different fimbriae) (Table 1). Also, *plo* gene in our previous study (Hadimli et al.<sup>18</sup>) were determined by used different specific primers.

PCR reactions were carried out in a 25 µl reaction mixture containing 25 pmol of each primer, 0.1 mM of each deoxynucleotide triphosphate (Fermentas), 1xPCR buffer, 2 mM MgCl<sub>2</sub>, 100 ng of genomic DNA and 1 U of Taq DNA polymerase (Fermentas). Except for the annealing temperatures, the thermal cycling conditions were identical for all sets of primers: 3 min at 94°C followed by 35 cycles of denaturation (94°C for 1 min), annealing (1 min) and extension (72°C for 3 min) and a final step at 72°C

for 7 min. The annealing temperature was set between 55°C and 60°C. Positive controls for each virulens gene were used (Fig. 1). Also, A PCR mixture without DNA was used as a negative control.

The identity of PCR products was initially confirmed by DNA sequencing. Amplification products were separated by electrophoresis in 2.5% or 1% agarose gel (*plo*, *cbpA*, *nanH*, *nanP*, *fimA*, *fimC*, *fimE* and *fimG*), stained with ethidium bromide and the bands were visualized using Infinity-Capture Version 12.4 (Vilbert Lourmat, France). The result was considered to be positive if the amplification product was of the expected molecular size.

## RESULTS

Haemolysins of all *A. pyogenes* strains had haemolytic effect on rabbit, sheep, cattle, chicken, dog and human erythrocytes. Furthermore, all isolates produced β-haemolysis on blood agar base containing sheep blood. In addition, all strains were found to be cytotoxic to Vero cell cultures *in vitro*.

### Virulence Factor Genes

The results obtained for clonal types and virulence factor genes, as well as the number of isolates, and sources of samples are shown in Table 2. While all *A. pyogenes* strains were positive for the *plo* gene, the positivity rate was 62% for the *nanH* gene, 84% for the *nanP* gene, 58% for the *cbp* gene, 96% for the *fimA* gene, 66% for the *fimC* gene, 42% for the *fimE* gene, and 10% for the *fimG* gene.

### Clonal Type Analysis

It was determined that, the *A. pyogenes* strains were of

**Table 1.** PCR primers used to amplify eight *A. pyogenes* virulence factor genes<sup>19</sup>

**Table 1.** *A. pyogenes*'in 8 virülens faktör genini çoğaltmak için kullanılan PZR primerleri<sup>19</sup>

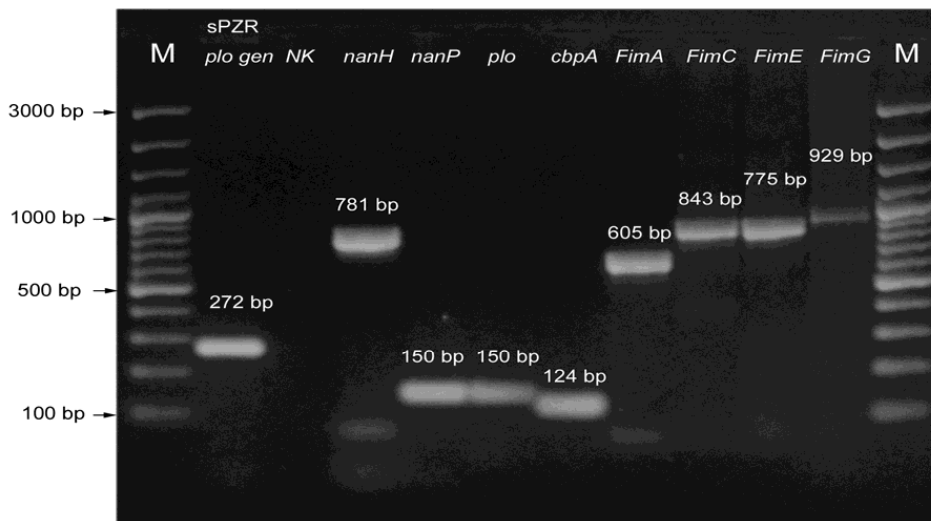
Target Gene	Primers	Annealing Temperature (°C)	PCR Product (bp)
<i>nanH</i>	Fw-cgctagtctgtagcgttgtaagt Rv-ccgaggagtttgactgactttgt	60	781
<i>nanP</i>	Fw-ttgagcgtacgcagctcttc Rv-ccacgaaatcggccttattg	60	150
<i>Plo</i>	Fw-tcatcaacaatcccacgaagag Rv-ttgctccagttgacgcttt	60	150
<i>cbpA</i>	Fw-gcagggttggtgaaagagtttact Rv-gcttgataacctcagaatttga	60	124
<i>fimA</i>	Fw-cactacgctcaccattcacaag Rv-gctgtaatccgctttgtctgtg	57	605
<i>fimC</i>	Fw-tgtcgaagggtgacgttcttcg Rv-caaggtcaccgagactgctgg	60	843
<i>fimE</i>	Fw-gcccaggaccgagagcgagggc Rv-gccttcacaataacagaacc	55	775
<i>fimG</i>	Fw-acgcttcagaaggtcaccagg Rv-atcttgatctgccccatgcg	57	929

**Table 2.** Correlation between clonal types, virulence factor genes and sources of samples**Tablo 2.** Klonal tipler, virülens factor genleri ve örneklerin kaynakları

BOX-PCR Type	No of Isolates	Source	Virulens Gene Factors							
			<i>plo</i> *	<i>nanH</i>	<i>nanP</i>	<i>cbpA</i>	<i>fimA</i>	<i>fimC</i>	<i>fimE</i>	<i>fimG</i>
I	4	BAL, CM, CL, SL	4/4	4/4	¾	4/4	4/4	3/4	2/4	0/4
II	7	BAL(2), CM(2), L(3)	7/7	5/7	4/7	6/7	6/7	5/7	5/7	0/7
III	1	CM	1/1	0/1	1/1	0/1	1/1	0/1	0/1	1/1
IV	6	CL (4), SL, L	6/6	6/6	5/6	6/6	6/6	6/6	6/6	0/6
V	1	CM	1/1	0/1	1/1	0/1	1/1	0/1	0/1	0/1
VI	8	CL (5), CM (2), L	8/8	5/8	7/8	5/8	7/8	6/8	3/8	0/8
VII	7	CL(4), CFJ, CM, ST	7/7	1/7	6/7	3/7	7/7	5/7	2/7	1/7
VIII	7	CL (6), CM	7/7	3/7	6/7	3/7	7/7	4/7	1/7	1/7
IX	1	CL	1/1	0/1	1/1	0/1	1/1	1/1	1/1	0/1
X	1	CL	1/1	0/1	1/1	1/1	1/1	1/1	0/1	0/1
XI	6	CL (3), CM, SL (2)	6/6	6/6	6/6	1/6	6/6	2/6	1/6	2/6
XII	2	CM (2)	2/2	2/2	2/2	1/2	2/2	1/2	1/2	1/2
<b>Total</b>	<b>51</b>		<b>51/51</b>	<b>32/51</b>	<b>43/51</b>	<b>30/51</b>	<b>49/51</b>	<b>34/51</b>	<b>22/51</b>	<b>6/51</b>

BAL: Broncho-alveolar lavage, CM: Cow's milk, CL: Cattle's liver, SL: Sheep liver, L: Cow's Lung, CFJ: Calf fluid of joint, ST: Suppurative tissue,

\* *plo* gen was determined using two different primers and all isolates given same result

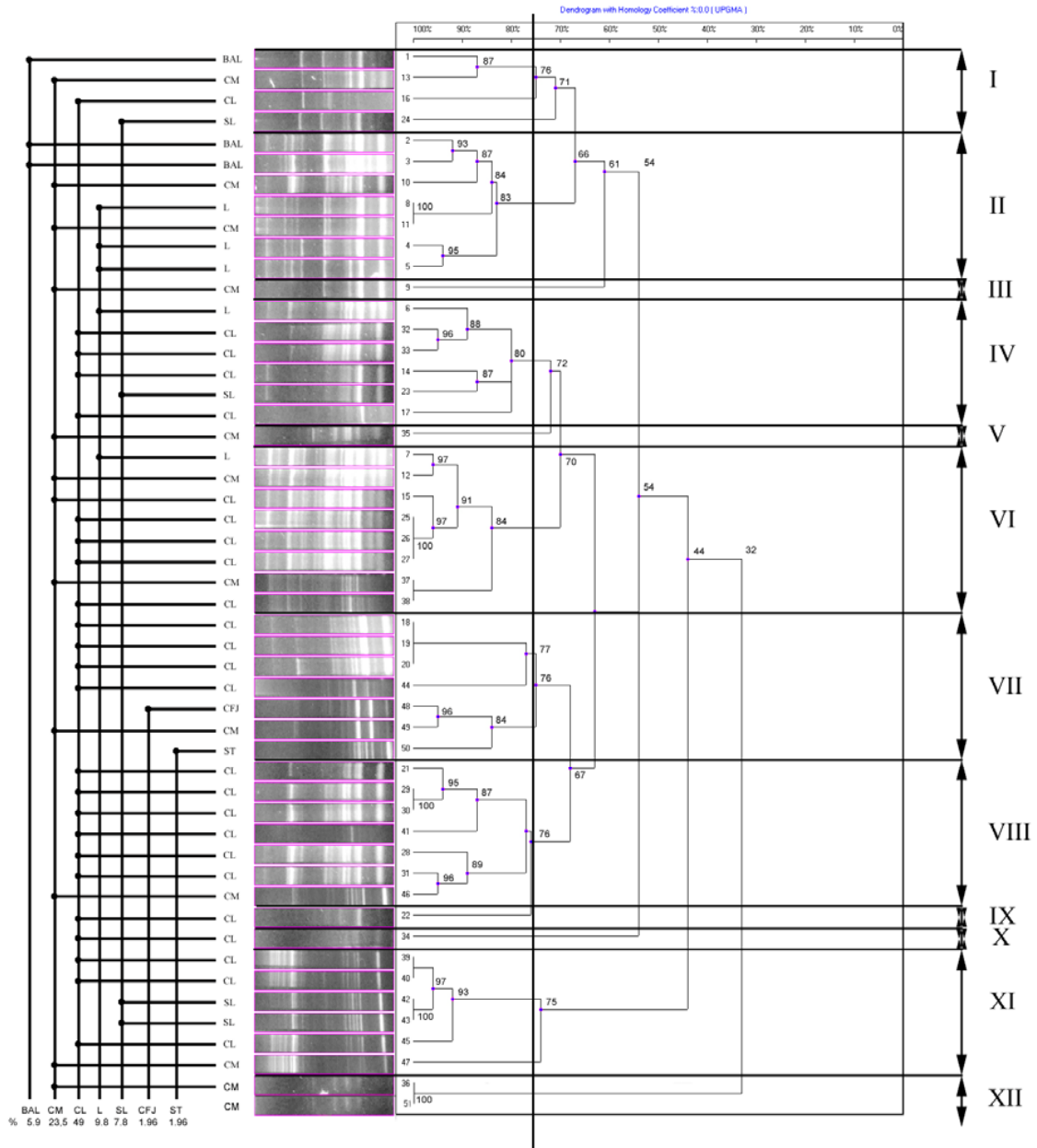
**Fig 1.** PCR analysis of virulence factor genes**Şekil 1.** Virülens faktör genlerinin PZR analizleri

12 clonal types, according to the source, case or sample they were recovered from. Of the 25 strains isolated from the liver of cattle, 6 belonged to type VIII, 5 to type VI, 4 to each of the types IV and VII, 3 to type XI and 1 to each of the types I, IX and X. It was determined that, of the 12 strains isolated from the milk of dairy cows with clinical mastitis, 2 belonged to each of the types II, VI and XII, and 1 to each of the types I, III, V, VII, VIII and XI. Of the isolates recovered from the lung samples of dairy cows, 3 were of the type II and 1 of each of the types IV and VI. On the other hand, of the strains isolated from the liver of sheep, 2 were classified as type XI and 1 as each of the types I and IV. Of the isolates recovered from broncho-alveolar lavage fluid (BAL), 2 belonged to type II and 1 to type I, whereas the two isolates recovered from the synovial fluid of a calf and from suppurative tissue were type VII.

## DISCUSSION

*Arcanobacterium pyogenes* is a ubiquitous opportunistic pathogen, which causes suppurative diseases in animals of economic value, including cattle and sheep, and in other animal species. *A. pyogenes* exists commensally in the upper respiratory tract and uro-genital system of animals<sup>1-8</sup>.

It has been reported that, synergistic and antagonistic haemolytic properties can be used as additional criteria for the identification of bacteria belonging to the genus *Arcanobacterium*<sup>20</sup>. In the present study, it was observed that, haemolysins of all *A. pyogenes* strains had a haemolytic effect on rabbit, sheep, cattle, chicken, dog and human erythrocytes. Furthermore, all isolates produced  $\beta$ -haemolysis on blood agar containing sheep blood.



**Fig 2.** Dendrogram of BOX-PCR from *A. pyogenes* 51 isolates. The degree of correlation showed on scala

**Şekil 2.** *A. pyogenes* 51 adet izolatlarının BOX-PCR ile Dendrogramı. Klonal tipler arasındaki benzerlik derecesi skala üzerinde gösterildi

The similar studies were reported that molecular epidemiological analyses of isolates were done from the same flock or cases of infection<sup>21, 22</sup>. In some cases, while it showed that isolates seems to be originate by a single clone, sometimes the heterogeneity determined among strains<sup>22</sup>. While some study<sup>23</sup>, were firstly given to date of properties of phenotypic and genotypic in the isolates from infections of animals, in other study<sup>24</sup>, molecular studies are undertaken to improve the diagnosis of the newly described bacteria.

Silva et al.<sup>19</sup> genotypically characterized the *A. pyogenes* isolates they recovered from dairy cows with either normal puerperium or clinical metritis, and identified the clonal

types of the isolates, which they had recovered from post-partum dairy cows belonging to the same herd. While some clonal types were found to be strictly associated with the development of clinical metritis, other types were identified from strains recovered from dairy cows with normal puerperium and clinical metritis. It was stated by these researchers that the presence of the virulence factor genes was not correlated with the capability of the agent in mastitis and other infections, and that *A. pyogenes* clonal types may not be a determinant factor in the development of the disease<sup>19,25</sup>. In the present study, the molecular identification of ovine and bovine *A. pyogenes* strains recovered from different cases of infection was assessed.

In the present study, it was determined using BOX-PCR that, the *A. pyogenes* isolates were of 12 clonal types, whilst Silva et al.<sup>19</sup> referred to 10 herd-specific clonal types (5 clonal types in each of the herds A and B). While Silva et al.<sup>19</sup> reported the rate of maximum similarity between the isolates of the two herds as 83.5%; in the present study, the maximum similarity rate of the isolates was determined to be 84.8%. In both studies, the maximum similarity rates of the isolates were rather high. While Silva et al.<sup>19</sup> reported that 82% of the isolates belonged to 5 clonal types, in the present study, 80.4% of the isolates belonged to 6 different clonal types (types II, IV, VI, VII, VIII and XI).

Silva et al.<sup>19</sup> pointed out to a vast heterogeneity among *A. pyogenes* isolates colonizing the uterus of dairy cows with normal puerperium and clinical metritis. Similarly, in the present study, it was determined that, based on the sample/case/source the isolates were recovered from, the *A. pyogenes* isolates displayed a high rate of heterogeneity for clonal types. The clonal type with greatest heterogeneity was determined as type VII (4 bovine livers, 1 calf synovial fluid, milk and suppurative tissue isolates); whilst, the clonal type with greatest homogeneity was type VIII (6 liver isolates and 1 milk isolate).

The number of clonal types was identified as 8 in isolates recovered from the liver of cattle slaughtered at a slaughterhouse, 9 in isolates recovered from the milk of dairy cows with mastitis, 3 in isolates recovered from the liver of sheep, and 2 in isolates recovered from the broncho-alveolar lavage fluid of dairy cows. The isolation of certain clonal types from different cases of infection and the identification of different clonal types from *A. pyogenes* isolates in the same case suggest that the pathogenicity of *A. pyogenes* should be further investigated in detail.

All of the clonal types identified were determined in isolates that were recovered from different sources and that are capable of inducing various infections. While the clonal types III, V and XII were similar strains isolated from only bovine mastitis, types IX and X were strains isolated from only bovine liver. On the other hand, some of the clonal types (I, II, IV, VI, VII, VIII, XI) were associated with strains isolated from different sources (broncho-alveolar lavage fluid, cow's milk, bovine liver, ovine liver, bovine lung, calf synovial fluid, suppurative tissue). The isolation of these clonal types from different infections and sources suggests that their potential to cause infection could be greater.

The genome of *A. pyogenes* contains a variety of known and putative virulence factors. The pyolysin is well known as a major virulence factor of this bacteria<sup>1,9,26</sup> and it is generally present in all *A. pyogenes* isolates<sup>8,17,20</sup>. In the present study, 2 different primers referred to by different researchers<sup>8,19</sup> were used for the *plo* gene. In result, all *A. pyogenes* isolates were found to be positive for the *plo* gene (band widths of 272 bp and 150 bp).

While Silva et al.<sup>19</sup> reported that all investigated *A. pyogenes* strains carried *nanH* and *nanP*, according to some studies<sup>1,25</sup>, some of *A. pyogenes* isolates were found to be positive for *nanH* (100% and 87%) and *nanP* (64.2% and 75%). In present study, positivity rates were determined as 62.7% and 84.3% for the *nanH* and *nanP*, respectively. The predominance of the *nanP* gene in bovine isolates has been indicated not to be surprising<sup>1,19</sup>. It has been stated that the *NanH* and *nanP* genes may contain different host substances and that the *nanP* gene may encode tropism for adhesion of the agent to host tissues<sup>12</sup>.

In many of studies<sup>1,19,25,27</sup> reported that *fimA* gene seems to be involved in adhesion processes and this gene was found to be high positive (98%, 94%, 100% and 90.9%) rates. The results of the present study corresponded to the findings of others<sup>1,19,25,27</sup>.

While Esmay et al.<sup>14</sup> reported the presence of the *cbpA* gene (encoding adhesion to the HeLA and 3T6 cells) in only 48% of the strains isolated from cases of different infections, in the present study, it was determined that 58.8% of the isolates were positive for the *cbpA* gene. While the rates of positivity determined in the present study were lower than those reported by Silva et al.<sup>19</sup>, it was observed that these results were close to those reported by Esmay et al.<sup>14</sup>.

The difference between the results obtained in this study and other researchers<sup>1,14,16,19,25,27</sup> is considered to have arisen from the isolates studied by the above mentioned researchers all being of uterine origin, and the isolates investigated in the present study having been recovered from different species, tissues and infections.

Silva et al.<sup>19</sup> reported that in *A. pyogenes* strains; clonal types, virulence gene profiles and pathogenicity (development of disease) were correlated with each other. However, no such correlation was determined in the present study. The underlying reason of the difference in the results of the two studies may be Silva et al.<sup>19</sup> having studied only on uterine samples obtained from two herds, and the strains investigated in the present study having been recovered from a wider range of sources.

The determination of the presence of virulence factors in the bacterial genome may not be adequate to show that the bacterium is capable of inducing disease. *A. pyogenes* isolates may possess their virulence factor genes differently under specific conditions. Host-intrinsic factors, synergic action between other bacteria and *A. pyogenes*, and differential gene expressions of virulence factor genes may have a more relevant effect on disease development<sup>19</sup>. The variety of factors expressed by *A. pyogenes* may explain how this microorganism is able to colonize many different host tissues and cause such diverse disease processes<sup>1,11,12,13,16</sup>.

In conclusion, the clonal types of *A. pyogenes* strains

isolated from different tissues and infections of sheep and cattle were identified in the present study. While all of the *A. pyogenes* isolates were positive for the *plo* gene, the positivity rates for other virulence factors differed among the isolates. Nonetheless, no correlation was determined between the virulence genes or clonal types and cases of infection.

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