Brucella melitensis Isolated from Aborted Cow and Sheep Fetuses in Northwest of China

Huan ZHANG^{1,#,a} Shengnan SONG^{1,#} Benben WANG^{3,#} Yali JIANG³ Wenxing WU¹ Fei GUO² Yang LIU¹ Qian WANG² Junbo ZHANG⁴ Hui ZHANG¹ Jinliang SHENG¹ Yuanzhi WANG² Chuangfu CHEN¹

[#] These authors contributed equally to this work

¹ School of Animal Science and Technology, Shihezi University, 832000-Shihezi City, Xinjiang, CHINA

² School of Medicine, Shihezi University, 832000-Shihezi City, Xinjiang, CHINA

³ School of Life Science, Shihezi University, 832000-Shihezi City, Xinjiang, CHINA

⁴ College of Agroforestry Engineering and Planning, Tongren University, 554300-Tongren City, Guizhou, CHINA ^a ORCID: 0000-0001-9366-2385

Article Code: KVFD-2017-18881 Received: 14.10.2017 Accepted: 25.12.2017 Published Online: 25.12.2017

How to Cite This Article

Zhang H, Song S, Wang B, Jiang Y, Wu W, Guo F, Liu Y, Wang Q, Zhang J, Zhang H, Sheng J, Wang Y, Chen C: Brucella melitensis isolated from aborted cow and sheep fetuses in Northwest of China. Kafkas Univ Vet Fak Derg, 24 (2): 307-310, 2018. DOI: 10.9775/kvfd.2017.18881

Abstract

This study aimed to investigate the causes of abortion in cows that were mixed fed with sheep and/or goats, and the presence of *B. melitensis* infection in cows. PCR of 34 (28%) specimens out of 120 samples were identified as *B. melitensis*. The traditional bacteriological tests identified all of the isolates from sheep/cow aborted fetuses and milk as *B. melitensis* biovar 3. This is the first study to demonstrate *B. melitensis* as the main etiological agent for cows mixed fed with sheep and/or goats in XUAR, northwest of China. This may cause severe infection in the local population, and pose a potential public health risk, especially when eating or drinking the products of contaminated milk.

Keywords: Brucella melitensis, Cow, Abortion, China

Kuzey Batı Çin'de Atık İnek ve Koyun Fetüslerinden *Brucella melitensis* İzolasyonu

Öz

Bu çalışmanın amacı koyun ve/veya keçilerle birlikte yetiştirilen ineklerde atıkların sebeplerini araştırmak ve *B. melitensis* enfeksiyonunun mevcudiyetini belirlemektir. Toplam 120 örneğin 34'ünde (%28) PCR ile *B. melitensis* tespit edildi. Klasik bakteriyolojik testler, koyun ve inek atık fetüsleri ile sütten elde edilen izolatların tümünde etkenin *B. melitensis* biovar 3 olduğunu belirledi. Bu çalışma, Çin'in Sincan Uygur Özerk Bölgesi'nde koyun ve/veya keçilerle birlikte yetiştirilen ineklerde *B. melitensis*'în atıklarda ana etiyolojik ajan olduğunu göstermektedir. Bu durum bölge popülasyonda ciddi enfeksiyona neden olabilir ve bu suretle özellikle kontamine süt ürünleri tüketiminde potansiyel halk sağlığı riski oluşturabilir.

Anahtar sözcükler: Brucella melitensis, İnek, Abort, Çin

INTRODUCTION

Brucellosis, a zoonosis of public health and economic importance worldwide, leads to great loss to domestic animals, principally in cows, sheep and goats. In some countries, particularly in southern Europe and Western Asia, where cattle are kept in close contact with sheep and goats, infection can also be caused by *B. melitensis*^[1],

İletişim (Correspondence)

- +86 0993 2058002 (Chuangfu Chen); +86 0993 2015620 (Yuanzhi Wang)
- ccf-xb@163.com (Chuangfu Chen); wangyuanzhi621@126.com (Yuanzhi Wang)

but the symptom of abortion in cows is not as severe as in sheep or goats. Mixed farming is also adopted by smallholder farmers in China.

Six classical species of *Brucella* have been identified, including *B. abortus*, *B. melitensis*, *B. suis*, *B. ovis*, *B. canis*, and *B. neotomae*. Although *B. abortus* is considered as the main etiological agent of infected dairy cows, *B. melitensis*

results in the greatest loss to domestic animal industry and public health ^[2,3]. Therefore, investigating the infection of *B. melitensis*, as a nonspecific and heterogeneous agent in dairy herds, is crucial.

Xinjiang Uygur Autonomous Region (XUAR) in the northwest of China, is the largest province in China, and the livestock industry is the main source for its economic growth ^[4]. Very few studies have been conducted on prevalence and distribution of brucellosis in remote areas. The aim of this study was to investigate the etiological agents responsible for abortions in sheep or cows in endemic areas of brucellosis, and the presence of *B. melitensis* as a heterogeneous agent in dairy cows.

MATERIAL and METHODS

Bacterial Strains

Reference strains of *Brucella melitensis* 16M, *B. abortus* 2308 and *Toxoplasma gondii* as well as *Campylobacter fetus spp*, *Theileria sergenti* and *Tritrichomonas fetus* were provided by Anthropozoonosis laboratory in Shihezi University.

Sample Collection and DNA Extraction

The samples including aborted fetuses (n = 120) and raw milk (n = 1) were collected from IIi region (northwest of XUAR) in some sheep and cow mix feeding farms between April and May in 2016. Samples of spleen, liver and lung tissues and stomach contents were collected aseptically from sheep or cow aborted fetuses and raw milk from a cow with clinical signs of joint swelling and abortion history. Then the DNA extraction from tissue samples was performed using the TIANamp Genomic DNA Kit (TIANGEN

BIOTECH CO., LTD) according to the manufacturer's instructions. The nucleic acid extraction from raw milk was performed as previously described ^[5]. DNA concentrations were determined by measuring the A_{260} , and the samples were stored at -20°C until further processing.

Synthetic Oligonucleotide Design

Oligonucleotide species-specific primers for *Brucella* genus ^[6], *Toxoplasma gondii* ^[7], *Campylobacter fetus* ^[8], *T. buffeli* ^[9], and *Tritrichomonas fetus* ^[10] are listed in *Table 1*.

PCR Amplification and Sequence Analysis

All samples were examined by PCR in a total volume of 30 μ L, with 12.5 μ L ddH₂O, 15 μ L mix, 0.5 μ L of each primer and 1.5 μ L DNA template. The reaction was performed in a DNA thermal cycler (Perkin-Elmer) and 2 μ L of the product was fractionated in a 1.5% or 2% agarose gel, stained with 0.5 mg/mL ethidium bromide solution, and visualized under UV light ^[11]. The positive amplication products were purified using the TIAN-gel Midi Purification Kit (TIANGEN, Beijing, China) and then subjected to sequencing. All of these data was analyzed using SPSS version 17.0 software.

Bacterial Isolation

Brucella was isolated from raw milk sample as previously described ^[12]. The tissue samples were homogenized before plating on the Brucella-selective agar. Then, 100 μ L of the homogenized suspension was inoculated onto Brucella-selective agar plates. The suspension was spread with a loop producing a depot followed by single colonies. All cultures were incubated at 37°C with 5% CO₂ for five days. Brucella identification and species differentiation were accomplished using PCR protocols ^[13]. Furthermore,

Table 1. PCR primers used for screening abortion-inducing pathogens in sheep				
Primer	Primer Sequences (5'-3')	Size (bp)		
Brucella-omp22- primer -F	TGATGGGAGGGACCGACTA	526		
Brucella-omp22- primer -R	TGGTTCTTCAGGTTGTTACGC	526		
B. abortus-IS711- primer -F	GACGAACGGAATTTTTCCAATCCC	524		
B. abortus- IS711- primer -R	TGCCGATCACTTAAGGGCCTTCAT	526		
B. melitensis- IS711- primer -F	AAATCGCGTCCTTGCTGGTCTGA	724		
B. melitensis- IS711- primer -R	TGCCGATCACTTAAGGGCCTTCAT	731		
C. fetus- sapB2- prime r-F	GCAAATATAAATGTAAGCGGAGAG	425		
C. fetus- sapB2- prime r-R	TGCAGCGGCCCCACCTAT	435		
T. buffeli-18S rRNA- primer -F	AAACTGCGAATGGCTCAT	016		
T. buffeli-18S rRNA- primer -R	ACATCCTTGGCAAATGCT	816		
<i>T. fetus-TFITS-</i> primer - F	CTGCCGTTGGATCAGTTTCG	200		
<i>T. fetus-TFITS-</i> primer - R	GCAATGTGCATTCAAAGATCG	208		
Toxoplasma gondii-specific-primer -F	CGCTGCAGGGAGGAAGACGAAAGTTG	520		
Toxoplasma gondii-specific-primer-R	CGCTGCAGACACAGTGCATCTGGATT	529		
^a The pair of primers of Brucella omp22 w	vere used to screen Brucella spp. in the first round identificatio	n: ^b The pair of primers of Brucella		

^a The pair of primers of Brucella omp22 were used to screen Brucella spp. in the first round identification; ^b The pair of primers of Brucella IS711 were used to differentiate the species of Brucella

309

biotyping of the *Brucella* was based on conventional bacteriological and typing methods ^[14]. This process was completed at the Center for Disease Prevention and Control (CDC) of China.

RESULTS

Molecular Detection

In the present study, molecularly positive products for *Brucella* genus were found but not for *T. gondii*, *C. fetus spp*, *T. sergenti* and *Tr. fetus*. Thirty-four (28.09%) samples, including 13 aborted sheep fetuses, 20 cow fetuses and one raw milk sample, were positive and further identified as *B. melitensis* by targeting *IS711* gene and only part of the positive samples were presented in *Fig.1*, the rest of data was not shown. *B. abortus* and other species were not detected in the organs of aborted sheep, cow fetuses and raw milk. The nucleotide sequences from our study have

been deposited in the GeneBank database (accession number: KY312521). There were no differences in DNA sequences as compared to that of *B. melitensis* biovar 3 NI strain (accession number: CP002931)^[15].

Isolation of Brucella spp. from Organs

Bacteria were isolated from 34 samples and positively detected by *IS711* gene. The *Brucella* wild strains were isolated from 13 aborted sheep fetuses, 20 aborted cow fetuses and one raw milk sample. The detailed information is shown in *Table 2*. Furthermore, the culture isolates were identified as *B. melitensis* biovar 3 by conventional bacterial tests.

DISCUSSION

In XUAR, brucellosis has prevailed for decades ^[16], where the seropositive rates for cows and sheep were 0.66%

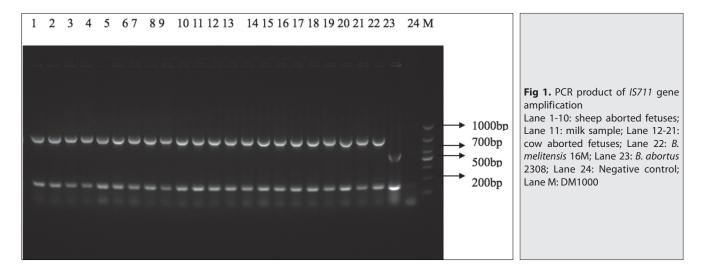


Table 2. Detection B. melitensis in in	dividual tissue or milk samples by
bacterial isolation	

Animal No	Organs	Host	Result		
1	Spleen Liver Lung	Cow	+ + -		
2	Spleen Liver Lung	Sheep	+ - -		
3	Spleen Liver Stomach contents	Sheep	- + +		
4	Spleen Liver Milk	Cow	+ - +		
5-15	Splen Liver	Sheep	+++++		
16-34	Splen Liver Stomach contents	Cow	+ + -		

and 3.25%, respectively, during 2013-2014 ^[17], and there are many pathogens could induce abortions in pregnant animals such as *Coxiella burnetii*, *Chlamydophila abortus*, *Salmonella enterica Serovar Abortusovis*, *T. gondii*, and *Neospora caninum* ^[18]. But, in the present study, *Brucella* was found to be the main pathogen responsible for livestock abortion and the rest of pathogens listed in *Table 1* were not found in these aborted fetuses, the result suggests that the *Brucella* pose the biggest threat to local livestock and people due to the infected cow could spread the disease through milk or contaminated dairy products. Interestingly, all of the isolates were identified as *B. melitensis* biovar 3 by conventional bacteriological and typing methods ^[14].

In Turkey, *B. melitensis* biovar 3 was first isolated from bovine aborted fetus ^[1]. In China, It was isolated in raw milk from an aborted cow at a farm that had about 300 sheep and 40 cows in Inner Mongolia, north of China ^[15]. The phenomenon of a host shift (i.e., the ability of a pathogen to colonize or infect a new host) is rare and appears in

resource-poor communities in China due to the mixed feeding of cows with infected sheep and/or goats and ignoring brucellosis quarantine. This study described *B. melitensis* isolates from aborted cow fetuses and raw milk. The result suggests that *B. melitensis* infection in cows is an emerging livestock industry and public health issue in China. As demonstrated in this study, *B. melitensis* can be shed in raw milk from infected cows. In addition, infection might spread to farm workers, slaughterers, and veterinarians through handling infected animals or organs after slaughter ^[19]. *B. melitensis* infection in cows may become more common in the future, although no data is available on brucellosis patients due to *B. melitensis* infection transmitted by raw milk or its products in China.

The problem of cows infected by *B. melitensis* has potentially important implications for the control programs of brucellosis in China. The clinical symptoms of *B. melitensis* infection in cows is not apparent as compared to that of *B. abortus* infection ^[20]. The infected cow, as a reservoir, is susceptible to disseminating contaminated milk to the local or neighboring population. This study recommends: i) avoiding intermixed feeding model of cows, sheep or/and goats in the same yard in endemic areas, and ii) increasing regular quarantine of brucellosis, and timely elimination of the infected sheep, goats and cows from the herd.

ACKNOWLEDGMENTS

This study was supported in part by grants from the National Key Research & Development plan (2017YFD0500304), National Natural Science Foundation of China (Grant Nos. U1503283, 81560338, 31572491, 31502067 and 31660705).

REFERENCES

1. Buyukcangaz E, Sen A: The first isolation of *Brucella melitensis* from bovine aborted fetus in Turkey. *J Biol Environ Sci*, 1 (3): 139-142, 2007.

2. Yazdi HS, Kafi M, Haghkhah M, Tamadon A, Behroozikhah AM, Ghane M: Abortions in pregnant dairy cows after vaccination with *Brucella abortus* strain RB5 1. *Vet Record*, 165 (19): 570-571, 2009. DOI: 10.1136/vr.165.19.570

3. Zowghi E, Ebadi A: Typing of *Brucella* strains isolated in Iran. *Arc Inst Razi (ARI)*, 33, 109-114, 1982.

4. Gao P, Huang L, Guo T, Sang D: A study of difficulties in developing herbivorous livestock in Xinjiang. *Finance Econ Xinjiang*, 2015. DOI: 10.3969/j.issn.1007-8576.2015.01.007

5. Cremonesi P, Castiglioni B, Malferrari G, Biunno I, Vimercati C, Moroni P, Morandi S, Luzzana M: Improved method for rapid DNA extraction of mastitis pathogens directly from milk. J Dairy Sci, 89 (1): 163169, 2006. DOI: 10.3168/jds.S0022-0302(06)72080-X

6. Khosravi AD, Abassi E, Alavi SM: Isolation of *Brucella melitensis* and *Brucella abortus* from brucellosis patients by conventional culture method and polymerase chain reaction technique. *Pak J Med Sci*, 22 (4): 396-400, 2005.

7. Homan WL, Vercammen M, De BJ, Verschueren H: Identification of a 200- to 300-fold repetitive 529 bp DNA fragment in *Toxoplasma gondii*, and its use for diagnostic and quantitative PCR. *Int J Parasitol*, 30 (1): 69-75, 2000. DOI: 10.1016/S0020-7519(99)00170-8

8. Wang G, Clark CG, Taylor TM, Pucknell C, Barton C, Price L, Woodward DL, Rodgers FG: Colony multiplex PCR assay for identification and differentiation of *Campylobacter jejuni, C. coli, C. lari, C. upsaliensis,* and *C. fetus* subsp. *fetus. J Clin Microbiol,* 40 (12): 4744-4747, 2002. DOI: 10.1128/JCM.40.12.4744-4747.2002

9. Jang S, Cho K, Chae JS, Kang SH: Fast diagnosis of bovine Theileriosis by whole blood PCR and microchip electrophoresis. *Bull Korean Chem Soc*, 25, 2004. DOI: 10.5012/bkcs.2004.25.5.757

10. Gookin JL, Birkenheuer AJ, Breitschwerdt EB, Levy MG: Singletube nested PCR for detection of tritrichomonas foetus in feline feces. *J Clin Microbiol*, 40 (11): 4126-4130, 2002. DOI: 10.1128/JCM.40.11.4126-4130.2002

11. Sambrook J, Fritsch EF, Maniatis T: Molecular Cloning: A Laboratory Manual, Second Ed., Cold Spring Harbor Laboratory Press, NY, 1989.

12. Langoni H, Ichihara SM, Silva AVD, Pardo RB, Tonin FB, Mendonça LJP, Machado JAD: Isolation of *Brucella* spp. from milk of brucellosis positive cows in São Paulo and Minas Gerais states. *Brazil J Vet Res Anim Sci*, 37 (6): 444-448, 2000. DOI: 10.1590/S1413-9596200000600004

13. Hinić V, Brodard I, Thomann A, Cvetnić Z, Makaya PV, Frey J, Abril C: Novel identification and differentiation of *Brucella melitensis, B. abortus, B. suis, B. ovis, B. canis,* and *B. neotomae* suitable for both conventional and real-time PCR systems. *J Microbiol Met,* 75 (2): 375-378, 2008. DOI: 10.1016/j.mimet.2008.07.002

14. Alton GG, Jones LM, Angus RD, Verger JM: Techniques for the brucellosis laboratory. 13 (6): 420, 1988.

15. Liu W, Jing Z, Ou Q, Cui B, He Y, Wu Q: Complete genome sequence of *Brucella melitensis* biovar 3 strain NI, isolated from an aborted bovine fetus. *J Bacteriol*, 194 (22): 6321, 2012. DOI: 10.1128/JB.01595-12

16. Muhtarhasan, Hai-Bo HE, Tai XP, Chen X, Tong SX, Wang Z: Analysis of surveillance data and epidemic situation of human brucellosis in Xinjiang, 2013. *Chin J Vec Biol Cont*, 2015. DOI: 10.11853/j. issn.1003.4692.2015.01.024

17. Zong-Lin HE, XIAKLA, Bai ZH, Kang Q, AYINUER, Yan-Rong MA, Zhu TT, Wan XJ, Song GH, Wu-Chang GU: Epidemiological survey of livestock brucellosis in Aksu area. *Grass-Feeding Livestock*, 2014. DOI: 10.3969/j.issn.1003-6377.2014.05.014

18. Masala G, Porcu R, Daga C, Denti S, Canu G, Patta C, Tola S: Detection of pathogens in ovine and caprine abortion samples from Sardinia, Italy, by PCR. *J Vet Diag Invest*, 19 (1): 96-98, 2007. DOI: 10.1177/ 104063870701900116

19. Di GE, De MF, Ancora M, Zilli K, Alessiani A: Typing of *Brucella* field strains isolated from livestock populations in Italy between 2001 and 2006. *Vet Italiana*, 44 (2): 383-388, 2008.

20. Sharifiyazdi H, Haghkhah M, Behroozikhah AM, Nematgorgani E: Bacteriological and molecular investigation of *B. melitensis* in dairy cows in Iran. *Comp Clin Pathol*, 21 (3): 269-273, 2012. DOI: 10.1007/ s00580-010-1090-6