Effect of Salting and Packaging Treatments on Fresh Rainbow Trout *(Oncorhynchus mykiss)* Fillets During Storage at Refrigerator Temperatures

Pınar OĞUZHAN * 🖍 Simay ANGİŞ **

* Ardahan University, The Faculty of Engineering, Food Engineering Department, TR-75000 Ardahan - TURKEY ** Ataturk University, The Faculty of Agriculture, Fisheries Department, TR-25240 Erzurum - TURKEY

Makale Kodu (Article Code): KVFD-2011-5633

Summary

In research was investigated for effect of salting and packaging treatments on the chemical and microbiological properties of rainbow trout fillets stored 25 d at $4\pm1^{\circ}$ C and packaged in vacuum or modified atmosphere (50% CO₂ + 50% N₂) and salted (dry salting and brine salting) with 20% NaCl. Fillets were subjected to microbiological (total aerobic mesophilic bacteria, psychrotrophic bacteria, *Pseudomonas*, lactic acid bacteria, Enterobacteriaceae, yeast and mould) and chemical (pH, thiobarbituric acid reactive substances-TBARS, total volatile base nitrogen-TVB-N) analyzes on certain days (0, 5, 10, 15, 20 and 25th days) of storage. TBARS and TVB-N values were established the highest increase in dry salted samples at the end of storage. Modified atmosphere packaging, in terms investigated of all parameters gave better results than vacuum packaging. According to the results of microbiological analyses, shelf life of rainbow trout fillets was estimated as 10, 15, 20, 25, 15 and 20 days for CV, CM, V1, M1, V2 and M2 respectively

Keywords: Rainbow trout, Salting, MAP, Vacuum, Shelf life

Buzdolabı Koşullarında Depolanan Taze Gökkuşağı Alabalığı (Oncorhynchus mykiss) Filetoları Üzerine Tuzlama ve Paketleme İşlemlerinin Etkisi

Özet

Araştırmada %20'lik sodyum klorür ile tuzlanan (salamura ve kuru tuzlama) vakum veya modifiye atmosferde (%50 CO₂ + %50 N₂) ambalajlanıp 4±1°C'de 25 gün depolanan gökkuşağı alabalığı (*Oncorhynchus mykiss*) filetolarının kimyasal ve mikrobiyolojik özellikleri üzerine tuzlama ve paketleme işlemlerinin etkisi incelenmiştir. Filetolar depolamanın belirli günlerinde (0, 5, 10, 15 ve 25. gün) mikrobiyolojik (toplam aerobik mezofilik bakteri, psikrotrofik bakteri, *Pseudomonas*, laktik asit bakterileri, Enterobacteriaceae, maya ve küf) ve kimyasal (pH, TBARS, TVB-N) açıdan analiz edilmiştir. Depolama sonunda TVB-N ve TBARS değerleri açısından en yüksek artış kuru tuzlanmış örneklerde tespit edilmiştir. Tüm parametreler incelendiğinde, modifiye atmosfer paketlemenin vakum paketlemeye göre daha iyi sonuç verdiği saptanmıştır. Mikrobiyolojik analiz sonuçlarına göre gökkuşağı alabalığının raf ömrü CV, CM, V1, M1, V2 ve M2 grubu örneklerde sırasıyla 10, 15, 20, 25, 15 ve 20 gün olarak belirlenmiştir.

Anahtar sözcükler: Gökkuşağı alabalığı, Tuzlama, MAP, Vakum, Raf ömrü

INTRODUCTION

Microorganisms can reproduce in the muscle tissue of aquatic products because these are not only sensitive but also has reach nutrient content. So, the conservation and handling of aquatic products are very important ¹. One of the conservation processes of aquatic products are salting ². Although salting process is itself a preservation technique,

for many processing technology (smoking, drying and marinating processes) is used as a preliminary operation. The main purpose of salting the fish meat is a part of the elimination of water. Bacterial activity is largely prevented in high salt concentrations. High salt concentration prevents the development of microflora causing spoilage in fish. In

+90 478 2111244

^{x20} İletişim (Correspondence)

pinaroguzhan@ardahan.edu.tr

addition, significantly increase the shelf life of fish ³. NaCl is an essential ingredient in processed meat products for its contribution to the water-holding capacity, prevention of microbial growth, reduction of water activity, facilitating the solubilisation of certain proteins and conferring a typical salty taste by enhancing the flavour of such food products⁴. Vacuum packaging method is a type of passive modified atmosphere. After placing the food into the packaging material suitable for this operation, the package is emptied in the air by vacuum and hermetic (air tight) is made a closure. This method is usually used for preservation of meat products. Vacuum packaging in vacuum, albeit a very small amount will remain at O₂. However, the low percentage of O₂ in the package as soon as possible is used by aerobic microorganisms and CO₂ is produced. These types of products, the proliferation of bacteria and oxidation products is prevented as air is not in the package 5-7. While abundant data exists on preservation of fish and fishery products using vacuum packaging, including salmon ⁸⁻¹⁰, ascidia ¹¹, sardine ^{12,13}, trout ¹⁴⁻¹⁶. Modified atmosphere packaging (MAP), is a protecting technique use to extent shelf-life of fish and fish products ¹⁷. Modified atmosphere packaging, elimination of oxygen from inside package and filled with different concentrations of CO₂ and N₂, however, refrigerated storage conditions for aerobic microorganisms, proteolytic bacteria, yeast and mold growth is inhibited ^{6,18,19}. There are a lot of research related to MAP that extension of the shelf life of fish, including chub mackarel ²⁰, cod ^{21,22}, swordfish ²³, rainbow trout ²⁴, salmon ²⁵, bass ²⁶, herring ²⁷, catfish ²⁸. There is not information in the literature regarding to the effects on chemical and microbiological properties of rainbow trout (Oncorhynchus mykiss) fillets prepared with brine method and dry salting method and packaged vacuum or modified atmosphere. The aim of this research is to determine with combined effects of salting (dry salting and brine salting) and packaging (vacuum and MAP) on the shelflife of refrigerated (4°C) rainbow trout fillets by evaluating certain microbiological and chemical parameters.

MATERIAL and METHODS

Preparing Samples

Fish material, rainbow trout (*O. mykiss*) (250±25 g) were obtained from Ataturk University Agricultural College Fisheries Department's rainbow trout breeding and research center. Fish were carried to laboratory and washed with tap water. The fish (72 samples) were eviscerated, stored until rigor had resolved and then filleted, 144 fillets in total ²⁹. Fillets were washed again for removing blood and mucous remains. All filleted samples including the control were packaged in obtained from the firm Südpack Verpackungen GmbH+Co (Germany) 15x25 cm PA/PE (Polyethylene/ Polyamide) (3-seal bags GB 70) in thickness having an O_2 permeability of 40 cm³/m²/day.atm.23°C; N_2 permeability of 24 cm³/m²/day.atm.23°C; CO₂ permeability of 145 cm³/ m²/day.atm.23°C and a water vapour permeability of <3 $g/m^2/day.atm.23$ °C). Fillets, divided into 3 groups. Fresh fish in the first group were evaluated as the control group. The second group were applied at 8°C for 1 h 20% dry salting (NaCl) for each fish. The third group were applied at 8°C for 1 h 20% brine (NaCl) for each fish. Then, fillets were packed by applying vacuum and modified atmosphere.

Treatments included: CV (control samples vacuum packaged) CM (control samples modified atmosphere packaged-50%CO₂ + 50%N₂), V1 (vacuum packaged-dry salted), M1 (modified atmosphere packaged-dry salted), V2 (vacuum packaged-brine) and M2 (modified atmosphere packaged-brine). Each group was included 24 fillets. Rainbow trout fillets stored under refrigeration (4±1°C) and samples were subjected to microbiological (total aerobic mesophilic bacteria, psychrotrophic bacteria, *Pseudomonas*, lactic acid bacteria, Enterobacteriaceae, yeast and mould) and chemical (pH, thiobarbituric acid reactive substances-TBARS, total volatile base nitrogen-TVB-N) analyzes on certain days (0, 5, 10, 15, 20 and 25th days) of storage.

Microbiological Analysis

A sample (25 g) was taken from each trout fillet, transferred aseptically into a stomacher bag containing 225 ml of 0.1% peptone water was added, and the mixture was homogenized for 60 s using a Stomacher (Lab Stomacher Blender 400-BA 7021 Sewardmedical, England) at room temperature. For microbial enumeration, 0.1 ml samples of serial dilutions (1:10, diluent, 0.1% peptonewater) of fish homogenates were spread on petri dish of various agar materials. Total aerobic mesophilic bacteria (TMAB) were enumerated on Plate Count Agar (PCA, Merck 1.05463.0500), and incubated at 30°C for 2 days. Psychrotrophic bacteria were enumerated on Plate Count Agar (PCA, Merck 1.05463.0500), and incubated at 10°C for 7 days. Pseudomonas were enumerated on cetrimide fusidin cephaloridine agar (CFC, Pseudomonas Agar Base-Oxoid CM0559 + CFC Selective Agar Supplement-Oxoid SR0103) and incubated at 25°C for 2 days. Lactic acid bacteria (LAB) were enumerated on de Man Rogosa Sharpe agar (MRS, de Man, Rogosa Sharpe Agar Oxoid CM0361) incubated at 30°C for 2 days. For Enterobacteriaceae (VRBD, Violet Red Bile Dextrose Agar Merck 1.10275.0500) incubated at 30°C for 2 days. Yeast and mould were enumerated on RBC (Rose Bengal Chloramphenicol) Agar (Merck 1.00467.0500) incubated at 25°C for 5 days.

Chemical Analysis

Total volatile basic nitrogen (TVB-N) was determined using the method of Malle and Tao³⁰. TVB-N contents were expressed as mg 100/g fish muscle. Thiobarbituric acid reactive substans (TBARS) was determined according to the method of Lemon³¹ and Kılıç and Richards³². TBARS content was expressed as µmol Malondialdehyde (MDA)/ kg fish muscle. pH was determined according to the method of Gökalp et al.³³.

Statistical Analysis

Experiments were replicated twice on two separate occasions with different fish samples. Analyses were run in duplicate for each replicate. All obtained data from this study were subjected to analysis of variance (ANOVA), and followed by Duncan's multiple range test to determine significant differences among means at $\alpha = 0.05$ level, using by SPSS ³⁴.

RESULTS

Microbiological Changes

Changes in TMAB of refrigerated fresh rainbow trout fillets during storage under vacuum and modified atmosphere packaging are shown in *Fig. 1a*. The initial (day 0) TMAB (*Fig. 1a*) of rainbow trout fillets was 3.12 log cfu/g. CV, CM, V1, M1, V2 and M2 rainbow trout fillets exceeded the value of 7 log cfu/g for TMAB, considered as the upper acceptability limit for fresh marine species ³⁵ on days 10, 15, 20, 25, 15 and 20 of storage, respectively. At the end of storage period CV, CM, V1, M1, V2 and M2 respectively, levels of 10.24, 9.94, 8.11, 7.24, 8.53 and 8.51 log cfu/g were reached.

The initial (day 0) Psychrotrophic bacteria (*Fig. 1b*) of rainbow trout fillets was 3.22 log cfu/g. CV, CM, V1, M1, V2 and M2 rainbow trout fillets exceeded the value of 7 log cfu/g for Psychrotrophic bacteria, considered as the upper acceptability limit for fresh marine species ³⁵ on days 10, 15, 20, 25, 15 and 15 of storage, respectively. At the end of storage period CV, CM, V1, M1, V2 and M2 respectively, levels of 10.60, 9.80, 8.24, 7.61, 8.88 and 8.66 log cfu/g were reached. Inhibiting effect of dry salting method was higher than the brine method on the number of total aerobic mesophilic bacteria and psychrotrophic bacteria of fillets during storage.

The initial (day 0) *Pseudomonas (Fig. 1c)* of rainbow trout fillets was 3.50 log cfu/g. At the end of storage period CV, CM, V1, M1, V2 and M2 respectively, levels of 8.33, 8.10, 6.62, 6.16, 7.26 and 7.16 log cfu/g were reached. Initial counts were 2.0 log cfu/g (LAB) (*Fig. 1d*) and 2.0 log cfu/g (Enterobacteriaceae) (*Fig. 1e*), at the end of storage period populations of LAB (8.46, 8.02, 8.52, 8.85, 7.49 and 7.48 log cfu/g) and Enterobacteriaceae (9.55, 9.0, 5.02, 5.33, 5.12 and 5.05 log cfu/g) were recorded for treatments CV, CM, V1, M1, V2 and M2, respectively.

The initial (day 0) yeast and mould (*Fig. 1f*) of rainbow trout fillets was 2.08 log cfu/g. At the end of storage period CV, CM, V1, M1, V2 and M2 respectively, levels of 5.65, 5.40, 4.27, 3.97, 3.96 and 3.84 log cfu/g were reached.

Chemical Changes

- TVB-Nitrogen: The amount of TVB-N is an important

criteriation in determining freshness of fish and fish products and TVB-N values are increasing in paralel spoilage ³⁶. The initial (day 0) TVB-N values (*Fig. 2a*) of rainbow trout fillets were 12.42 mg/100 g. CV, CM, V2, M1 and M2 rainbow trout fillets exceeded the value of 25 mg/100 g which an upper acceptable limit TVBN value of 25 mg N/100 for rainbow trout was suggested by ³⁷ on days 15 of storage. V1 rainbow trout fillets exceeded the value of 25 mg/100 g on days 10 of storage.

- **Lipid Oxidation:** Oxidative rancidity may become a problem if higher than normal levels of oxygen are used. Rancidity due to oxidation of polyunsaturated fatty acids (PUFA) in some fish may be a problem in modified atmosphere with O_2 ³⁷⁻³⁹.

Initial TBARS values (*Fig. 2b*) were 1.95 µmol MDA/kg. At the end of storage period TBARS values 9.19, 8.37, 10.83, 9.99, 9.81 and 9.06 µmol MDA/kg were recorded for treatments CV, CM, V1, M1, V2 and M2, respectively.

- **pH:** pH value of fish meat usually ranges from 5.7-6.6. Fresh fish is close to neutral pH, after the death with be formed the lactic acid firstly falling and then rising again with spoilage ⁴⁰. pH values of rainbow trout fillets (*Fig. 2c*) were 6.31. At the end of storage period pH values 6.70, 6.39, 6.29, 6.25, 6.34 and 6.33 were recorded for treatments CV, CM, V1, M1, V2 and M2, respectively.

DISCUSSION

Bacterial growth of modified atmosphere packaged samples are lower than vacuum packaged samples because of CO₂ gas is exist in MAP. The carbon dioxide atmosphere can be considered effectively inhibitory on the total aerobic mesophilic and psychrotrophic flora. Similary results were observed by many other researchers ^{16,24,41-47}. The application of MAP has been previously reported to extend the shelf life of herring ²⁷, rainbow trout ³⁷, salmon ⁴⁸, hake ^{49,50}, cod fillets ^{22,51}, Baltic herring fillets ⁵² and eel ⁵³. Inhibiting effect of dry salting method was higher than the brine method on the number of total aerobic mesophilic bacteria and psychrotrophic bacteria of fillets during storage.

Similar initial *Pseudomonas* (day 0) were reported for rainbow trout by Frangos et al.¹⁶, Pyrogotou et al.⁴⁶ and Mexis et al.⁵⁴. *Pseudomonas* spp. and *Shewanella putrefaciens* were early recognised as putative spoilage inducers in fish muscle and have since then been found in various fish species from fresh- and marine waters as well as in other foods ⁵⁵⁻⁵⁷.

The initial LAB and Enterobacteriaceae numbers 2.0 log cfu/g while this values increased during storage time all groups. Similar findings were found for sardine by Can⁴⁷.

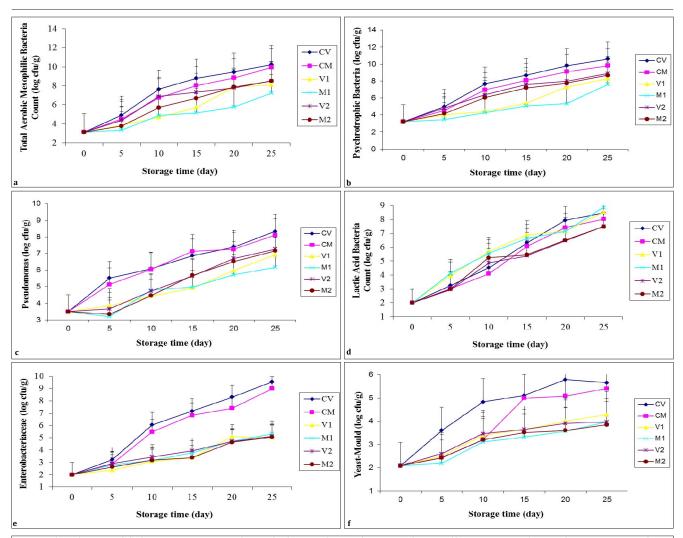


Fig 1. Total aerobic mesophilic bacteria counts (a), psychrotrophic bacteria (b), *Pseudomonas* (c), lactic acid bacteria counts (d), Enterobacteriaceae (e) and yeast and mould (f) changes of salted (brine and dry salting) rainbow trout fillets during cold storage in vacuum and MAP conditions at 4°C Sekil 1. Soğuk depolama boyunca tuzlanmış (salamura ve kuru tuzlama) alabalık filetolarının 4°C' de vakum ve modifiye atmosfer paketleme şartlarında toplam aerobik mezofilik bakteri (a), psikrotrofik bakteri (b), *Pseudomonas* (c), laktik asit bakterileri (d), Enterobacteriaceae (e), mava ve küf (f)'deki değişiklikler

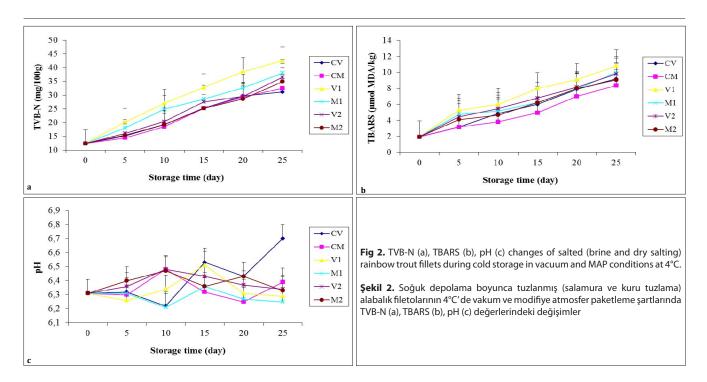
Similar initial yeast and mould (day 0) were reported for rainbow trout by Can ⁴⁷ and mud eel by Vishwanath et al.⁵⁸.

TVB-N may be considered as a quality index for fish. Its increase related to the activity of spoilage bacterai and endogenous enzymes ⁵⁹⁻⁶¹. The initial (day 0) TVB-N numbers 12.42 mg/100 g while this values increased in the duration of storage time all groups. Similarly, TVB-N values have been reported for salmon ⁴¹, red mullet ⁴³, trout ⁶², chub mackarel ⁶³, Atlantic bonito ⁶⁴ and mahi sefid ⁶⁵.

In "perfect material" TBA value should be less than 3 mg malonaldehyde/kg, in "good material" TBA value should not be more than 5 mg malonaldehyde/kg and consumption limit for TBA value is between 7 and 8 mg malonaldehyde/kg. TBA values showed the degree of rancidity in the products, and values greater than 3-4 mg malonaldehyde/kg indicated a loss of product quality ²⁴. Initial TBARS values were 1.95 µmol MDA/kg. Increased TBARS values in the duration of storage time all groups. Similarly, TBARS values have been reported for rainbow trout ²⁴, catfish ²⁸, anchovy ³⁶, mahi sefid ⁶⁵ and whitefish ⁶⁶.

The pH in fresh fish flesh in almost neutral. In the post-mortem period, decomposition of nitrogenous compounds leads to an increase in pH in the fish flesh. The increase in pH indicates the loss of quality ⁴⁷. pH values of rainbow trout fillets were 6.31. Similarly, pH values have been reported for red mullet ⁴³, sardine ⁴⁷, mahi sefid ⁶⁵, sea bream ⁶⁷ and tilapia ⁶⁸. pH values of modified atmosphere packaged control group are lower than vacuum packaged samples. This result is occured due to carbonic acid the conversion of carbon dioxide. However pH values of the brine method was higher than the dry salting method. Similary findings were observed by Bilgin et al.⁴⁰.

The highest TBARS and TVB-N values were established in the dry salted and vacuum packaged samples while the lowest pH value was determined in the dry salted and modified atmosphere packaged samples at the end of storage.



According to microbiological analyses data, the shelf life of CV, CM, V1, M1, V2 and M2 to 10, 15, 20, 25, 15 and 20 days respectively.

The present study concluded that dry salting and modified atmosphere packaging combination can retard total aerobic mesophilic bacteria, psychrotrophic bacteria, *Pseudomonas* of samples but increase lactic acid bacteria and brine salting and modified atmosphere packaging combination can delay Enterobacteriaceae, yeast and mould of samples during refrigerated storage.

REFERENCES

Babadoğan G: Su Ürünleri Sektör Araştırması. İGEME, s. 124, Ankara, 1998.
Bellagha S, Sahli A, Farhat A, Kechaou N, Glenza A: Studies on salting and drying of sardine (*Sardinella aurita*): Experimental kinetics and modeling. *J Food Engine*, 78, 947-952, 2007.

3. Ismail N, Wootton M: Fish salting and drying a review. *ASEAN Food J*, 7 (4): 175-183, 1992.

4. Armenteros M, Aristoy MC, Barat JM, Toldrá F: Biochemical changes in dry-cured loins salted with partial replacements of NaCl by KCl. *Food Chem*, 117, 627-633, 2009.

5. Keles F: Gıda ambalajlama ilkeleri, Atatürk Üniversitesi Ziraat Fakültesi Yayınları No: 189, Atatürk Üniversitesi Ziraat Fakültesi Ofset Tesisi, Erzurum, 1998.

6. Gülyavuz H, Ünlüsayın M: Su Ürünleri İşleme Teknolojisi. Süleyman Demirel Üniversitesi Eğridir Su Ürünleri Fakültesi, İsparta, 1999.

7. Kılınç B, Çaklı Ş: Paketleme tekniklerinin balık ve kabuklu su ürünleri mikrobiyal florası üzerine etkileri. J Fisheries & Aquat Sci, 18 (1-2): 279-291, 2001.

8. Leroi F, Joffraud JJ, Chevalier F: Effect of salt and smoke on the microbiological quality of cold-smoked salmon during storage at 5°C as estimated by the factorial design method. *J Food Prot*, 63 (4): 502-508, 2000.

9. González-Rodríguez MN, José-Javier S, Santos JA, Otero A, García-López MA: Numbers and types of microorganisms in vacuum-packed coldsmoked freshwater fish at the retail level. *Int J Food Microbiol*, 77, 161-168, 2002. **10.** Martinez O, Salerón J, Guillén MD, Casas C: Effect of freezing on the physicochemical, textural and sensorial characteristics of salmon (*Salmo salar*) smoked with a liquid smoke flavouring. *LWT-Food Sci and Technol*, 43, 910-918, 2010.

11. Stamatis N, Arkoudelos J, Vafidis D: Differences in chemical, microbial and sensory quality parameters of the marinated ascidia *Microcosmus sabatieri* Roule, 1885 during storage at 6°C under vacuum conditions. *Int J Food Sci and Technol*, 43, 1705-1713, 2008.

12. Senesi E, Bertolo G, Torreggiani D, Cesare L, Caserio G: The utilization of mediterranean sardines by means of smoking. *Advances in Fish Sci and Technol*, 290-293, 1980.

13. Gómez-Estaca J, Gimènez B, Gómez-Guillén C, Montero P: Influence of frozen storage on aptitude of sardine and dolphinfish for cold-smoking process. *LWT-Food Sci and Technol*, 43, 1246-1252, 2010.

14. Schulze K: Untersuchungen zur Mikrobiologie, Haltbarkeit und Zusammensetzung von Raucherforellen aus einer Aquakultur. *Arch Lebensmittelhyg*, 36 (4): 82-85, 1985.

15. Lyhs U, Björkroth J, Hyytia E, Korkeala H: The spoilage flora of vacuumpackaged, sodium nitrite or potasium nitrate treated, cold- smoked rainbow trout stored at 4°C or 8°C. *Int J Food Microbiol*, 45 (2): 135-142, 1998.

16. Frangos L, Pyrgotou N, Giatrakou V, Ntzimani A, Savvaidis IN: Combined effects of salting, oregano oil and vacuum-packaging on the shelf-life of refrigerated trout fillets. *Food Microbiol*, 27, 115-121, 2010.

17. Özoğul Y, Özoğul F, Küley E: Modifiye edilmiş atmosfer paketlemenin balık ve balık ürünlerine etkisi. *J Fisheries & Aquat Sci*, 23 (1-2): 193-200, 2006.

18. Swiderski F, Russel S, Waszkiewiez-Robak B, Cholewinska E: Evaluation of vacuum-packaged poultry meat and its products. *J Sci Food Agric*, 48, 193-200, 1997.

19. Kılınç B, Çaklı Ş: Su ürünlerinin modifiye atmosferde paketlenmesi. *J Fisheries & Aquat Sci,* 21 (3-4): 349-353, 2004.

20. Erkan N, Özden Ö, Inuğur M: The effects of modified atmosphere and vacuum packaging on quality of chub mackerel. *Int J Food Sci and Technol*, 42, 1297-1304, 2007.

21. Cann DC, Smith GL, Houston NC: Further studies of the packaging of marine fish products Under Modified Atmospheres, Torry Research Station. Abendeen-UK, 1983.

22. Debevere J, Boskou G: Effect of modified atmosphere packaging on the TVB/TMA-producing microflora of cod fillets. *Int J Food Microbiol*, 31 (1-3): 221-229, 1996.

23. Muratore G, Licciardella F: Effect of vacuum and modified atmosphere packaging on the shelf life liquid smoked swordfish (Xiphias gladius). Food

Chem and Toxic, 70, 359-363, 2005.

24. Çaklı Ş, Kılınç B, Dinçer T, Tolasa Ş: Comparison of the shelf lifes of Map and vacuum packaged hot smoked rainbow trout (*Oncorhynchus mykiss*). *Eur Food Res Technol*, 224, 19-26, 2006.

25. Paludan-Müller C, Dalgaard P, Huss HH, Gram L: Evaluation of the role of *Carnobacterium piscicola* in spoilage of vacuum and modified atmosfere packed cold smoked salmon stored at 5°C. *Int J Food Microbiol*, 39, 155-166, 1998.

26. Torrieri E, Cavella S, Villani F, Masi P: Influence of modified atmosphere packaging on the chilled shelf life of gutted farmed bass (*Dicentrarchus labrax*). J Food Engine, 77, 1078-1086, 2006.

27. Lyhs U, Lahtinenb J, Schelvis-Smit R: Microbiological quality of maatjes herring stored in air and under modified atmosphere at 4 and 10°C. *Food Microbiol*, 24, 508-516, 2007.

28. Göktepe I, Moody MW: Effect of modified atmosphere packaging on the quality of smoked catfish. *J Muscle Foods*, 9, 375-389, 1998.

29. Robb DHF, Kestin SC, Warriss PD, Nute GR: Muscle lipid content determines the eating quality of smoked and cooked Atlantic salmon (*Salmo salar*). *Aquat*, 205, 345-358, 2002.

30. Malle P, Tao SH: Rapid quantitative determination of trimethylamine using steam distillation. *J Food Prot*, 50 (9): 756-760, 1987.

31. Lemon DW: An Improved TBA Test for Rancidity, New Series Circular, May 8, No:51, Halifax Laboratory, Halifax Mova, 1975.

32. Kılıç B, Richards MP: Lipid oxidation in poultry döner kebabi. Prooxidative and anti-oxidative factors. *J Food Sci*, 68 (2): 690-696, 2003.

33. Gökalp HY, Kaya M, Zorba Ö, Tülek Y: Et ve ürünlerinde kalite kontrolü ve laboratuar uygulama kılavuzu, Atatürk Üniversitesi Ziraat Fakültesi Yayın No: 318, Ders Kitabı: 69. Erzurum, 1999.

34. SPSS: SPSS for windows release 10.0, SPSS Inc. Chicago-USA, 1999.

35. ICMSF: International commission on microbiological specifications for foods. Sampling plans for fish and shellfish. **In**, ICMSF, Microorganisms in Foods. Sampling for Microbiological Analysis: Principles and Scientific Applications. Toronto, Ontario, Canada: University of Toronto Press, 1992.

36. Köse S, Koral S: Tütsülenmiş hamsinin buzdolabı koşullarında (4±1°C) depolanması esnasında kalite değişimlerinin belirlenmesi, *XIII. Ulusal Su Ürünleri Sempozyumu*,1-4 Eylül, Çanakkale, 2005.

37. Aras Hisar Ş, Hisar O, Kaya M, Yanık T: Effects of modified atmosphere and vacuum packaging on microbiological and chemical properties of rainbow trout (*Oncorhynchus mykiss*) Fillets. *Int J Food Microbiol*, 97 (2): 209-214, 2004.

38. Finne G: Modified and controlled atmosphere storage of muscle foods. *Food Technol*, 36 (2): 128-133, 1982.

39. Stammen K, Gerdes D, Caporaso F: Modified atmosphere packaging of seafood. *Critical Reviews in Food Sci and Technol*, 29, 301-331, 1990.

40. Bilgin Ş, Ertan OÖ, Günlü A: The effects on chemical composition of *Salmo trutta macrostigma* Dumeril, 1858 of different salting techniques. *J Fisheries & Aquat Sci*, 24 (3-4): 225-232, 2007.

41. Leroi F, Joffraud JJ, Chevalier F, Cardinal M: Study of the microbial ecology of cold-smoked salmon during storage of 8°C. *Int J Food Microbiol*, 39, 111-121, 1998.

42. Dondero M, Cisternas F, Carvaja L, Simpson R: Changes in qulity of vacuum-packed cold-smoked salmon (*Salmo salar*) as a function of storage temperature. *Food Chem*, 83, 543-550, 2004.

43. Gümüş B, İkiz R, Ünlüsayin M, Gülyavuz H: Quality changes of salted red mullet (*Mullus barbatus* L., 1758) during vacuum packaged stored at +4°C. *J Fisheries & Aquat Sci*, 25 (2): 101-104, 2008.

44. Kilinc B, Cakli S, Dincer T, Tolasa S: Microbiological, chemical, sensory, color, and textural changes of rainbow trout fillets treated with sodium acetate, sodium lactate, sodium citrate, and stored at 4°C. *J Aquat Food Product Technol*, 18, 3-17, 2009.

45. Erkan N: The Effect of thyme and garlic oil on the preservation of vacuum-packaged hot smoked rainbow trout (*Oncorhynchus mykiss*). Food Bioprocess Technol, 1-9, 2010.

46. Pyrgotou N, Giatrakou V, Ntzimani A, Savvaidis IN: Quality assessment of salted, modified atmosphere packaged rainbow trout under treatment with oregano essential oil. *J Food Sci*, 75 (7): 406-411, 2010.

47. Can ÖP: Combine effect of potassium sorbate and dry salting on the shelf life sardine (*Sardina pilchardus*). *J Food Tech*, 9 (1): 43-49, 2011.

48. Pastoriza L, Sampedro G, Herrera JJ, Cabo ML: Effect of carbon dioxid atmosphere on microbial growth and quality of salmon slices. *J Sci Food Agric*, 72, 348-352, 1996.

49. Pastoriza L, Sampedro G, Herrera JJ, Cabo ML: Effect of modified atmosphere packaging on shelf-life of iced fresh hake slices. *J Sci Food Agric*, 71, 541-547, 1996.

50. Pastoriza L, Sampedro G, Herrera JJ, Cabo ML: Influence of sodium chloride and modified atmosphere packaging on microbiological, chemical and sensorial properties in ice storage of slices of hake (*Merluccius merluccius*). *Food Chem*, 61 (1-2), 23-28, 1998.

51. Lauzon HL, Magnússon H, Sveinsdóttir K, Gudjónsdóttir M, Martinsdóttir E: Effect of brining, modified atmosphere packaging and superchilling on the shelf life of cod (*Gadus morhua*) loins. *J Food Sci*, 74 (6): 258-267, 2009.

52. Randell K, Hattula T, Ahvenainen R: Effect of packaging method on the quality of rainbow trout and baltic herring fillets. *LWT*, 36, 56-61, 1997.

53. Arkoudelos J, Stamatis N, Samaras F: Quality attributes of farmed *eel (Anguilla anguilla)* stored under air, vacuum and modified atmosphere packaging at 0°C. *Food Microbiol*, 24, 728-735, 2007.

54. Mexis SF, Chouliara E, Komtominas MG: Combined effect of an oxygen absorber and oregano essential oil on shelf life extension of rainbow trout fillets stored at 4°C. *Food Microbiol*, 26, 598-605, 2009.

55. Castell CH, Greenough MF: The action of *Pseudomonas* on fish muscle: I. Organisms responsible for odour produced during incipient spoilage of chilled fish muscle. *J Fish Res Board Can*, 16, 13-19, 1957.

56. Macdonell MT, Colwell RR: Phylogeny of the Vibrionaceae and recommendation for two new genera, *Listonella and Shewanella*. *Syst Appl Microbiol*, 6, 171-182, 1985.

57. Reynisson E, Lauzon HL, Magnússon H, Jónsdóttir R, Ólafsdóttir G, Marteinsson V, Hreggviðsson GÓ: Bacterial composition and succession during storage of North-Atlantic cod (*Gadus morhua*) at superchilled temperatures. *BMC Microbiol*, 9, 1-12. 2009.

58. Vishwanath W, Lilabati H, Bijen M: Biochemical, nutritional and microbiolgical quality of fresh and smoked mud eel fish *Monopterus albus*-a comparative study. *Food Chem*, 61 (1/2): 153-156, 1998.

59. Özoğul Y, Polat A, Özoğul F: The effect of modified atmosphere packaging and vacuum packaging on chemical, sensory, and microbiological changes of sardines (*Sardina pilchardus*). *Food Chem*, 85, 49-57, 2004.

60. Ruiz-Capillas C, Moral A: Sensory and biochemical aspects of quality of whole big eye tuna (*Thunnus obesus*) during bulk storage in controlled atmospheres. *Food Chem*, 89, 347-354, 2005.

61. Öksüztepe G, Emir Çoban Ö, Güran Ş: Sodyum laktat ilavesinin taze gökkuşağı alabalığından (*Oncorhynchus mykiss*) yapılan köftelere etkisi. *Kafkas Univ Vet Fak Derg,* 16 (Suppl-A): S65-S72, 2010.

62. Zorn VW, Greul E, Kramez J: Beurteilung des hygienestatus geraucherter, vakuumverpackter forellenfillets. *Arch Lebensmittelhyg*, 44, 81-104, 1993.

63. Goulas AE, Kontominas MG: Effect of salting and smoking-method on the keeping quality of chub mackarel (*Scomber japonicus*): Biochemical and sensory attribuetes. *Food Chem*, 93, 511-550, 2005

64. Duyar HA, Erdem ME, Samsun S, Kalaycı F: The effects of the different woods on hot-smoking vacuum packed Atlantic Bonito (*Sarda sarda*) stored at 4°C. *J Anim Vet Adv*, 7 (9): 1117-1122, 2008.

65. Zolfaghari M, Shabanpour B, Fallahzadeh S: Quality preservation of salted, vacuum packaged and refrigerated mahi sefid (*Rutilus frisii kutum*) fillets using an onion (*Allium cepa*) extract. *Aquat Res*, 41, 1123-1132, 2010.

66. Cuppett SL, Gray JI, Booren AM, Price JF, Stachiw MA: Effect of processing variables on lipid stability in smoked great lakes whitefish. *J Food Sci*, 54 (1): 52-54, 1989.

67. Goulas AE, Kontominas MG: Combined effect of light salting, modified atmosphere packaging and oregano essential oil on the shelf-life of sea bream (*Sparus aurata*): Biochemical and sensory attributes. *Food Chem*, 100, 287-296, 2007.

68. Chaijan M: Physicochemical changes of tilapia (*Oreochromis niloticus*) muscle during salting. *Food Chem*, 129, 1201-1210, 2011.