

The Role of Oxidant and Antioxidant Parameters in the Infectious Diseases: A Systematic Literature Review

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Abstract

The formation of reactive oxygen species is a physiological event in aerobic life. In case of infection, if the increased oxidant substances cannot be cleaned sufficiently by antioxidants, oxidative stress occurs. As a result, a number of pathological problems occur by damaging DNA, protein, carbohydrate and lipids. In biological systems, the balance between oxidants and antioxidants is important. In oxidative stress situations where the balance cannot be achieved sufficiently, support can be provided with the use of exogenous antioxidants. However, the molecular structure, route of administration and concentrations of these exogenous antioxidants are important. Otherwise, they may show a pro-oxidative effect. The present review study makes a general overview of oxidative-nitrosative stress markers commonly used in infective clinical studies, antioxidant enzymes and parameters and antioxidant supplements.

Keywords: *Infectious diseases, Oxidant, Antioxidant, Oxidative stress*

Enfektif Hastalıklarda Oksidan ve Antioksidan Parametrelerin Rolü: Sistemik Bir Literatür Değerlendirmesi

Öz

Reaktif oksijen türlerinin oluşumu arerobik yaşamda fizyolojik bir olaydır. Bu serbest radikal türlerinin daha fazla üretilmesine neden olan paraziter, bakteriyel ve viral enfeksiyonlarda antioksidan sistemin kapasitesi bu reaktifleri yeterince temizleyemez ise oksidatif stres gelişir. Sonuçta DNA, protein, karbonhidrat ve lipidlerde hasar oluşarak bir takım patolojik problemler meydana gelir. Biyolojik sistemlerde oksidan ve antioksidanlar arasındaki denge önemlidir. Dengenin yeterince sağlanamadığı oksidatif stres durumlarında bazan eksojen antioksidan kullanımı ile destek sağlanabilir. Ancak bu eksojen antioksidanların molekül yapısı, veriliş yolu ve konsantrasyonları önemlidir. Aksi takdirde pro-oksidatif etki gösterebilirler. Bu çalışma ile yaygın olarak görülen enfektif klinik çalışmalarda kullanılan oksidatif-nitrozatif stres belirteçlerinin, antioksidan enzim ve parametreleri ile ileriye yönelik olarak yapılacak bazı antioksidan supplementlerinin genel bir durumu değerlendirilmiştir.

Anahtar sözcükler: *Enfeksiyöz hastalıklar, Oksidanlar, Antioksidanlar, Oksidatif stres*

INTRODUCTION

Free radicals are high energy atoms or molecules that contain one or more unpaired electrons in their outer orbitals. Nitric oxide (·NO), nitrogen dioxide (NO₂·), superoxide (O₂⁻·), hydroxyl (·OH), lipid peroxy (LOO·), peroxy (ROO·) and alkoxy (RO·) radicals can be given as examples^[1,2]. Of these free radicals, those originating from oxygen area called reactive oxygen species (ROS) and those originating from nitrogen are called reactive nitrogen species (RNS)^[2]. Under physiological conditions, the most important source of intracellular reactive products is mitochondria. These reactive products are produced in large quantities in neutrophils, macrophages and monocytes. In addition to being produced

with mitochondrial electron transfer system (ETS) chain and phagocyte activation, endogenously produced free radicals are also released as a result of the activity of many enzymes such as xanthine oxidase (XOD), nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, neutrophil myeloperoxidase (MPO), cyclooxygenase, lipoxxygenase^[3-6]. Sources of exogenous free radicals (ROS) are UV, X-ray, gamma and microwave rays, air pollutants such as asbestos, benzene, carbon monoxide, formaldehyde, ozone and toluene, chemicals such as cleaning products, glue, paint, thinner, perfumes and pesticides, sudden and excessive oxygen entry, medical hyperbaric oxygen exposure, increase in catecholamines, increased lactic acid in muscles and blood, elevation in lytic enzyme activities such as lactate



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dehydrogenase, creatine phosphokinase, different and difficult physiological conditions such as exercise, pregnancy and old age, intense stress, cigarette and alcohol use, large amounts of and long-term consumption of diets containing polyunsaturated and easily peroxidable fats. Antioxidant system deficiencies or exceeding the defensive wall disrupts the oxidant-antioxidant balance in favour of oxidants and oxidative stress occurs [7-10]. Because reactive species have very short shelf life and difficult measurement, NO, lipid peroxidation marker malondialdehyde (MDA), total oxidant capacity (TOC), which is easy to measure with kits and 8-hydroxydeoxyguanosine as the marker of ROS in DNA oxidation are the most used parameters [11-14].

A large number of studies have been published on oxidative stress markers in the field of veterinary science. This review makes a general overview of oxidative-nitrosative stress markers commonly used in infective clinical studies, antioxidant enzymes and parameters and antioxidant supplements to be made prospectively.

PHYSIOLOGICAL STATES OF RADICALS

In addition to being harmful, free radicals also fulfil some physiological functions depending on their concentrations. For example, NO is an important mediator in the relaxation of smooth muscles and regulation of microcirculation as an intracellular messenger; additionally in addition, free radicals are used in vascular tone and signal transmission, protein phosphorylation, transcription factor activation, cell differentiation, apoptosis, oocyte maturation, embryonic development, steroidogenesis, spermatogenesis, pregnancy and immune defence system [15-20]. NO leukocyte adhesion, which is used by endothelial cells, is necessary for leukocyte adhesion, platelet aggregation, angiogenesis, thrombosis and for vascular smooth muscles to regulate blood pressure. In addition to these, NO produced by neurons is an important transmitter substance and has a key in role for neural plasticity. On the other hand, NO produced by macrophages is an important mediator to create an immune response. Superoxide and H₂O₂ can act like second messengers. NO is a molecule that has significant cytotoxic effect in defence against tumour cells, parasite, fungi, protozoa, helminths and mycobacteria, but it is not effective against extracellular pathogens. O₂⁻ can stimulate collagen synthesis through fibroblast proliferation, H₂O₂ has a role in NFκB activation. They can be said to have important roles on cellular signals such as killing cancer cells with cytotoxic lymphocytes and macrophages during phagocytosis in viral, parasitic and microbial infections; detoxification of xenobiotics with p450; activation of ROS and RNS nuclear transcription factors, Ca release from intracellular stores, tyrosine phosphorylation amino acid; activation of non-receptor tyrosine kinase and activation of some cytokines and growth factor signals. In addition to being produced with phagocyte activation, mitochondrial ETS chain also occurs

as a result of the activation of many enzymes such as XOD, NADPH oxidase, MPO, cyclooxygenase, lipoxygenase. Apart from these, ROS participates in the biosynthesis of molecules such as prostaglandin and thyroxin and stimulates the development of these processes. In luteal phase in the oestrus cycle and in follicular (oestrus) period, oxidation level is high. Reactive oxygen species are dissolved in cell and are used in the regulation of guanylate cyclase activity and in vital activities such as gene transcription [2,21-30].

DISEASES AND SYNDROMES REPORTED TO BE CAUSED BY OXIDATIVE DAMAGE

There are a large number of studies reporting that oxidative stress markers are the basis of many diseases. MDA, hydroxy-2-nonenal (HNE), 2-propenal (acrolein), isoprostanes, oxide glutathione (GSSG), NO, total oxidant capacity (TOC), 8-hydroxy-2-deoxyguanosine (8-OHdG) are among the most studied oxidative markers clinically for this purpose. Although oxidative stress has been studied with too many diseases and syndromes, it has been reported to be effective in the development of cancer, cardiovascular disease, neural disorders, Alzheimer's disease, mild cognitive disorder, Parkinson's disease, alcohol-related liver disease, ulcerative colitis, aging and atherosclerosis, lead poisoning, liver damage due to carbon tetrachloride, aminoglycoside, reactions from drugs and toxins, such as heavy metal toxicity, chronic and degenerative diseases such as glomerulonephritis, emphysema, porphyria, bronchopulmonary dysplasia, atherosclerosis, pancreatitis, rheumatoid arthritis, aging, neurodegenerative disorders, hemolytic anemia, cardiovascular diseases, pneumonia, sepsis, mastitis, metritis, retentio secundinarum, genital tract inflammation, acidosis, tongue-playing, sepsis, mastitis, ketosis, enteritis, respiratory, joint diseases and autoimmune disorders [14,26,28,31-43].

ROS FORMATION IN INFECTIOUS CONDITIONS

In many animal studies, the common picture in the clinical biochemistry of parasitic, microbial or viral diseases is the increase in oxidant parameters such as ROS and NO in the cell in order to fight the infectious agent. An increase in MDA level has been reported as a result of lipid peroxidation with the increase in cell damage. Although ROS production is useful in removing invasive pathogens, its excessive and prolonged production can cause permanent damage to host and non-infected cells. ROS production can affect the pro-inflammatory response of inflammatory cell significantly. Intracellular ROS formation by NADPH oxidase triggers pro-inflammatory cytokine production in macrophages, neutrophils and microglia. Free radicals are particularly known to be effective in the last step of phagocytosis, that is, in the step of killing microorganisms. The rapid production of free radicals by a mechanism induced by

inflammation and known as Respiratory Burst causes oxidative stress and cell damage. During this event, O_2 consumption in phagocytic cells increases 4 to 100 times. Activated phagocytic cells (neutrophil, eosinophil and all types of macrophages) produce $O_2^{\cdot-}$ with NADPH oxidase. This function is important in cleaning phagocytosed bacteria. There are five mechanisms under the control of the oxidative explosion;

Endogenous GTPase limits NADPH oxidase activation,

Lactoferrin in phagocyte granules binds free iron,

Phagocytes have self-protective antioxidant systems such as superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase; dense taurine substance is thought to neutralize hypochlorous acid (HOCl),

Some cells sacrifice themselves apoptosis,

Liquids and cells surrounding the target have protective systems [30,44].

When cells are activated, as a result of the activation of macrophages against the agent, chemotactic chemokine such as tumour necrosis factor- alpha (TNF- α), interleukin-6 (IL-6), interferon gamma (IFN- γ), interleukin-8 (IL-8), monocyte chemo attractant protein-1 (MCP-1), macrophage inflammatory protein 2 (MIP-2) are released. As an early response to inflammation, they can cause an abnormal persistent cascade known as cytokine storm as a result of excessive release of inflammatory cytokines. $O_2^{\cdot-}$ is released as a result of the reaction of NADPH oxidase enzyme with respiratory burst in order to eliminate the pathogen in the pathogenesis of the infection. $O_2^{\cdot-}$ radical formed as a result of reaction is converted into H_2O_2 , a cytotoxic molecule catalysed by SOD. The most toxic radical, hydroxyl radical (HO) is produced as a result of H_2O_2 and $O_2^{\cdot-}$ reaction (Heber-Weiss reaction) or H_2O_2 and Fe^{2+} reaction (Fenton reaction), while hypochlorous acid is produced as a result of the reaction of H_2O_2 and chlorine (Cl) catalysed by the enzyme myeloperoxidase. This acid produced forms a part of the antimicrobial defence by destroying bacterial DNA and causing DNA replication to stop. The events that characterize leukocyte activation are rapid increase in oxygen use, increase in glycogenolysis and glucose oxidation and rapid formation of ROS. Especially neutrophils and monocytes form highly reactive oxidants by using the H_2O_2 -MPO-Halid system. Chloride, bromide, iodide and thiocyanate can be used as substrate in this system; however, due to its *in vivo* concentrations, chloride (Cl) appears to be the most preferred halide. Highly reactive hypochlorous porphyrins react rapidly with a large number of molecules such as proteins and amines and kill microorganisms with halogenation, protein, lipid peroxidation reactions [30,45-51].

OXIDATIVE STRESS IN PARASITIC INFESTATIONS

Parasites cause tissue and cell damage in the host as they increase the amount of free radical and lipid peroxidation

in the tissues, organs and cells they settle. It has been reported that following *Nippostrontronylus brasilliensis*, *Trichinella spiralis* and *Dictyocaulus viviparus* infestation, MDA level increases significantly in the lung tissue of the host and that this increase may have been caused by the damage of free radicals to various cell components during infection. Increased lipid peroxidation has been associated with lung lesions due to parasite and oxidative stress caused by respiratory distress that occurs as a result of these lesions [52-54].

It is known that cellular and humoral immune response occurs against cystic echinococcosis, T-lymphocytes play an important role in the immunological control of parasite and macrophage and neutrophils fight metasestodes [55]. NO produced by macrophages is an important mediator to create immune response. In addition, superoxide and hydrogen peroxide increase in macrophage and neutrophils due to XO, MPO and NADPH oxidase activity, which causes an increase in oxidant production [56]. Lipid peroxidation caused by ROS causes cell membrane disruption and ultimately necrotic deaths [57]. In studies conducted on patients infected with liver cystic echinococcosis, an increase was found in MDA levels, while a decrease was found in antioxidant levels; thus, oxidative stress was reported [55-61]. In addition, in the study conducted by Heidarpour et al. [59], the decrease in albumin level was associated with its being used as free radical scavenger in oxidative process. In lung infections, an increase occurs in TOC level, while a decrease occurs in TAC level and has in OSI index [60,62].

Similarly, oxidative stress increase was found in *Toxocara vitulorum*, *Eimeria spp* and coccidiosis, *Nematodiasis*, *Leishmaniasis*, *Hypoderma spp.*, *Toxoplasma gondii*, *Eimeria*, *Anaplasmosis*, *Theileriosis*, *Babesiosis*, *Cryptosporidium parvum* infections in other parasitic diseases involving intestines, kidneys and blood, etc. [12,13,63-72]. In Helminth infestations, these are fought thanks to oxidizing enzymes of increased eosinophil [73]. When studies conducted were examined, the common point found was increased oxidation in parasitic infestations, and increased oxidative stress as a result of decreased antioxidative defence. Specifically, the use of oxidant-antioxidant parameters in differential diagnosis is difficult because oxidative stress is similarly increased in the pathogenesis of many parasitic infestations. On the contrary, these parameters can be used to get information about the healing success or clinical course of the disease.

OXIDATIVE STRESS IN BACTERIAL INFESTATIONS

Although there are different types of microorganisms causing respiratory diseases, studies conducted on beef cattle with microorganisms such as *Actinomyces spp.*, *Staph. epidermidis*, *Corynebacterium spp.*, *Pasteurella multocida*, *Manheimia haemolytica*, *S. aureus* and *Escherichia coli* have reported increased oxidative stress [74]. In studies conducted with different types and biological materials, similar to

bacterial pneumonia, increase was found in serum, broncho-alveolar fluid, while decrease was found in antioxidant system. *M. bovis* increases neutrophil apoptosis and ROS production, while it decreases NO production [30,75-77]. High oxidative stress index in brucella can be used in showing the severity of inflammation [67,78]. Paratuberculosis, one of the inflammatory bowel diseases of the digestive system, *Mycobacterium avium* subtype paratuberculosis (MAP) infections in the aetiology of Johne and Crohn disease are frequently encountered [79]. In the pathophysiology of such inflammatory bowel diseases, high production of ROS and RNS species is closely associated with decreased antioxidant activity and oxidative stress shaped with increased glutathione peroxidase (GPx) activity. In addition, in nonsteroidal anti-inflammatory drugs, it signals apoptosis with oxidative stress through mitochondrial pathway. As a result of increased oxidative stress, the functions of fatty acids and proteins in cell membrane deteriorate, even causing DNA damage and mutations. Another factor contributing to these inflammatory diseases is T-helper cells releasing high amount of interferon (Th-1, Th-2). With the deterioration of epithelial barrier, intestinal permeability increases and inflammation becomes uncontrolled [1,80-82]. While an increased picture is seen in studies conducted in general on infective diseases due to an increase in oxidative parameters and a decrease in antioxidant activity, Johne and Crohn reported increased oxidation and increased GPx activity unlike other studies [83,84]. In Crohn's aetiology, there is no change in GPx activity in case of absence of MAP, while typical finding in those with MAP is an increase in GPx activity. It has been reported that the consistent correlation between MAP infection and GPx activity can potentially be used to find out the MAP infection status.

In a study conducted on calves with septicaemia caused by *E. coli*, while no change was reported in MDA and albumin levels, increase was reported in SOD and GPx activity. The absence of increase in MDA was attributed to the important increase in bilirubin level [85]. Negative correlation was reported between MDA and hyperbilirubinemia [86].

OXIDATIVE STRESS IN VIRAL INFECTIONS

Parvovirus infection in dogs has been stated to be significantly associated with oxidative stress and reactive oxygen/nitrogen species, lipid peroxidation and poor antioxidant reserve. In a study conducted by Aydoğdu et al. [87] TOS was increased, while no change was seen in TAS and naturally OSI index was high. There are also situations in which increase was detected in SOD and GPx activity with oxidants such as MDA, H₂O₂ and decrease in CAT activity [88]. In sheep infected with sheeppox virus, an increase was found in MDA, while a decrease was found in GSH and albumin [89]. This decrease in albumin results from suppression of the antioxidative system and negative acute phase reactant due to infection.

In studies conducted on bulls, cattle and sheep with Foot-and-mouth disease (FMD), the common point is the increase in MDA, NO, TOC level and decrease in GSH and TAC level [90-94]. Researchers have reported that oxidant stress is strong in such a viral disease. In viral infectious diseases, even if the disease changes, results are similar in oxidative stress markers. In malignant catarrhal fever (MCF) disease seen in cattle, while MDA and NO levels were increased GSH was decreased [95,96]. In the pneumonia table in goats, MDA was increased, while SOD, GPx, GSH were decreased [97]. Low level of GSH causes higher ROS production, resulting in unbalanced immune response, inflammation and susceptibility to infection.

Increased RNS production in viral infections is increased through induction of inducible NO synthase enzyme. Cytokines related to Th1 trigger ROS/ RNS production in tissues in infected host tissues. Imbalance in ROS/RNS production and its removal results in oxidative/nitrosative stress that can increase virus replication and the mutation rate of viral RNA, resulting in increased damage to host tissues [98-101]. In viral hepatitis, the main mechanism that triggers cell death is reactive oxygen species formed against pathogen in neutrophil and kupffer cells. ROS increase causes an increase in collagen production by stimulating profibrinogenic cytokines such as TGF-β, PDGF and the regulation of collagen gene transcription in fibroblasts, which is involved in the pathogenesis of the formation of fibrosis. Increased 8-OHdG with infection can be used as a marker in hepatocarcinogenesis [102-104]. While a decrease has been reported in GPx and SOD activity in Herpes virus infections, increase in GPx activity and suppression of SOD activity has been reported in BHV-1 infections [96,102-106]. Since an increase in MDA and NO and decrease is seen in GSH in zoonotic viral diseases such as ecthyma contagiosum, oxidative stress can be mentioned in the pathogenesis [107].

We have stated that the weapon used by the body to phagocytize the pathogen as a defence system to prevent parasitic, bacterial and viral infections from entering the cell and infecting the host is free radicals. The common point in various diseases mentioned above was the increase of oxidants in biological fluids such as serum, tissue, etc. and increase in oxidative stress as a result of the deficiency or suppression in the antioxidative defence system. Oxidative stress (OS) occurs when the antioxidant defence system is insufficient against free radical products. If this stress continues excessively and for a long time, lipid, protein and DNA modifications are seen. Oxidative damage to DNA can form base or sugar lesions of DNA, single and double-strand breaks, abasic regions, DNA-protein or cross-linking between strands. Damaged nucleosides accumulate in both nuclear and mitochondrial DNA. Disruption of this redox balance triggers cell signal change, causing loss of basic cellular functions, tissue inflammation, ageing, apoptosis and ultimately tissue damage [2,6,108-113].

ANTIOXIDANTS

High doses of oxidants used clinically to clarify the pathogenesis of the disease cause pathophysiological changes, especially in parasitic, bacterial and viral infective conditions. The defence system developed by the organism to decrease oxidative stress in the face of this increased oxidation is antioxidants. While endogenous antioxidant enzymes form the superoxide dismutase, catalase, glutathione peroxidase and glutathione reductase (GR) enzymatic defence line; endogenous protective antioxidants are non-enzymatic antioxidant defence systems such as glutathione (GSH), uric acid, melatonin, bilirubin, coenzyme Q10, albumin, α -lipoic acid, lactoferrin, ferritin, haptoglobin, ceruloplasmin, transferrin. Cleansing or chain breaking antioxidants taken exogenously are lipid or water soluble materials such as tocopherol, carotenoids, ascorbate, urate. Besides, zinc, selenium, lycopene, lutein, zeaxanthin, chlorogenic acids, gallic acid, caffeic acid, quercetin, kaempferol, myricetin, naringenin, eriodictyol, hesperetin, luteolin and apigenin are also antioxidants that can be taken in the organism exogenously. These react with free radicals before living structures, take this burden on them and form a product that cannot sustain the harmful reaction. These antioxidant defence systems are parameters used in determining oxidative stress index in serum, plasma, tissue and urine. CAT, GSH, GPx, GR activities and total antioxidant capacity (TAC), ceruloplasmin, albumin, bilirubin parameters are frequently measured in clinical studies as antioxidant markers. Exogenous antioxidants and endogenous antioxidants work together to maintain redox balance [2,6,114,115]. Deficiencies in advanced age caused by insufficient intake or excessive consumption of antioxidant vitamins disrupt immune regulation [116]. As with inhibition by Vit-C or regeneration of Vit-E by GSH, endogenous and exogenous antioxidants work together to maintain redox balance. XO activity, which is accepted as the primary source of ROS, is inhibited by some flavonoids such as quercetin, silibin and luteolin. The most studied polyphenols in clinical infective animal studies due to both their anti-inflammatory and antioxidant effects are resveratrol, quercetin, rutin, tangeretin, homoplantagin, ellagic acid, catechin, apigenin [117-119]. While flavonoids decrease peroxidase activity, they inhibit the release of free radicals by neutrophils and the activation of these cells by α 1-antitrypsin [120]. Anti-inflammatory activities of polyphenols such as quercetin, rutin, morin, hesperetin and hesperidin have been reported in acute and chronic inflammation of animal models. For example, rutin is only effective in chronic inflammatory processes, especially in arthritis, while flavones are effective in the neurogenic inflammation caused by xylene. It has been reported that inflammatory reaction induced by LPS injection can be modulated with daidzin, glycidyl, genistein and their glycosides. Also, exogenous antioxidants such as polyphenols can affect enzymatic activity such as protein kinase and signalling systems during inflammation process,

and these enzymes play a role in cell activation processes such as cell proliferation, B lymphocyte activation or cytokine production by stimulated monocytes. They also significantly inhibit the release of arachidonic acid from the cell membrane [117,121,122]. *In vivo* and *in vitro* studies conducted show that plant derived polyphenol molecules have anti-genotoxic and anti-cytotoxic effect on cells exposed to oxidative stress [123]. Again, the ability of quercetin and naringenin to inhibit cytochrome P450 enzymes which include bioactivation of chemical carcinogens constitutes the other chemopreventive mechanism of polyphenols against cancer development, including lung cancer [124]. Exogenous antioxidants also show pro-oxidant activity in high concentrations and especially in the presence of metal ions such as iron and copper. Pro-oxidative effects of polyphenol compounds such as quercetin, catechin and gallic acids, the antioxidant properties of which are known and emphasized in previous studies, have also come to the fore in recent studies. It has been shown that prooxidant activity in flavonoids is associated with the number of hydroxyls in the molecule and flavonoids containing more than three OH in B group increase the production of hydroxyl radicals [2,119,125-128]. For this reason, molecular structures and concentrations of antioxidant substances to be used in studies should be well determined.

ANTIOXIDANTS IN INFECTIOUS ANIMAL DISEASES

If considered in a broad sense, it can be thought that antioxidant stress resulting from the excessive or inappropriate inflammatory response in severe-fulminant infections and complications of chronic infections may be at the centre of events that harm the patient or even lead to death to overcome the disease. In which infections, duration and doses can supplemental antioxidant treatment be used for the benefit of the host? There are many unclear issues about how these treatments will affect the immune response. Therefore, it may be recommended to study the effects of antioxidant supplements first *in vitro* and then start *in vivo* studies to clarify the situation in the whole system.

ANTIOXIDANT USE IN PARASITIC INFESTATIONS

Studies on the use of antioxidant supplements in parasitic infestations are more limited compared to viral and bacterial infections. The nephrotoxic effect occurs as a side effect during the application of first generation platinum containing cisplatin, which is mostly used as an antineoplastic for the treatment of Leishmaniasis. The application of antioxidant complex (Vit-C, Vit-E, silibinin) to reduce nephrotoxicity has been described as a promising study because of the decrease in the parasite load and toxic effect [129]. Ram et al. recommend the use of copper, manganese, selenium and zinc injections in newborn calves with theileriosis since they help improve immunological

imbalance [130]. Similarly, it is stated that N-acetyl-L-cysteine, which is known to have antioxidant activity, can be used as a drug in the treatment of babesiosis [131]. Vit-E and selenium application can be used as an adjunct therapeutic agent to regulate intravascular hemolysis caused by oxidative stress in babesiosis in cattle [132].

ANTIOXIDANT USE IN BACTERIAL INFECTIONS

In the samples of the infectious studies that we found in our review, oxidative stress has been reported in the pathogenesis of patients with sepsis and it has been suggested mitochondrial dysfunction may be a causal factor in the development of multiple organ failure. This is because ROS, which increases as a result of oxidative stress in the cell, simultaneously causes the collapse of mitochondrial membrane potential and pathological ROS burst due to ETZ. These may cause ROS release in neighbouring mitochondria by being released into the cytosol; that is, ROS-induced ROS release occurs [133-137]. It has been shown that mitochondrial functions are significantly impaired in the created liver sepsis model and the impairment is strongly associated with the extent of mitochondrial ultra-structural abnormalities [138]. Oxidative stress is one of the main pathogenic factors causing mitochondrial dysfunction in acute kidney injury. Since the kidney suffers from oxidative stress during sepsis, one of the most promising approaches to mitigate such damaging results has been proposed as the use of antioxidants. For this purpose, the results of the study in which the mitochondria targeted antioxidant, plastoquinol decylrhodamine 19 (SkQR1) is applied, showed that antioxidant use is beneficial against renal tissue damage. In order not to disturb the redox balance, it has been deemed appropriate to use mitochondria-specific oxidative explosion-extinguishing supplements instead of high-dose traditional antioxidant application [139]. Mitochondria are one of the key organelles involved in the development of pathogenic cascades under septic conditions, acting both as a source and as a target for ROS. For this reason, antioxidant use has been recommended to prevent the development of oxidative stress by stopping or decreasing pathological ROS production in mitochondria. Besides, antioxidants such as multi-antioxidant Ceria-Zirconia nanoparticles developed to remove ROS for sepsis treatment have been studied and recommended for the treatment of inflammatory diseases [140]. It has been stated that selenium, which is used for antioxidant purposes in animal models of bacterial infections such as *Escherichia coli*, *Listeria monocytogenes*, *Dichelobacter nodosus*, *Staphylococcus aureus*, has positive effects and can be used in healing [141-144]. In the experimental study of *Microcystis aeruginosa* against Microcystin LR toxin, melatonin, Vit-E and Vit-C, which are used as a supplement to reduce the cell damage and increased 8-OH-dG, have a very high protective effect. Among these antioxidants, the effect of melatonin was 60 times higher against Vit-C and 70 times

higher against Vit E [11]. Melatonin protects the DNA against oxidative damage by activating antioxidant enzymes and inhibiting prooxidative enzymes. Additionally, it is unique as an antioxidant in its ability to cross biological barriers and multiple action pathways. It is also known to have minimum toxicity even in high doses and thus to be within a wide range of dosage [145-147].

ANTIOXIDANT USE IN VIRAL DISEASES

It has been reported that the application of Vit-E, Vit-C and Zn as antioxidants provides antioxidant protection during the treatment process against oxidative damage caused by both viral infection and antiviral therapy in common viral hepatitis [148]. In studies conducted with selenium, it has been reported that selenium not only enhances Th1 type host immunity against viral infections, but also inhibits the evolution of more virulent strains of viral pathogens in RNA viruses [149-151]. Resveratrol has been reported to be effective as an inhibitor against viral replication and viral-induced inflammation in diseases caused by various pathogenic viruses, including respiratory viruses such as RSV, HCoV and HRV, influenza virus [152]. Anti-inflammatory and antioxidant activities of resveratrol may contribute to alleviating the symptoms of the virus associated with pathological signs [153]. Although it has been reported that the application of N-acetylcysteine (NAC) in infection with influenza virus clinically decreases the incidence of the disease, there are also some researchers who think that it is not healthy to recommend NAC without conducting further studies since it may decrease GSH while increasing the amount of GSSG as a result of showing prooxidant effect when the dose gets higher [154-156]. It has been reported that high doses of ascorbic acid in viral infections clears superoxide anion, inhibits virus proliferation and decreases expression of viral antigens and cellular viral load. It has also been reported to have immunomodulatory characteristics, concentrate in leukocytes, lymphocytes and macrophages, heal chemotaxis, increase neutrophil phagocytic capacity and oxidative killing, support lymphocyte proliferation and function, significantly restore decreased mitochondrial membrane potential and decrease gene expression of pro-inflammatory cytokines [157,158]. It has been reported that ROS production increases for viral replication in BHV-1 infection and causes mitochondrial dysfunction in the cell [159]. SOD activation decreases in cells infected with this virus, quercetin application increases SOD activation and ceruloplasmin level and number of apoptotic cells [105]. In a study conducted on poultry, it was suggested that oxidative stress with duodenal and jejunal mucosa in Newcastle disease virus infection causes pathological damage, while this damage can be eliminated with Vit-E given as supplement and can be used in the treatment of Newcastle disease [160]. It has also been reported that resveratrol application in poultry is useful both as antiviral and as an antioxidant [161]. It has been reported that the application of antioxidant prepartes (zinc, methionine,

Vit-E, selenium) to sheep with foot-and-mouth disease improves the general health conditions and performance of animals since it increases TAC and GPx activity and decreases DNA damage [94]. It has been stated that in viral hepatitis, antioxidants can be used against viruses because they reduce virus replication and oxidant damage due to virus and increase in response to oxidative damage. Antioxidants may be effective in weakening replication and making antiviral interferon therapy more effective [162].

CONCLUSION

There is a great deal of research in animal studies on the mechanism of oxidative stress in the pathogenesis of the disease. Basically, the common point in infectious diseases is the increase in oxidative stress. When the organism encounters an infective pathogen, it increases the amount of ROS in neutrophils to protect the immune defence system. With the high and prolonged continuation of ROS which increases initially to remove the pathogen agent, oxidation and DNA modifications start in lipids and proteins. Disruption of this redox balance triggers cell signal change, causing loss of basic cellular functions, change in tissue structure, apoptosis and eventually tissue damage. It is difficult to use increased oxidative parameters in the differential diagnosis of diseases; however, it will be useful to determine oxidative stress index in determining the pathogenesis and severity of the infection and following the treatment process. On the other hand, antioxidant parameters vary in infective disease studies conducted. While GPx activity and ceruloplasmin increased in some of the studies conducted, there are also studies reporting a decrease. Since SOD activity and TAC are generally suppressed, redox balance cannot be maintained and oxidative stress occurs in the organism. Studies to supplement with antioxidants exogenously are carried out so as not to cause to give more damage with the increasing OSI. In some of these studies, prooxidative effects are seen in the presence of especially metal ions such as iron and copper depending on the molecular structure and concentrations of antioxidants, contrary to what is expected. Studies investigating the preventive effects of rations or supplements rich in antioxidants on the development, progression and treatment in studies on infectious ill patients or in experimental studies are limited or in dose-determination stages. As the pro-oxidative effects of polyphenol compounds such as quercetin, catechin and gallic acid have come to the fore in recently conducted studies, the need for *in vitro* and *in vivo* studies on the dose, solubility, reliability and administration route of antioxidative substances has been increasing. Therefore, epidemiological studies on the effects of antioxidant molecules in healthy and sick animals will play a key role in research.

REFERENCES

1. Halliwell B, Gutteridge JMC: Free Radicals in Biology and Medicine. 4th ed.,

Oxford: Oxford University Press; 2007.

2. Valko M, Leibfritz D, Moncola J, Cronin MTD, Mazur M, Telser J: Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol*, 39, 44-84, 2007. DOI: 10.1016/j.biocel.2006.07.001
3. Nakayama T, Church DF, Pryor WA: Quantitative analysis of the hydrogen peroxide formed in aqueous cigarette tar extracts. *Free Radic Biol Med*, 7 (1): 9-15, 1989. DOI: 10.1016/0891-5849(89)90094-4
4. Janssen YM, Van Houten B, Borm PJ, Mossman BT: Cell and tissue responses to oxidative damage. *Lab Invest*, 69 (3): 261-274, 1993. DOI: 10.1016/j.tox.2007.10.009
5. Soehnlein O, Kenne E, Rotzius P, Eriksson EE, Lindbom LL: Neutrophil secretion products regulate anti-bacterial activity in monocytes and macrophages. *Clin Exp Immunol*, 151(1): 139-145, 2008. DOI: 10.1111/j.1365-2249.2007.03532.x
6. Kalyanaraman B: Teaching the basics of redox biology to medical and graduate students: Oxidants, antioxidants and disease mechanisms. *Redox Biol*, 1, 244-257, 2013. DOI: 10.1016/j.redox.2013.01.014
7. Sarma K, Saravanan M, Mondal DB, De UK, Kumar M: Influence of natural infection of *Toxocara vitulorum* on markers of oxidative stress in Indian buffalo calves. *Indian J Anim Sci*, 82, 1142-1145, 2012. DOI: 10.1016/j.redox.2013.01.014
8. Sen S, Chakraborty R, Sridhar C, Reddy YSR, Debnath B: Free radicals, antioxidants, diseases and phytomedicines: Current status and future prospect. *Int J Pharm Sci Res*, 3 (1): 91-100, 2010.
9. Augustin W, Wiswedel I, Noack H, Reinheckel T, Reichelt O: Role of endogenous and exogenous antioxidants in the defence against functional damage and lipid peroxidation in rat liver mitochondria. *Mol Cell Biochem*, 174 (1-2): 199-205, 1997.
10. Karabulut H, Gülay MŞ: Free Radicals. *MAKÜ Sağlık Bil. Enst. Derg.*, 4 (1): 50-59, 2016.
11. Al-Jassabi S, Khalil AM: Microcystin-induced 8-hydroxydeoxyguanosine in DNA and its reduction by melatonin, vitamin C, and vitamin E in mice. *Biochemistry*, 71, 1115-1119, 2006. DOI: 10.1134/S0006297906100099
12. Cenesiz M, Sagkan Ozturk A, Dalgın D, Yarım GF, Ciftci G, Ozdemir R, Guzel M, Kazak F, Cenesiz S: Investigation of acute phase reactants and antioxidant capacity in calves infected with *Cryptosporidium parvum*. *Kafkas Univ Vet Fak Derg*, 23 (3): 481-485, 2017. DOI: 10.9775/kvfd.2016.17183
13. Merhan O, Taşçı GT, Bozukluhan K, Aydın N: Determination of oxidative stress index and total sialic acid in cattle infested with *Hypoderma* spp. *Kafkas Univ Vet Fak Derg*, 26 (5): 633-636, 2020. DOI: 10.9775/kvfd.2020.24071
14. Celi P: Biomarkers of oxidative stress in ruminant medicine. *Immunopharmacol Immunotoxicol*, 33 (2): 233-240, 2011, DOI: 10.3109/08923973.2010.514917
15. Murad F: Nitric oxide and cyclic GMP in cell signaling and drug development. *N Engl J Med*, 355, 2003-2011, 2006. DOI: 10.1056/NEJMs063904
16. Miller JK, Brzezinska-Slebodzinska E, Madsen FC: Oxidative stress, antioxidants, and animal function. *J Dairy Sci*, 76, 2812-2823, 1993. DOI: 10.3168/jds.S0022-0302(93)77620-1
17. Agarwal A, Gupta S, Sharma RK: Role of oxidative stress in female reproduction. *Reprod Biol Endocrinol*, 3:28 (2005), 2005. DOI: 10.1186/1477-7827-3-28
18. Dröge W: Free radicals in the physiological control of cell function. *Physiol Rev*, 82, 47-95, 2002. DOI: 10.1152/physrev.00018.2001
19. Misro MM, Choudhury L, Upreti K, Gautam D, Chaki SP, Mahajan AS, Babbar R: Use of hydrogen peroxide to assess the sperm susceptibility to oxidative stress in subjects presenting a normal semen profile. *Int J Androl*, 27, 82-87, 2004. DOI: 10.1046/j.0105-6263.2003.00451.x
20. Guérin G, El Mouatassim S, Ménéz Y: Oxidative stress and protection against reactive oxygen species in the pro-implantation embryo and its surroundings. *Hum Reprod Update*, 7, 175-189, 2001. DOI: 10.1093/humupd/7.2.175
21. Winrow VR, Winyard PG, Morris CJ, Blake DR: Free radicals in inflammation; second messengers and mediators of tissue destruction. *Br Med Bul*, 49, 506-522, 1993. DOI: 10.1093/oxfordjournals.bmb.a072627
22. Devasagayam TPA, Tilak JC, Boloor KK, Sane KS, Ghaskadbi SS, Lele RD: Free radicals and antioxidants in human health: Current status and future prospects. *JAPI*, 52, 794-804, 2004.
23. Dröge W: Free radicals in the physiological control of cell function. *Physiol Rev*, 82, 47-95, 2002. DOI: 10.1152/physrev.00018.2001
24. Fang YZ, Yang S, Wu G: Free radicals, antioxidants, and nutrition. *Nutrition* 18, 872-879, 2002. DOI: 10.1016/S0899-9007(02)00916-4
25. Lander HM: An essential role for free radicals and derived species in signal transduction. *FASEB*, 11 (2): 118-124, 1997. DOI: 10.1096/fasebj.11.2.9039953
26. Kuru M, Ögün M, Kulaksız R, Kükürt A, Oral H: Comparison of oxidative/nitrosative stress, leptin and progesterone concentrations in pregnant and non-pregnant Abaza goats synchronized with controlled internal drug release application. *Kafkas Univ Vet Fak Derg*, 24 (6): 887-892, 2018. DOI: 10.9775/kvfd.2018.20222
27. Schreck R, Baeuerle PA: A role for oxygen radicals as second messengers. *Trends Cell Biol*, 1 (2-3): 39-42, 1991. DOI: 10.1016/0962-8924(91)90072-H

- 28. Fujii J, Iuchi Y, Okada F:** Fundamental roles of reactive oxygen species and protective mechanisms in the female reproductive system. *Reprod Biol Endocrinol*, 3:43 (2005), 2005. DOI: 10.1186/1477-7827-3-43
- 29. Martin KR, Barrett JC:** Reactive oxygen species as double-edged swords in cellular processes: Low-dose cell signaling versus high-dose toxicity. *Hum Exp Toxicol*, 21 (2): 71-75, 2002. DOI: 10.1191/0960327102ht2130a
- 30. Ni SL, Gao F, Zuo CX, Tang XD, Liu MJ, Chang JJ, Wang Y, Chen DK, Ma WT:** Neutrophils: A critical participant in common diseases of ruminants. *Kafkas Univ Vet Fak Derg*, 26 (5): 719-727, 2020. DOI: 10.9775/kvfd.2020.24012
- 31. Kinnula VL, Crapo JD:** Superoxide dismutases in malignant cells and human tumors. *Free Radic Biol Med*, 36 (6): 718-744, 2004. DOI: 10.1016/j.freeradbiomed.2003.12.010
- 32. Singh U, Jialal I:** Oxidative stress and atherosclerosis. *Pathophysiology*, 13, 129-142, 2006. DOI: 10.1016/j.pathophys.2006.05.002
- 33. Smith MA, Rottkamp CA, Arun AN, Raina AK, Perry G:** Oxidative stress in Alzheimer's disease. *Biochim Biophys Acta Mol Basis Dis*, 1502 (1): 139-144, 2000. DOI: 10.1016/S0925-4439(00)00040-5
- 34. Kalantari H, Forouzandeh H, Khodayar MJ, Siahpoosh A, Saki N, Kheradmand P:** Antioxidant and hepatoprotective effects of *Capparis spinosa* L. fractions and quercetin on tertbutyl hydroperoxide-induced acute liver damage in mice. *J Tradit Complement Med*, 8 (1): 120-127, 2017. DOI: 10.1016/j.jtcme.2017.04.010
- 35. Sas K, Robotka H, Toldi J, Vecsei L:** Mitochondria, metabolic disturbances, oxidative stress and the kynureninesystem, with focus on neurodegenerative disorders. *J Neurol Sci*, 257, 221-239, 2007. DOI: 10.1016/j.jns.2007.01.033
- 36. Bolton JD, Trush MA, Penning TM, Dryhurst G, Monks TJ:** Role of quinones in toxicology. *Chem Res Toxicol*, 13 (3): 135-160, 2000. DOI: 10.1021/tx9902082
- 37. Hayden MS, Ghosh S:** Shared principles in NF-kappaB signaling. *Cell*, 132, 344-362, 2008. DOI: 10.1016/j.cell.2008.01.020
- 38. Guidi I, Galimberti D, Lonati S, Novembrino C, Bamonti F, Tiriticco M, Fenoglio C, Venturelli E, Baron P, Bresolin N, Scarpin E:** Oxidative imbalance in patients with mild cognitive impairment and Alzheimer's disease. *Neurobiol Aging*, 27, 262-269, 2006. DOI: 10.1016/j.neurobiolaging.2005.01.001
- 39. Ateel GE:** Oxidants and antioxidants in alcohol-induced liver disease. *Gastroenterology*, 124 (3): 778-790, 2003. DOI: 10.1053/gast.2003.50087
- 40. Sundari PN, Wilfred G, Ramakrishna B:** Does oxidative protein damage play a role in the pathogenesis of carbon tetrachloride-induced liver injury in the rat? *Biochim Biophys Acta Mol Basis Dis*, 1362, 169-176, 1997. DOI: 10.1016/S0925-4439(97)00065-3
- 41. Hyun DH, Emerson SS, Jo DG, Mattson MP, Cabo R:** Calorie restriction up-regulates the plasma membrane redox system in brain cells and suppresses oxidative stress during aging. *Proc Natl Acad Sci USA*, 103 (52): 19908-19912, 2006. DOI: 10.1073/pnas.0608008103
- 42. Upston JM, Kritharides L, Stocker R:** The role of vitamin E in atherosclerosis. *Prog Lipid Res*, 42 (5): 405-422, 2003. DOI: 10.1016/S0163-7827(03)00024-9
- 43. Kirmizigül AH, Özcelik M, Ogun M, Erkilic EE, Paksoy N, Merhan O, Uzlu E:** Serum Cu, Mn and Zn levels and oxidative stress in cattle performing tongue-playing. *Kafkas Univ Vet Fak Derg*, 25 (6): 787-791, 2019. DOI: 10.9775/kvfd.2019.21861
- 44. Moslen MT:** Reactive oxygen species in normal physiology, cell injury and phagocytosis. In: Armstrong D (Ed): *Advances in Experimental Medicine and Biology, Free Radicals in Diagnostic Medicine*. 17-27, New York: Plenum Press, 366, 1994.
- 45. Doherty PC, Turner SJ, Webby RG, Thomas PG:** Influenza and the challenge for immunology. *Nat Immunol*, 7, 449-455, 2006. DOI: 10.1038/ni1343
- 46. Perrone LA, Plowden JK, García-Sastre A, Katz JM, Tumpey TM:** H5N1 and 1918 pandemic influenza virus infection results in early and excessive infiltration of macrophages and neutrophils in the lungs of mice. *PLoS Pathog*, 4 (8): e1000115, 2008. DOI: 10.1371/journal.ppat.1000115
- 47. Weiss SJ, Klein R, Sliwka A, Wei M:** Chlorination of taurine by human neutrophils: Evidence for hypochlorous acid generation. *J Clin Invest*, 70 (3): 598-607, 1982. DOI: 10.1172/JCI110652
- 48. Maeda H, Akaike T:** Oxygen free radicals as pathogenic molecules in viral diseases. *Proc Soc Exp Biol Med*, 198, 721-727, 1991. DOI: 10.3181/00379727-198-43309C
- 49. Clark RA, Stone PJ, El Hag A, Calore JD, Franzblau C:** Myeloperoxidase-catalyzed inactivation of alpha 1-protease inhibitor by human neutrophils. *J Biol Chem*, 256, 3348-3353, 1981.
- 50. Vlahos R, Selemidis S:** NADPH oxidases as novel pharmacologic targets against influenza A virus infection. *Mole Pharmacol*, 86, 747-759, 2014. DOI: 10.1124/mol.114.095216
- 51. Yan LJ, Traber MG, Kobuchi H, Matsugo S, Tritschler HJ, Packer L:** Efficacy of hypochlorous acid scavengers in the prevention of protein carbonyl formation. *Arch Biochem Biophys*, 327 (2): 330-34, 1996. DOI: 10.1006/abbi.1996.0130
- 52. McNeil KS, Knox DP, Proudfoot L:** Anti-inflammatory responses and oxidative stress in *Nippostrongylus brasiliensis* induced pulmonary inflammation. *Parasite Immunol*, 24, 15-22, 2002. DOI: 10.1046/j.0141-9838.2001.00428.x
- 53. Dzik JM, Gołos B, Jagielska E, Kapala A, Wałajczyk-Rode AI:** Early response of guinea-pig lungs to *Trichinella spiralis* infection. *Parasite Immunol*, 24, 369-379, 2002. DOI: 10.1046/j.1365-3024.2002.00474.x
- 54. Değer S, Değer Y, Ertekin A, Gül A, Biçek K, Özdal N:** *Dictyocaulus viviparus* ile enfekte sığırlarda lipit peroksidasyon ve antioksidan durumunun saptanması. *Türkiye Parazitoloj Derg*, 32 (3): 234-237, 2008.
- 55. Nisbet C, Çenesiz S, Açııcı Z, Umur Ş:** Determination of the serum malondialdehyde, ceruloplasmin, adenosine deaminase levels in cattle with cystic echinococcosis. *J Erciyes Univ Vet Fak Derg*, 5 (1): 1-4, 2008.
- 56. Bargagli E, Olivieri C, Bennett D, Prasse A:** Oxidative stress in the pathogenesis of diffuse lung diseases: A review. *Respir Med*, 103 (9): 1245-1256, 2009. DOI: 10.1016/j.rmed.2009.04.014
- 57. Heidarpour M, Mohri M, Borji H, Moghdass E:** Oxidant/antioxidant status in cattle with liver cystic echinococcosis. *Vet Parasitol*, 195 (1-2): 131-135, 2013. DOI: 10.1016/j.vetpar.2013.01.018
- 58. Heidarpour M, Mohri M, Borji H, Moghdass E:** Oxidative stress and trace elements in camel (*Camelus dromedarius*) with liver cystic echinococcosis. *Vet Parasitol*, 187 (3-4): 459-463, 2012. DOI: 10.1016/j.vetpar.2012.01.015
- 59. Heidarpour M, Mohri M, Borji H, Moghdass E:** Oxidant/antioxidant balance and trace elements status in sheep with liver cystic echinococcosis. *Comp Clin Pathol*, 22, 1043-1049, 2013. DOI: 10.1007/s00580-012-1523-5
- 60. Irak K, Celik AB, Karakoc Z, Çelik ÖY, Mert H, Mert N, Kaya OM:** Oxidant/antioxidant status, PON1 and ARES activities, trace element levels, and histological alterations in sheep with cystic echinococcosis. *Iran J Parasitol*, 13 (3): 448-456, 2018.
- 61. Sagkan-Ozturk A, Durgut R, Ozturk OH:** Oxidant/antioxidant status in lambs and sheep with liver and lung cystic echinococcosis diagnosed by ultrasonography and necropsy. *Vet Parasitol*, 208 (3-4): 280-285, 2015. DOI: 10.1016/j.vetpar.2014.12.034
- 62. Hanedan B, Kirbas A, Kandemir FM, Ozkaraca M, Kilic K, Benzer F:** Arginase activity and total oxidant/antioxidant capacity in cows with lung cystic echinococcosis. *Med Weter*, 71 (3): 167-170, 2015.
- 63. Sarma AD, Mallick AR, Ghosh AK:** Free radicals and their role in different clinical conditions: An overview. *Int J Pharm Sci Res*, 1 (3): 185-192, 2010.
- 64. Bozukluhan K, Merhan A, Özcan A, Gokce H, Gokce G:** Investigation of the levels of serum haptoglobin, oxidative indicators and some biochemical parameters in calves naturally infected with *Toxocara vitulorum*. *Ankara Univ Vet Fak Derg*, 64, 75-79, 2017.
- 65. Özkurt G, Gökçen A, Çamkerten İ, Şahin T, Balkan BM, Boz M:** Erythrocyte SOD, CAT, GPx enzymes activity and MDA level in Kilis goats with naturally-occurred nematodiasis. *Harran Univ Vet Fak Derg*, 1 (2): 107-110, 2012.
- 66. Gültekin M, Paşa S, Ural K, Balıkcı C, Sevrî G, Açııcı E, Gültekin G:** Oxidative status and lipid profile among dogs at different stages of visceral leishmaniasis. *Türkiye Parazitoloj Derg*, 41, 183-187, 2017. DOI: 10.5152/tpd.2017.5398
- 67. Merhan O, Bozukluhan K, Gokce H:** Acute phase proteins and biochemical and oxidative stress parameters in *Hypoderma* spp. infested cattle. *J Hellenic Vet Med Soc*, 68 (4): 535-540, 2017. DOI: 10.12681/jhvm.16049
- 68. Bozukluhan K, Merhan O, Kızıltepe S, Harmankaya A, Gökçe G:** Determination of oxidative stress and ceruloplasmin levels in sheep with toxoplasmosis. *Van Vet J*, 31 (2): 83-86, 2020. DOI: 10.36483/vanvetj.646976
- 69. Nasreldin N, Ewida RM, Hamdon H, Elnaker YF:** Molecular diagnosis and biochemical studies of tick-borne diseases (anaplasmosis and babesiosis) in Aberdeen Angus cattle in New Valley. *Egypt Vet World*, 13 (9): 1884-1891, 2020. DOI: 10.14202/vetworld.2020.1884-1891
- 70. Esmailnejad B, Tavassoli M, Dalir-Naghadeh B, SepidehRajabi A, Mohammadi V, Anassori E, Ehteshamfar S:** Status of oxidative stress, trace elements, sialic acid and cholinesterase activity in cattle naturally infected with *Babesia bigemina*. *Comp Immunol Microbiol Infect Dis*, 71:10150, 2020. DOI: 10.1016/j.cimid.2020.101503
- 71. Pajić M, Aleksić N, Vejnović B, Polaček V, Novakov N, Andrić DO, Stanimirović Z:** Influence of anticoccidials on oxidative stress, production performance and faecal oocyst counts in broiler chickens infected with *Eimeria* species. *Kafkas Univ Vet Fak Derg*, 25 (3): 379-385, 2019. DOI: 10.9775/kvfd.2018.21021
- 72. Kızıl M, Baydar E, Kızıl Ö:** Tayleriyozisli sığırlarda antioksidan parametrelerdeki değişiklikler. *FÜ Sağlık Bil Vet Derg*, 25 (2): 53-56, 2011.
- 73. Amy D, Klion MD, Thomas B, Nutman MD:** The role of eosinophils in host defense against helminth parasites. *J Allergy Clin Immunol*, 113 (1): 30-37, 2004. DOI: 10.1016/j.jaci.2003.10.050
- 74. Özçelik M, İssi M, Gül Y, Güler O, Şimşek H, Özdemir N, Kılıç A:** Bakteriyeel pnömonili besi sığırlarında oluşan serbest radikal hasarının antioksidan aktivite ve bazı mineral maddeler üzerine etkisi. *Erciyes Univ Vet Fak Derg*, 11 (2): 111-116, 2014.
- 75. Joshi V, Gupta VK, Bhanuprakash AG, Mandal RSK, Dimri U, Ajith Y:** Haptoglobin and serum amyloid A as putative biomarker candidates of naturally occurring bovine respiratory disease in dairy calves. *Microb Pathog*, 116, 33-37, 2018.

DOI: 10.1016/j.micpath.2018.01.001

- 76. Alaka OO, Emikpe BO, Jarikre TA, Ola OO:** Pathology and bronchoalveolar lavage fluid cellular and oxidative stress changes in canine pneumonia in Nigeria. *Comp Clin Pathol*, 28, 1681-1687, 2019. DOI: 10.1007/s00580-019-03000-2
- 77. Ismael M, El-Sayed MS, Metwally AM, Ibrahim, Zeinab KI, El-Saman, RM:** Clinical and haematobiochemical evaluation of pneumonia in calves with special reference to oxidant/antioxidant indices. *AJVS*, 54 (2): 40-44, 2017. DOI: 10.5455/ajvs.246119
- 78. Pancarci B, Unver A, Cetinkol Y, Atakisi O:** Nitric oxide levels and oxidant-antioxidant capacities in patients with brucellosis. *Acta Med Mediterr*, 32, 69-73, 2016. DOI: 10.19193/0393-6384_2016_1_10
- 79. Rhodes G, Richardson H, Hermon-Taylor J, Weightman A, Higham A, Pickup R:** *Mycobacterium avium* Subspecies *paratuberculosis*: Human exposure through environmental and domestic aerosols. *Pathogens*, 3 (3): 577-595, 2014. DOI: 10.3390/pathogens3030577
- 80. Wendland BE, Aghdassi E, Tam C, Carrier J, Steinhart AH, Wolman SL, Baron D, Allard JP:** Lipid peroxidation and plasma antioxidant micronutrients in Crohn disease. *Am J Clin Nutr*, 74, 259-264, 2001. DOI: 10.1093/ajcn/74.2.259
- 81. Banerjee RK:** Indomethacin inactivates gastric peroxidase to induce reactive oxygen mediated gastric mucosal injury and curcumin protects it by preventing peroxidase inactivation and scavenging reactive oxygen. *Free Radic Biol Med*, 40, 1397-1408, 2006. DOI: 10.1016/j.freeradbiomed.2005.12.016
- 82. Oudkerk Pool M, Bouma G, Visser JJ, Kolkman JJ, Tran DD, Meuwissen SG, Peña AS:** Serum nitrate levels in ulcerative colitis and Crohn's disease. *Scand J Gastroenterol*, 30, 784-788, 1995. DOI: 10.3109/00365529509096328
- 83. Cenesiz M, Ciftci G, Dalgin D, Kilic Y, Yarim GF, Cenesiz S:** Evaluation of oxidant and antioxidant capacity in paratuberculosis positive cattle. *Pakistan J Zool*, 48 (5): 1603-1606, 2016.
- 84. Qasem A, AbdelAty A, AbuSuwa H, Naser SA:** Oxidative stress due to *Mycobacterium avium* subspecies *paratuberculosis* (MAP) infection upregulates selenium-dependent GPx activity. *Gut Pathog*, 8, 12, 2016. DOI: 10.1186/s13099-016-0090-8
- 85. Meral Ö, Ercan N, Fidancı UR:** Septisemili buzağalarda lipid peroksidasyon düzeyi ve antioksidan enzim aktiviteleri. *Ankara Üniv Vet Fak Derg*, 64, 161-164, 2017.
- 86. Kumar A, Pant P, Basu S, Rao GRK, Khanna HD:** Oxidative stress in neonatal hyperbilirubinemia. *J Trop Pediatr*, 53, 69-71, 2007. DOI: 10.1093/tropej/fml060
- 87. Aydoğdu U, Coşkun A, Başbuğ O, Ağaoglu ZT:** Parvoviral enteritisli köpeklerde total oksidan-antioksidan durum ile oksidatif stres indeksinin değerlendirilmesi. *FÜ Sağlık Bil Vet Derg*, 32 (3): 161-164, 2018.
- 88. Elsayed NM, Kubesy AA, Salem NY:** Altered blood oxidative stress biomarkers in association with canine parvovirus enteritis. *Comp Clin Pathol*, 29, 355-359, 2020. DOI: 10.1007/s00580-019-03067-x
- 89. Bozukluhan K, Merhan O, Gokce HI, Ogun M, Atakisi E, Kızıltepe S, Gokce G:** Determination of some acute phase proteins, biochemical parameters and oxidative stress in sheep with naturally infected sheepox virus. *Kafkas Univ Vet Fak Derg*, 24 (3): 437-441, 2018. DOI: 10.9775/kvfd.2017.19167
- 90. Uzlu E, Karapehlivan M, Erdoğan HM, Kızıltepe Ş, Erkilic EE, Devci HA, Gökçe E, Kaya İ, Çitil M:** Serum and saliva sialic acid and oxidative stress parameters changes in bulls with Foot and Mouth Disease. *Kafkas Univ Vet Fak Derg*, 22 (3): 321-325, 2016. DOI: 10.9775/kvfd.2015.13114
- 91. Mousa SA, Galal MKH:** Alteration in clinical, hemobiochemical and oxidative stress parameters in Egyptian cattle infected with Foot and Mouth Disease. *J Anim Sci Adv*, 3, 485-491, 2013.
- 92. Devci HA, Kükürt A, Nur G, Alpay M, Merhan O, Bozukluhan K, Yılmaz V, Karapehlivan M:** Serum paraoxonase activity and total sialic acid in sheep with foot and mouth disease. *Med Weter*, 74 (3): 199-202, 2018. DOI: 10.21521/mw.6078
- 93. Yarim GF, Nisbet C, Cenesiz S, Coşkun A:** Şap hastalıklı koyunlarda serum nitrik oksit düzeyi ve adenoazin deaminaz aktivitesinin araştırılması. *Ankara Üniv Vet Fak Derg*, 53, 161-164, 2006.
- 94. Abou-Zeina HAA, Nasr SM, Nassar SA, Farag TK, El-Bayoumy MK, Ata EB, Hassan NMF, Abdel-Aziem SH:** Beneficial effects of antioxidants in improving health conditions of sheep infected with foot-and-mouth disease. *Trop Anim Health Prod*, 51 (8): 2379-2386, 2019. DOI: 10.1007/s11250-019-01952-9
- 95. Erkilic EE, Ögün M, Kırmızıgül AH, Adalı Y, Ermutlu CŞ, Eroğlu HA, Kükürt A, Çitil M, Uzlu E:** Determination of some oxidative stress and inflammation markers in serum, blood and CSF in cattle with head-eye form of malignant catarrhal fever. *Kafkas Univ Vet Fak Derg*, 23 (4): 515-519, 2017. DOI: 10.9775/kvfd.2016.17166
- 96. Koçak G:** The effect of quercetin on oxidative stress in madin-darby bovine kidney cell culture infected with bovine herpes 1 virus. *Ondokuz Mayıs Üniv. Sağlık Bil. Enst.*, 2019.
- 97. Jarikre TA, Ohore GO, Oyagbemi AA, Emikpe BO:** Evaluation of oxidative stress in caprine bronchoalveolar lavage fluid of pneumonic and normal lungs. *Inter J Vet Sci Med*, 5, 143-147, 2017. DOI: 10.1016/j.ijvsm.2017.09.001
- 98. De Mochel NSR, Seronello S, Wang SH, Ito C, Zheng JX, Liang TJ, Lambeth JD, Choi J:** Hepatocyte NAD(P)H oxidases as an endogenous source of reactive oxygen species during hepatitis C virus infection. *Hepatology*, 52, 47-59, 2010. DOI: 10.1002/hep.23671
- 99. Reshi ML, Su YC, Hong JR:** RNA viruses: ROS-mediated cell death. *Int J Cell Biol*, 2014:467452, 2014. DOI: 10.1155/2014/467452
- 100. Sies H:** Biochemistry of oxidative stress. *Angew Chem Int Ed Engl*, 25, 1058-1071, 1986. DOI: 10.1002/anie.198610581
- 101. Molteni CG, Principi N, Esposito S:** Reactive oxygen and nitrogen species during viral infections. *Free Radic Res*, 48, 1163-1169, 2014. DOI: 10.3109/10715762.2014.945443
- 102. Urtasun R, Lopategi A, George J, Leung TM, Lu Y, Wang X, Ge X, Fiel MI, Nieto N:** Osteopontin, an oxidant stress sensitive cytokine, up-regulates collagen-I via integrin alpha(V)beta(3) engagement and PI3K/pAkt/NFkappaB signaling. *Hepatology*, 55, 594-608, 2012. DOI: 10.1002/hep.24701
- 103. Ivanov AV, Valuev-Elliston VT, Tyurina DA, Ivanova ON, Kochetkov SN, Bartosch B, Isaguliants MG:** Oxidative stress, a trigger of hepatitis C and B virus-induced liver carcinogenesis. *Oncotarget*, 8 (3): 3895-3932, 2017. DOI: 10.18632/oncotarget.13904
- 104. Fujita N, Sugimoto R, Ma N, Tanaka H, Iwasa M, Kobayashi Y, Kawanishi S, Watanabe S, Kaito M, Takei Y:** Comparison of hepatic oxidative DNA damage in patients with chronic hepatitis B and C. *J Viral Hepatitis*, 15, 498-507, 2008. DOI: 10.1111/j.1365-2893.2008.00972.x
- 105. Fu X, Jiang X, Chen X, Zhu L, Zhang G:** The differential expression of mitochondrial function-associated proteins and antioxidant enzymes during bovine herpesvirus 1 infection: A potential mechanism for virus infection-induced oxidative mitochondrial dysfunction. *Mediat Inflamm*, 2019:7072917, 2019. DOI: 10.1155/2019/7072917
- 106. Sartori G, Jardim NS, Sari MHM, Flores EF, Prigol M, Nogueir CW:** Diphenyl diselenide reduces oxidative stress and toxicity caused by HSV-2 infection in mice. *J Cell Biochem*, 118 (5): 1028-1037, 2017. DOI: 10.1002/jcb.25667
- 107. Devci HA, Kükürt A, Uzlu E, Sözdutmaz İ, Merhan O, Aktaş S, Alpay M, Kaya İ, Karapehlivan M:** Evaluation of paraoxonase activity, total sialic acid and oxidative stress in sheep with Ecthyma Contagiosa. *Kafkas Univ Vet Fak Derg*, 23 (3): 453-457, 2017. DOI: 10.9775/kvfd.2016.17001
- 108. Aguilar A, Alvarez-Vijande R, Capdevila S, Alcobero J, Alcaraz A:** Antioxidant patterns (superoxide dismutase, glutathione reductase, and glutathione peroxidase) in kidneys from non-heart-beating-donors. *Transplant Proc*, 39, 249-252, 2007. DOI: 10.1016/j.transproceed.2006.10.212
- 109. Yeum KJ, Russell RM, Krinsky NI, Adlani G:** Biomarkers of antioxidant capacity in hydrophilic and lipophilic compartments of human plasma. *Arch Biochem Biophys*, 430, 97-103, 2004. DOI: 10.1016/j.abb.2004.03.006
- 110. Henle ES, Linn S:** Formation, prevention, and repair of DNA damage by iron/hydrogen peroxide. *J Biol Chem*, 272 (31): 19095-19098, 1997. DOI: 10.1074/jbc.272.31.19095
- 111. Chiurchiù V, Maccarrone M:** Chronic inflammatory disorders and their redox control: From molecular mechanisms to therapeutic opportunities. *Antioxid Redox Signal*, 15 (9): 2605-2641, 2011. DOI: 10.1089/ars.2010.3547
- 112. Paap B, Wilson DM, Sutherland BM:** Human abasic endonuclease action on multilesion abasic clusters: Implications for radiation-induced biological damage. *Nucleic Acids Res*, 36 (8): 2717-2727, 2008. DOI: 10.1093/nar/gkn118
- 113. Janssen-Heininger YMW, Mossman BT, Heintz NH, Forman HJ, Kalyanaraman B, Finkel T, Stamler JS, Rhee SG, van der Vliet A:** Redox-based regulation of signal transduction: Principles, pitfalls, and promises. *Free Radic Biol Med*, 45 (1): 1-17, 2008. DOI: 10.1016/j.freeradbiomed.2008.03.011
- 114. Bouayed J, Bohn T:** Exogenous antioxidants-Double-edged swords in cellular redox state health beneficial effects at physiologic doses versus deleterious effects at high doses. *Oxid Med Cell Longev*, 3 (4): 228-237, 2010. DOI: 10.4161/oxim.3.4.12858
- 115. Marrocco I, Altieri F, Pelusa I:** Measurement clinical significance of biomarkers of oxidative stress in human. *Oxid Med Cell Longev*, 2017, 6501046, 2017. DOI: 10.1155/2017/s6501046
- 116. Bendich A:** Antioxidant vitamins and human immune responses. *Vitam Hom*, 52, 35-62, 1996. DOI: 10.1016/s0083-6729(08)60406-9
- 117. Rotelli AET, Guardia T, Juarez AO, De La Rocha NE, Pelzer LE:** Comparative study of flavonoids in experimental models of inflammation. *Pharmacol Res*, 48 (6): 601-606, 2003. DOI: 10.1016/s1043-6618(03)00225-1
- 118. Heim KE, Tagliaferro AR, Bobilya DJ:** Flavonoid antioxidants: Chemistry, metabolism and structure-activity relationships. *J Nutr Biochem*, 13, 572-584, 2002. DOI: 10.1016/S0955-2863(02)00208-5
- 119. Cao G, Sofic E, Prior RL:** Antioxidant and prooxidant behavior of flavonoids: Structure-activity relationships. *Free Radic Biol Med*, 22, 749-60, 1997. DOI: 10.1016/S0891-5849(96)00351-6
- 120. Nijveldt RJ, van Nood E, Van Hoorn DEC, Boelens PG, van Norren K, van Leeuwen, PA:** Flavonoids: A review of probable mechanisms of action and potential

applications. *Am J Clin Nutr*, 74 (4): 418-425, 2001. DOI: 10.1093/ajcn/74.4.418

121. Campbell MA, Sefton CM: Protein tyrosine phosphorylation is induced in murine B lymphocytes in response to stimulation with anti-immunoglobulin. *EMBO J*, 9 (7): 2125-2131, 1999. DOI: 10.1002/j.1460-2075.1990.tb07381.x

122. Tordera M, Ferrandiz ML, Alcaraz MJ: Influence of anti-inflammatory flavonoids on degranulation and arachidonic acid release in rat neutrophils. *Zeitschrift für Naturforschung Section C*, 49 (3-4): 235-240, 1994. DOI: 10.1515/znc-1994-3-412

123. Kocyigit A, Koyuncu I, Taskin A, Dikilitas M, Bahadori F, Turkkan B: Antigenotoxic and antioxidant potentials of newly derivatized compo-und naringenin-oxime relative to naringenin on human mononuclear cells. *Drug Chem Toxicol*, 39 (1): 66-73, 2016. DOI: 10.3109/01480545.2015.1026973

124. Merchant LL, Murphy SP, Hankin JH, Wilkens LR, Kolonel LN: Intake of flavonoids and lung cancer. *J Nat Cancer Inst*, 92 (2): 154-160, 2000. DOI: 10.1093/jnci/92.2.154

125. Azam S, Hadi N, Khan NU, Hadi SM: Prooxidant property of green tea polyphenols epicatechin and epigallocatechin-3-gallate: Implications for anticancer properties. *Toxicol In Vitro*, 18, 555-561, 2004. DOI: 10.1016/j.tiv.2003.12.012

126. Robaszekiewicz A, Balcerczyk A, Bartosz G: Antioxidative and prooxidative effects of quercetin on A549 cells. *Cell Biol Int*, 31, 1245-50, 2007. DOI: 10.1016/j.cellbi.2007.04.009

127. Jamshidi-Kia F, Wibowo JP, Elachouri M, Masumi R, Salehifard-Jouneghani, Abolhassanzadeh Z, Lorigooini Z: Battle between plants as antioxidants with free radicals in human body. *J Herbmed Pharmacol*, 9 (3): 191-199, 2020. DOI: 10.34172/jhp.2020.25

128. MÜderrisoglu S, Cenesiz S: Sodyum Florür Toksikasyonunda Kuersetinin Oksidan- antioksidan Etkileri. LAP LAMBERT Academic Publishing, 2017.

129. Sharma M, Sehgal R, Kaur S: Evaluation of nephroprotective and immunomodulatory activities of antioxidants in combination with cisplatin against murine visceral leishmaniasis. *PLoS Negl Trop Dis*, 6 (5):e1629, 2012. DOI: 10.1371/journal.pntd.0001629

130. Ram PK, Singh SK, Srivastava A, Kumar G, Jaiswal AK, Yadav B, Garg SK: Effects of injectable trace minerals (ITMs) on Th1/Th2 cytokine balance of newborn calves with tropical theileriosis. *Biol Trace Elem Res*, 2020. DOI: 10.1007/s12011-020-02263-z

131. Rizk MA, El-Sayed SA, AbouLaila M, Yokoyama N, Igarashi I: Evaluation of the inhibitory effect of N-acetyl-L-cysteine on *Babesia* and *Theileria* parasites. *Exp Parasitol*, 179, 43-48, 2017. DOI: 10.1016/j.exppara.2017.06.003

132. Shinde RM, Bhikane AU, Naraladkar BW, Masare PS: Antioxidants as an adjunct therapy in clinical management of babesiosis in cattle: A novel approach. *Ruminant Sci*, 8 (1): 93-100, 2019.

133. Singer M: The role of mitochondrial dysfunction in sepsis-induced multi-organ failure. *Virulence*, 5 (1): 66-72, 2014. DOI: 10.4161/viru.26907

134. Zorov DB, Juhaszova M, Sollott SJ: Mitochondrial ROS-induced ROS release: An update and TAS review. *Biochim Biophys Acta*, 1757 (5-6): 509-517, 2006. DOI: 10.1016/j.bbabi.2006.04.029

135. Brady NR, Hamacher-Brady A, Westerhoff HV, Gottlieb RA: A wave of reactive oxygen species (ROS)-induced ROS release in a sea of excitable mitochondria. *Antioxid Redox Signal*, 8 (9-10): 1651-1665, 2006. DOI: 10.1089/ars.2006.8.1651

136. Plotnikov EY, Kazachenko AV, Vysokikh MY, Vasileva AK, Tcvirkun DV, Isaev NK, Kirpatovskiy VI, Zorov DB: The role of mitochondria in oxidative and nitrosative stress during ischemia/reperfusion in the rat kidney. *Kidney Int*, 72 (12): 1493-1502, 2007. DOI: 10.1038/sj.ki.5002568

137. Silachev DN, Plotnikov EY, Zorova LD, Pevzner IB, Sumbatyan NV, Korshunova GA, Gulyaev MV, Pirogov YA, Skulachev VP, Zorov DB: Neuroprotective effects of mitochondria-targeted plastoquinone and thymoquinone in a rat model of brain ischemia/reperfusion injury. *Molecules*, 20, 14487-14503, 2015. DOI: 10.3390/molecules200814487

138. Crouser ED, Julian MW, Blaho DV, Pfeiffer DR: Endotoxin-induced mitochondrial damage correlates with impaired respiratory activity. *Crit Care Med*, 30 (2): 276-284, 2002. DOI: 10.1097/00003246-200202000-00002

139. Plotnikov EY, Pevzner IB, Zorova LD, Chernikov VP, Prusov AN, Kireev II, Silachev DN, PV Skulachev, Zorov DB: Mitochondrial damage and mitochondria-targeted antioxidant protection in LPS-induced acute kidney injury. *Antioxidants*, 8, 176, 2019. DOI: 10.3390/antiox8060176

140. Soh M, Kang DW, Jeong HG, Kim D, Kim DY, Yang W, Song C, Baik S, Choi IY, Ki SK, Kwon HJ, Kim T, Kim CK, Lee SH, Hyeon T: Ceria-zirconia nanoparticles as an enhanced multi-antioxidant for sepsis treatment. *Angew Chem Int Ed Engl*, 56 (38): 11399-11403, 2017. DOI: 10.1002/anie.201704904

141. Kim HW, Ha US, Woo JC, Kim SJ, Yoon BI, Lee SJ, Cho YH: Preventive effect of selenium on chronic bacterial prostatitis. *J Infect Chemother*, 18, 30-34, 2012. DOI: 10.1007/s10156-011-0276-4

142. Wang C, Wang H, Luo J, Hu Y, Wei L, Duan M, He H: Selenium deficiency impairs host innate immune response and induces susceptibility to *Listeria monocytogenes* infection. *BMC Immunol*, 10:55, 2009. DOI: 10.1186/1471-2172-10-55

143. Hall JA, Vorachek WR, Stewart WC, Gorman ME, Mosher WD, Pirelli GJ, Bobe G: Selenium supplementation restores innate and humoral immune responses in footrot-affected sheep. *PLoS ONE*, 8 (12):e82572, 2013. DOI: 10.1371/journal.pone.0082572

144. Smith KL, Hogan JS, Weiss WP: Dietary vitamin E and selenium affect mastitis and milk quality. *J Anim Sci*, 75 (6): 1659-1665, 1997. DOI: 10.2527/1997.7561659x

145. Arriaga CR, Gonzales AP, Reina M, Galano A: Computer-designed melatonin derivatives: potent peroxyl radical scavengers with no prooxidant behavior. *Theor Chem Acc*, 139:133, 2020. DOI: 10.1007/s00214-020-02641-9

146. Sharman EH, Bondy SC: Melatonin: A Safe Nutraceutical and Clinical Agent. Nutraceuticals Efficacy, Safety and Toxicity, 501-509, Academic Press, 2016.

147. Galano A, Tan DX, Reiter RJ: Melatonin: A versatile protector against oxidative DNA damage. *Molecules*, 23 (3): 530, 2018. DOI: 10.3390/molecules23030530

148. Farias MS, Budni P, Ribeiro CM, Parisotto EB, Dias JF, Dalmarco EM, Fröde TS, Pedrosa RC, Filho DW: Antioxidant supplementation attenuates oxidative stress in chronic hepatitis C patients. *Gastroenterol Hepatol*, 35 (6): 386-394, 2012. DOI: 10.1016/j.gastrohep.2012.03.004

149. Huang Z, Rose AH, Hoffmann PR: The role of selenium in inflammation and immunity: From molecular mechanisms to therapeutic opportunities. *Antioxid Redox Signal* 16 (7): 705-743, 2012. DOI: 10.1089/ars.2011.4145

150. Guillin OM, Vindry C, Ohlmann T, Chavatte L: Selenium, selenoproteins and viral infection. *Nutrients*, 11:2101, 2019. DOI: 10.3390/nu11092101

151. Harthill M: Review: micronutrient selenium deficiency influences evolution of some viral infectious diseases. *Biol Trace Elem Res*, 143, 1325-1336, 2011. DOI: 10.1007/s12011-011-8977-1

152. Abba Y, Hassim H, Hamzah H, Noordin MM: Antiviral activity of resveratrol against human and animal viruses. *Adv Virol*, 2015:184241, 2015. DOI: 10.1155/2015/184241

153. Filardo S, Pietro MD, Mastromarino P, Sessa R: Therapeutic potential of resveratrol against emerging respiratory viral infections. *Pharmacol Ther*, 214:107613, 2020. DOI: 10.1016/j.pharmthera.2020.107613

154. Carr AC, Maggini S: Vitamin C and immune function. *Nutrients*, 9 (11): 1211, 2017. DOI: 10.3390/nu9111211

155. Geiler J, Michaelis M, Naczek P, Leutz A, Langer K, Doerr HW, Cinatl J: N-acetyl-L-cysteine (NAC) inhibits virus replication and expression of pro-inflammatory molecules in A549 cells infected with highly pathogenic H5N1 influenza A virus. *Biochem Pharmacol*, 79 (3): 413-20, 2010. DOI: 10.1016/j.bcp.2009.08.025

156. Van Hecke O, Lee J: N-acetylcysteine: A rapid review of the evidence for effectiveness in treating COVID-19. *CEBM*, 2020.

157. Jariwalla RJ, Roomi MW, Gangapurkar B, Kalinovsky T, Niedzwiecki A, Rath M: Suppression of influenza A virus nuclear antigen production and neuraminidase activity by a nutrient mixture containing ascorbic acid, green tea extract and amino acids. *Biofactors*, 31, 1-5, 2007. DOI: 10.1002/biof.5520310101

158. Dey S, Bishayi B: Killing of *S. aureus* in murine peritoneal macrophages by ascorbic acid along with antibiotics chloramphenicol or ofloxacin: Correlation with inflammation. *Microb Pathog*, 115, 239-250, 2018. DOI: 10.1016/j.micpath.2017.12.048

159. Zhu L, Yuan C, Zhang D, Ma Y, Ding X, Zhu G: BHV1 induced oxidative stress contributes to mitochondrial dysfunction inMDBK cell. *Vet Res*, 47:47, 2016. DOI: 10.1186/s13567-016-0332-2

160. Rehman ZU, Che L, Ren S, Liao Y, Qiu X, Yu S, Sun Y, Tan L, Song C, Liu W, Ding Z, Munir M, Nair V, Meng C, Ding C: Supplementation of vitamin E protects chickens from Newcastle Disease Virus mediated exacerbation of intestinal oxidative stress and tissue damage. *Cell Physiol Biochem*, 47, 1655-1666, 2018. DOI: 10.1159/000490984

161. Sebastiano M, Eens M, Messina S, AbdElgawad H, Pineau K, Beemster G TS, Chastel O, Costantin D: Resveratrol supplementation reduces oxidative stress and modulates the immune response in free-living animals during a viral infection. *Funct Ecol*, 32, 2509-2519, 2018. DOI: 10.1111/1365-2435.13195

162. Paracha UZ, Fatima K, Alqahtani M, Chaudhary A, Abuzenadah A, Damanhuri G, Qadri I: Oxidative stress and hepatitis C virus. *Virology*, 10:251, 2013. DOI: 10.1186/1743-422X-10-251