RESEARCH ARTICLE

Investigation of Pathological Findings Infected with *Aeromonas* salmonicida in Rainbow Trout (*Oncorhynchus mykiss* Walbaum, 1792)^[1]

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Summary

Experimental furunculosis caused by *Aeromonas salmonicida* was induced in rainbow trout (*Oncorhynchus mykiss* Walbaum, 1792) using intraperitoneal injection and immersion challenge methods. A total of 65 (50 experimental and 15 control) rainbow trouts (weight 155 ± 15 g and 20-25 cm in size) were used for this experimental work. *Aeromonas salmonicida* at a concentration of $3x10^{\circ}$ cfu/ml (0.1 ml per fish) was intraperitoneally injected to twenty-five fish (first group). A second group of 25 fish was immersed in 30 l water inoculated to which 3 ml of bacterial inoculum ($3x10^{\circ}$ cfu/ml) had been added for one hour. Sterile saline solution (0.1 ml/fish) was intraperitoneally injected to control group. Necropsy of the fish was performed, macroscopic and microscopic findings were evaluated. The chronic form of disease was observed in both groups. Anorexia, irregular swimming, hemorrhages at the dorsal, ventral and pectoral base of fins were the main clinic and macroscopic findings. However, in some fish, diffused or focal scale eruption on the dermis from operculum to caudal fin, periocular hemorrhages and exophtalmos were observed. Similar microscopic findings were determined in the both experiment groups. Microscopically, foci of bacteria with no inflammatory response especially in the muscles, gills, heart, stomach, pyloric caeca, intestines, kidney, spleen and liquefactive necrosis in the muscles were the main microscopic findings.

Keywords: Aeromonas salmonicida, Rainbow trout, Experimental infection, Pathological findings

Aeromonas salmonicida ile Enfekte Edilen Gökkuşağı Alabalıklarında (Oncorhynchus mykiss Walbaum, 1792) Patolojik Bulguların İncelenmesi

Özet

Bu araştırma *Aeromonas salmonicida* ile deneysel Furunkülozis'in intraperitoneal yol ve immersiyon yöntemiyle oluşturulması amacıyla yapıldı. Bu amaç için, 50 adet deneme grubu, 15 adet kontrol grubu olmak üzere toplam 65 adet, 155±15 g ağırlığında, 20-25 cm büyüklüğünde gökkuşağı alabalığı (*Oncorhynchus mykiss* Walbaum, 1792) kullanıldı. İntraperitoneal yolla oluşturulan enfeksiyonda (birinci grup) 25 adet balığa 3x10^s hücre/ml olacak şekilde 0.1 ml bakteri inokulatı verildi. İmmersiyon yöntemiyle uygulama yapılan diğer gruptaki (ikinci grup) 25 adet balığa ise 0.1 ml bakteri inokulatı verildi. İmmersiyon yöntemiyle uygulama yapılan diğer gruptaki (ikinci grup) 25 adet balık, 3x10^s hücre/ml olacak şekilde 3 ml bakteri inokulatı verilen 30 l suda 1 saat bekletildi. Kontrol grubundaki 15 balığa ise 0.1 ml serum fizyolojik intraperitoneal olarak verildi. Bu balıkların sistemik nekropsileri yapıldı, makroskobik ve mikroskobik bulgular değerlendirildi. Her iki deneme grubunda da hastalığın kronik formu şekillendi. Balıklarda klinik ve makroskobik olarak, iştahsızlık, deri renginde koyulaşma, yüzme bozuklukları, dorsal, ventral ve pektoral yüzgeçlerin tabanında kanamalar görüldü. Ayrıca, bazı balıklarda, operkulumdan kuyruk yüzgecine kadar uzanan bölgede pullarda dökülmeler, perioküler kanamalar ve ekzoftalmus dikkati çekti. Her iki deneme grubunda da, mikroskobik olarak benzer bulgular görüldü. Balıklarda mikroskobik olarak kaslarda, solungaçlarda, kalpte, midede, pilorik keselerde, bağırsaklarda, böbreklerde ve dalakta yangısal bir yanıt görülmeksizin bakteri kümeleri ve kaslarda erime nekrozları saptandı.

Anahtar sözcükler: Aeromonas salmonicida, Gökkuşağı alabalığı, Deneysel enfeksiyon, Patolojik bulgular

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INTRODUCTION

Furunculosis, caused by *Aeromonas salmonicida*, is a septicaemic and contagious disease characterized by the furuncle formation on the skin with necrotic and hemorrhagic muscle lesions ^{1,2}, and leads to outbreaks in many families of fish, especially Salmonidae and Cyprinidae ^{3,4}. In Turkey, *Aeromonas salmonicida* was isolated from the three fish hatcheries located in West-Aegean region in 2003 ⁵. The disease may be seen in peracute, acute, subacute and chronic forms. The peracute form of furunculosis frequently occurs in the fingerling fish. The acute form of furunculosis is common in growing fish and fish usually die within 2-3 days. Subacute and chronic forms are more common in older fish, and at the beginning of infection, deaths occur rarely, but the deaths rate gradually increases ⁶.

Aeromonas salmonicida is a gram-negative, aerobic, non-motile, fermentative, non-spore forming, bacillus approximately 1.0 μm x 2.0 μm in size ^{1,3,6}. The cell wall of Aeromonas salmonicida possesses some virulent factors such as an additional cell envelope protein (A-Layer Protein) and lipopolysaccharid ^{7:10}. A-Layer Protein is important for virulence. Because Aeromonas salmonicida, possesses A-Layer Protein, has the ability to adhere, enter and to survive within macrophages ^{7,11,12}. In addition, the extracellular products (ECP) of Aeromonas salmonicida consisted of 70-kDa serine protease (extracellular protease, EP) and glycerophospholipid:cholesterol acyltransferase complexed with lipopolysaccharide (GCAT/LPS) play an important role in pathogenesis and virulence ¹³⁻¹⁵.

The aim of this study was to investigate macroscopic and microscopic findings in rainbow trouts experimentally infected with *Aeromonas salmonicida* via the intraperitoneal route and immersion challenge methods.

MATERIAL and METHODS

The ethics committee of Adnan Menderes University approved the design and protocol of the experiments (15.01.2004-2004/0013). In this research, a total of 65 (50 fish in experimental groups and 15 fish in control group) rainbow trouts (weight 155±15 g and 20-25 cm in size) obtained from a commercial fish farm with no history of furunculosis, were used. Fish were divided into three groups: First group (25 fish), second group (25 fish) and control group (15 fish). They were acclimatized for 14 days in three separate pools (one for control group and two for experimental groups). Water temperature was maintained at 15-18°C. During the experimental period, external filter (600 l/h) was used in all aquariums. Dissolved oxygen of the water and pH in all pools were uniform (pH 7.25 and dissolved oxygen 9.1-9.4 mg/l). The fish were fed daily with a commercial feed (No.2 Bağcı). Before the intraperitoneal injections, fish were anaesthetized with 2-phenoxyethanol (0.25%) ¹⁶. Aeromonas salmonicida (field strain) at a concentration of 3x10⁵cfu/ml (0.1 ml per fish) was intraperitoneally injected to twenty-five fish in the first group. The other twenty-five fish (the second group) were immersed into 30 I water inoculated with 3 ml of bacterial inoculum (3x10⁵ cfu/ml) for one hour. The same procedure was repeated after three days. Sterile saline solution (0.1 ml per fish) was intraperitoneally injected to fifteen fish in the control group. Experiment was performed for 31 days. Necropsy of the fish was performed and tissue samples were collected and fixed in 10% formalin solution, embedded in paraffin, sectioned at 5 µm and stained routinely with Hematoxylin-Eosin (H&E). The selected kidney, liver, spleen, heart and gills were stained by Brown and Brenn staining method for the isolation of bacteria. In addition, to evaluate the presence of lipid in the liver, Oil Red O staining method was used ¹⁷.

RESULTS

In this experiment, chronic form of furunculosis was developed in the both groups. In the first group, the deaths began on 6th day and reached to maximum rate on 27-28th day; in the second group, the deaths began on 7th day and reached maximum rate on 23-25th day. The surviving fish were euthanasied on the 31st day of the experiment [total 7 fish (3 fish from first group, 4 fish from second group)]. Fish in the control group were euthanasied and changes were observed comparatively. *Aeromonas salmonicida* was reisolated from the infected fish and no *Aeromonas salminicida* or any other pathogenic bacteria were isolated from fish in the control group.

Clinical signs consisted of anorexia, darkening in the color or depigmentation of the skin and irregular swimming in the both groups.

Macroscopic Findings

In both groups, macroscopically, hemorrhage at the dorsal, ventral and pectoral base of fins, diffused or focal scale eruption on the dermis from operculum to caudal fin were often noted. Furthermore, diffuse hemorrhages in the kidney, stomach and intestines; periocular hemorrhages and exophtalmos; anemia; hyperemia or anemia in the gills; a great reduction in the size of kidney tissue; ascites and petechial hemorrhages on the perivisceral tissue (*Fig. 1*) were determined in some fish. Generally, due to anorexia, the stomach, pyloric caeca and intestines were empty.



Fig 1. Petechial hemorrhages on the perivisceral tissue (arrow) **Şekil 1.** Periviseral dokuda peteşiyal kanamalar (ok)

Microscopic Findings

Similar microscopic findings were determined in the both experiment groups. The main microscopic findings were the foci of bacteria with no inflammatory response in the different tissue and organs, and liquefactive necrosis in the muscles.

The liquefactive necrosis was observed in the muscles (*Fig. 2A*). The sizes and the locations of the necrosis varied among the fish. Some of the necrotic muscle bundles were filled with the destruction products or they were completely empty.

Foci of bacteria with no inflammatory response were observed in the necrotic areas of muscle (*Fig. 2B*); around secondary lamellae in the gills; in the heart; serosal surface of the stomach, pyloric caeca and intestines; in the tubular lumen of the kidney; capsular surface of the spleen and kidney.

In the liver, discrete vacuoles were present in the cytoplasm of the hepatocytes. These vacuoles were

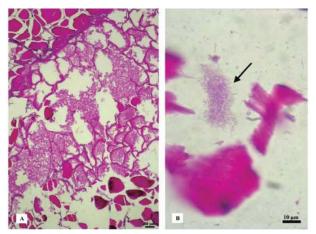


Fig 2. *A:* The liquefactive necrosis in the muscle tissue. HE. Bar, 50 μ m, *B:* Foci of bacteria with no inflammatory response in the muscle tissue (arrow), HE. Bar, 10 μ m

Şekil 2. A: Kaslarda erime nekrozu. HE. Bar, 50 μm, **B:** Kaslarda yangısal bir yanıt görülmeksizin bakteri kümeleri (ok), HE. Bar, 10 μm

recognized as fat droplets stained with Oil Red O method in frozen sections.

Hydropic degeneration in the proximal and distal tubules, reduction in the lymphoid cells of hematopoietic tissue (*Fig. 3*), and glomerular athrophy were determined in the kidney. The most important microscopic lesion in the spleen was necrosis in the lymphoid tissue.

The primary and secondary lamellar lesions consisted of edema, adhesion, hyperplasia, hyperemia and telangiectasia in the gills. Marked edema in the secondary lamellae was often present.

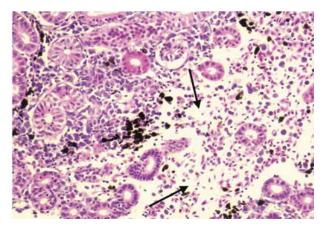


Fig 3. Reduction in the lymphoid cells of hematopoietic tissue in the kidney (arrows), HE. Bar, 50 $\,\mu m$

Şekil 3. Böbrekte hematopoietik dokunun lenfoid hücrelerinde azalma (oklar), HE. Bar, 50 µm

In addition, elevated numbers and degranulation of eosinophilic granular cells (EGC) in the submucosa of the stomach and pyloric caeca were determined. However, same finding was detected in only one fish's gill.

DISCUSSION

In the chronic form of furunculosis anorexia, exophtalmos, darkening in the color of skin, irregular swimming, hemorrhages at the dorsal, ventral and pectoral bases of fins, and furuncle formation at the skin were reported as the main clinic and macroscopic findings ^{6,12,14}. In the present study, anorexia, irregular swimming, darkening in the color of skin, hemorrhages at the bases of fins, scale eruptions were observed following the inoculations. But furuncle formation was not seen.

It has been reported that the 70-kDa serine protease causes of liquefaction necrosis in the muscles ¹³. In addition, on the other hand, GCAT/LPS combination with 70-kDa serine protease induces liquefaction necrosis in the muscles ¹⁸. In the present study, the liquefaction

necroses in the muscles were located especially around foci of bacteria, and we suggested that liquefaction necrosis in the muscles could be related to the extracellular products of *Aeromonas salmonicida*.

One of the important microscopic findings of furunculosis is foci of bacteria with no inflammatory response in the dermis, muscle and internal organs ^{1,19}. Recent studies suggest that GCAT/LPS have haemolytic, leucocytolytic and cytotoxic activities ^{6,12,20}. That is because of these effects of GCAT/LPS, destruction of hematopoietic tissue occurs in kidney and spleen. Moreover, a few authors suggested that Aeromonas salmonicida, possesses A-Layer Protein, the ability to adhere, enter and, to survive within macrophages bring about phagocytosis inhibition ^{7,11}. Phagocytosis inhibition and hematopoietic tissue destruction constitute foci of bacteria with no inflammatory response in the dermis, muscles and internal organs. In this study, foci of bacteria with no inflammatory response in the muscles, gills, heart, stomach, pyloric caeca, intestines, kidney and spleen, and reduction in the lymphoid cells of hematopoietic tissue of the kidney were seen. As a matter of fact the selected tissue sections were stained by Brown and Brenn staining method for bacteria and positive results were obtained. We suggested that foci of bacteria with no inflammatory response in the muscles, gills and internal organs might be related to A Layer Protein and activity of GCAT/LPS.

In conclusion, in both groups, macroscopic and microscopic findings of this study might be considered important pathological findings of furunculosis in rainbow trout.

REFERENCES

1. Ribelin WE, Migaki G: The Pathology of Fishes. 1st ed., The University of Wisconsin Press, Wisconsin, 1975.

2. Roberts RJ: Fish Pathology. 3rd ed., W.B. Saunders, Toronto, 2001.

3. Austin B: Progress in understanding the fish pathogen *Aeromonas salmonicida. Mar Biotechnol,* 15, 131-134, 1997.

4. Erer H: Balık Hastalıkları, 2. Baskı, Selçuk Üniversitesi Basımevi, Konya, 2002.

5. Kırkan Ş, Göksoy EÖ, Kaya O: Isolation and antimicrobial susceptibility of *Aeromonas salmonicida* in rainbow trout (*Oncorhynchus mykiss*) in Turkey hatchery farms. *J Vet Med B Infect Dis Vet Public Health*, 50, 339-342, 2003.

6. Woo PTK, Bruno DW: Fish Diseases and Disorders: Viral, Bacterial and Fungal Infections. 1st ed., CABI Publishing,

New York, 1999.

7. Garduno RA, Thornton JC, Kay WW: Aeromonas salmonicida grown in vivo. Infect Immun, 61 (9): 3854-3862, 1993.

8. Ellis AE, Vale AD, Bowden TJ, Thompson K, Hastings TS: *In vivo* production of A-protein, lipopolysaccharide, iron-regulated outer membrane proteins and 70-kDa serine protease by *Aeromonas salmonicida subsp. salmonicida*. *FEMS Microbiology Letters*, 149, 157-163, 1997.

9. Dalmo RA, Seternes T, Arnesen SM, Jorgensen TO, Bogwald J: Tissue distribution and cellular uptake of *Aeromonas salmonicida* lipopolysaccharide (LPS) in some marine fish species. *J Fish Dis*, 21, 321-334, 1998.

10. Simko E, Kocal TE, Quinn BA, Ostland VE, Ferguson HW, Hayes MA: Influences of *Aeromonas salmonicida* lipopolysaccharide, prednisolone and water temperature on plasma protein composition in salmonids. *J Fish Dis,* 22, 91-100, 1999.

11. Garduno RA, Moore AR, Olivier G, Lizama AL, Garduno E, Kay WW: Host cell invasion and intracellular residence by *Aeromonas salmonicida*: Role of the S-layer. *Can J Microbiol,* 46, 660-668, 2000.

12. Cipriano RC, Bullock GL: Furunculosis and other diseases caused by *Aeromonas salmonicida*. *Fish Disease Leaflet* 66, pp. 1-19, 2001.

13. Lee KK, Ellis AE: The role of the lethal extracellular cytolysin of *Aeromonas salmonicida* in the pathology of furunculosis. *J Fish Dis,* 14, 453-460, 1991.

14. Vipond R, Bricknell IR, Durant E, Bowden TJ, Ellis AE, Smith M, Macintyre S: Defined deletion mutants demonstrate that the major secreted toxins are not essential for the virulence of *Aeromonas salmonicida*. *Infect Immun*, 66 (5): 1990-1998, 1998.

15. Arnesen JA, Eggset G: Isolation and characterisation of two extracellular metalloproteases from *Aeromonas* salmonicida ssp. salmonicida. J Fish Dis, 22, 35-43, 1999.

16. Avci H, Birincioğlu SS: Pathological findings in rainbow trout (*Oncorhynchus mykiss* Walbaum, 1792) experimentally infected with Yersinia ruckeri. Turk J Vet Anim Sci, 29, 1321-1328, 2005.

17. Luna GL: Histologic Staining Methods of The Armed Forces Institute of Pathology. 3rd ed., Mcgraw Hill Book Company, US, 1968.

18. Ellis AE: An appraisal of the extracellular toxins of *Aeromonas salmonicida ssp. salmonicida. J Fish Dis,* 14, 265-277, 1994.

19. Bricknell IR, Bruno DW, Stone J: *Aeromonas salmonicida* infectivity studies in goldsinny wrasse, *Ctenolabrus rupertis. J Fish Dis,* 19, 469-474, 1996.

20. Lee KK, Ellis AE: Glycerophospholipid:Cholesterol Acyltransferase complexed with Lipopolysaccharide (LPS) is a major lethal exotoxin and cytolysin of *Aeromonas salmonicida*: LPS stabilizes and enhances toxicity of the enzyme. *J Bacteriol*, 172 (9): 5382-5393, 1990.