Optimisation of Indirect ELISA by Comparison of Different Antigen Preparations for Detection of Antibodies Against Schmallenberg Virus [1][2]

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Abstract

Schmallenberg virus (SBV) infection, discovered in 2011, was reported in Europe including Turkey, Africa and recently in some Asian countries. Commercial enzyme-linked immunosorbent assay (ELISA) kits were widely used by researchers in many epidemiological studies and SBV diagnosis. The aim of this study was to optimise indirect in-house ELISA that is based on different antigen preparations of cell-culture derived whole SBV particle. Antigen preparations were maintained with various methods: PEG precipitation, ultracentrifugation, dialysis, and antigen inactivation. Following antigen optimisation, steps of antigen coating, blocking, conjugate and stop solution were optimised and in-house ELISA was compared to commercial indirect SBV ELISA kit. The best result in ELISA antigen preparation for SBV was gained by 30% PEG purification method followed by formaldehyde inactivation. Although results of this study demonstrated that in-house ELISA for detection of SBV specific antibodies was equally sensitive and specific as commercial kit, purified SBV antigen based in-house ELISA development could increase S/P ratios.

Keywords: Diagnosis, Dialysis, ELISA, PEG, Schmallenberg virus

INTRODUCTION

Schmallenberg virus (SBV), first identified in 2011 in Germany, is classified in Orthobunyavirus genus of Peribunyaviridae family. SBV causes abortion, stillbirths, and congenital malformation in ruminants [1,2]. Since 2011, SBV infection in wild and domestic ruminants was reported in many countries in Europe including Turkey [3-6]. SBV infection was found in some African and Asian countries recently [7-9]. Seroepidemiological data of SBV infection in Turkey...
revealed that SBV seropositivity in cattle, sheep, goats, and Anatolian water buffalo was 39.8%, 1.6%, 2.8%, and 1.5%, respectively [3]. SBV seropositivity in some European countries was estimated to be up to 98.5% in cattle, 89% in sheep, and 50.8% in goat [10]. SBV infection has been detected in both domestic and wild ruminants, including cattle, sheep, goat, buffalo, deer, and bison [3].

There are many assays developed for diagnosis of SBV infection to date. Reverse transcriptase PCR (RT-PCR), real-time RT-PCR assays, loop-mediated isothermal amplification (LAMP), and recombinase polymerase amplification (RPA) are developed for molecular diagnosis of SBV [11-13]. For detection of SBV specific antibodies, virus neutralisation, plaque reduction neutralisation, and enzyme-linked immunosorbent assay (ELISA) tests were used [14-17]. Indirect and competitive ELISA kits which are based on nucleoprotein (N) of SBV are commercially available for detection of SBV specific antibodies in serum, plasma and milk samples. The commercial kits are used by many researchers to determine SBV seropositivity for seroepidemiological surveys and diagnosis [15,18]. On the other hand, some researchers developed in-house ELISA to determine SBV specific antibody based on whole virus particle [16,17]. In this study it is aimed to optimise an indirect ELISA assay based on cell-culture derived whole viral particle for serological diagnosis of SBV infection by preparation and comparison of different SBV ELISA antigens.

**MATERIAL and METHODS**

**Schmallenberg Virus**

Schmallenberg virus isolate (strain F6; GenBank accessions: KC355457-KC355459) was kindly provided by Prof. Dr. Wim van der Poel (Wageningen Bioveterinary Research, Netherlands). SBV was propagated on Vero cells which were grown using Dulbecco modified Eagle medium (DMEM) with 10% foetal bovine serum (FBS) and incubated at 37°C with 5% CO₂. The viral titer was determined by plaque titration assay as described previously [19].

**Serum Samples**

Serum samples taken from 300 cattle in previous studies were tested with both commercial indirect ELISA kit (IDEXX, Westbrook, Maine, USA) and competitive ELISA kit (IDVet, Grabels, France) in order to determine SBV-seropositive and SBV-seronegative samples. Both commercial kits were used as following the instructions of the manufacturers. The positive and negative sera according to results of these commercial kits were accepted as SBV-seropositive and SBV-seronegative samples. In-house ELISA results were compared to commercial indirect ELISA results.

**Preparation of ELISA Antigen**

Vero cells were infected with SBV and cells were frozen after 80% or over of cells showed cytopathic effect. Freeze-thaw process was maintained for 2-3 times. Different methods which have implemented and/or not been implemented before such as, polyethylene glycol (PEG) precipitation, ultracentrifugation, dialysis, and inactivation for antigen preparation were carried out.

**Precipitation with 50% PEG:** 50% PEG 8000 (w/v) (Sigma, Missouri, USA) and 23% NaCl (w/v) (Merck, Darmstadt, Germany) were dissolved in distilled water. Supernatant of SBV infected cells was collected and centrifuged at 3000 rpm for 30 min at 4°C. The supernatant was mixed with 50% PEG and 23% NaCl solution was added to the mixture. Virus-PEG mixture was stirred at 100 rpm for 16 h at 4°C. The mixture was centrifuged at 12000 rpm for 30 min at 4°C. Pellet was suspended in 1× TNE buffer (pH 7.2) and centrifuged at 5500 rpm for 20 min at 4°C. After centrifugation supernatant was collected and centrifuged at 24000 rpm for 2 h at 4°C. Supernatant was discarded and the pellet was suspended in 1× TNE buffer and stored at -80°C until used.

**Precipitation with 30% PEG:** Viral supernatant was mixed at a ratio of 2:1 with 30% PEG 8000 (w/v) which was prepared in 0.4 M NaCl [19]. The mixture was incubated at 4°C overnight. The virus was recovered by centrifugation at 3200×g for 30 min at 4°C. Supernatant was discarded and the virus was suspended in PBS and stored at -80°C until used.

**Ultracentrifugation:** Supernatant of SBV infected cells was collected and centrifuged at 2000 rpm for 10 min at 4°C. After centrifugation, supernatant was collected and filtered with using 0.22 µm pore filter membranes. The filtrate was ultracentrifuged (in Department of Virology, Faculty of Veterinary Medicine, Firat University, Elazig, Turkey) using SW-28 rotor (Beckman Coulter, Brea, California, USA) at 25,000 rpm for 2 h at 4°C. DMEM containing 1% BSA was poured onto the viral pellet, incubated for 2 h at 4°C, and re-suspended.

**Commercial PEG precipitation kit:** SBV antigen was purified with PEG virus precipitation kit (Biovision, California, USA) according to manufacturer’s instruction. Briefly, cells infected with SBV were centrifuged at 3200×g for 15 min at 4°C. Supernatant was collected, mixed with 5× PEG solution and incubated overnight at 4°C. The mixture was centrifuged the following day at 3200×g for 30 min at 4°C. The viral pellet was suspended in virus suspension buffer and stored at -80°C until used.

**Dialysis:** Dialysis was carried out with two different systems: 12-14 kDa cut-off dialysis tubes (Biovision, California, USA) and Spectra/Por2 dialysis membrane 12-14 kDa MWCO (Spectrum, Waltham, Massachusetts, USA). The dialysis tubes were soaked with distilled water and the tubes were dialyzed against water overnight at 4°C and against PBS for 4 h at 4°C [17]. After dialysis, virus collected and stored at -80°C until used. The dialysis membrane was initially
soaked in distilled water for 30 min. The supernatant was
dialyzed against type I MilliQ water during day and over-
night at 4°C and water was changed at least 4 times. Final
dialysis was carried out against PBS overnight at 4°C [21].
The virus was collected and stored at -80°C until used.

**Antigen inactivation:** To inactivate SBV, the antigens were
treated by Triton X-100 with final concentration of 1% [17]
or formaldehyde with final concentration of 2% [22] and
incubated in room temperature for 1 h.

**SDS-PAGE and Western Blotting**

SDS-PAGE and western blotting was carried out to confirm
SBV isolate and antigen preparation method and to
determine SBV-seropositive serum samples were reacting
against viral proteins. For separation of protein suspensions,
protein electrophoresis was carried out in 10% SDS-PAGE
gel. The proteins were transferred onto a PVDF membrane
(Thermo Scientific, Waltham, Massachusetts, USA) and
the membrane was blocked overnight at 4°C with 5%
skimmed milk powder in phosphate buffered saline with
0.05% Tween-20 (PBST). SBV-seropositive and -seronegative
cattle sera were used as primary antibody and the membrane
was incubated in sera diluted 1:100 in 0.01% PBST at room
temperature for 2 h. Washing step was carried out with 0.1%
PBST for three times. Rabbit anti-bovine IgG HRP secondary
antibody (Life Technologies, Carlsbad, California, USA) diluted
1:1000 in 0.01% PBST was added onto the membrane and
incubated at room temperature for 1 h. Following
washing with 0.1% PBST, TMB substrate (Amresco, Solon,
Ohio, USA) was poured onto the membrane, incubated for 5 min and evaluated with pre-stained protein marker
(Thermo Scientific, Waltham, Massachusetts, USA).

**In House Indirect ELISA Optimisation**

Concentrations of differently prepared antigens were
measured by Bradford protein assay kit (Thermo Fisher
Scientific, Waltham, Massachusetts, USA). In-house indirect
ELISA was optimised by performing checkerboard titrations
of antigen and conjugate using positive and negative
serum samples which were defined by commercial indirect
and competitive ELISA kits. Further optimisations
of SBV antigen, coating buffer, blocking, diluents, and
stop solution were implemented for the in-house indirect
ELISA. SBV antigen was diluted in both 0.05 M carbonate/
bicarbonate buffer and PBS. Fifty µL of antigen solution
at 10 µg/mL concentration was coated onto polystyrene
ELISA plates by incubating at 4°C overnight. Undiluted sera,
1/2, 1/10 and 1/100 dilutions of sera are tested for sample
step. The washing steps were carried out with 0.05% PBST.
Two different blocking agents, 10% skimmed milk powder and
1:10 diluted foetal bovine serum (FBS), were tried for
blocking step. The conjugate (Life Technologies, Carlsbad,
California, USA) was diluted in different diluents, such as
5% skimmed milk powder (in 0.05% PBST) and 3% FBS (in
0.05% PBST). TMB substrate was used and the reaction
was stopped by using 2 M sulphuric acid or 1% sodium
dodecyl sulphate (SDS). Results were read at 450 nm using
a spectrophotometer.

**RESULTS**

Three hundred cattle sera were tested by both commercial
indirect and competitive ELISA kits and 22 out of 300 sera
were positive for SBV specific antibody in both commercial
kits. Positive and negative sera were used for development
of in-house indirect ELISA. Differently prepared ELISA
antigens were tested by SDS-PAGE and western blotting
and SBV nucleoprotein (25 kDa) and Gc protein (110 kDa)
were detected with SBV seropositive serum (Fig. 1), whereas
SBV seronegative serum cannot detect any SBV proteins
by western blotting analysis (data not shown).

After checkerboard titrations of antigen and conjugate,
optimisation steps of SBV antigen, coating buffer, blocking

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Fig 1. Western blotting results of different SBV
antigen and bovine serum albumin (BSA). All
differently prepared SBV antigens (Lines 1-3) showed
Gc protein (110 kDa) and nucleoprotein (N) (25
kDa) bands in western blotting. M: Protein marker;
1: SBV antigen prepared by ultracentrifugation, 2:
SBV antigen prepared by PEG precipitation; 3: SBV
antigen gained from cell culture; 4: BSA (66.5 kDa)
as control
agent, and stop solution were implemented. SBV antigen was diluted at concentration of 10 µg/mL and coated successfully in PBS, not in 0.05 M carbonate/bicarbonate buffer. Antigen coating with PBS results in OD of 0.773±0.094 for positive sera and 0.241±0.098 for negative sera, whereas carbonate/bicarbonate buffer coating results in 0.4255±0.152 for positive sera and 0.234±0.067 for negative sera (Fig. 2-A). Because coating with PBS resulted in a higher OD value for positive sera, PBS was preferred for the antigen coating step.

To optimise blocking step of in-house ELISA, skimmed milk powder and FBS were tested. The positive sera OD was 0.579±0.001 and negative sera OD was 0.3045±0.0015 in blocking with 10% skimmed milk powder, whereas carbonate/bicarbonate buffer coating results in 0.4255±0.152 for positive sera and 0.234±0.067 for negative sera (Fig. 2-A). Because coating with PBS resulted in a higher OD value for positive sera, PBS was preferred for the antigen coating step.

For sample step, undiluted sera, 1/2 and 1/10 dilutions of sera gave same O.D. results, whereas 1/100 dilution O.D. was undetectable. Therefore, optimisation of sample step was implemented with 1/10 dilutions of sera.

Skimmed milk powder and FBS were used as diluents of the conjugate to optimise conjugate step of in-house ELISA. The optimisation of conjugate step was maintained in commercial indirect ELISA kit (IDEXX, Westbrook, Maine, USA) with following the instructions of the manufacturer, except for conjugate step which was carried out with three different conjugates. The first one is the conjugate of commercial kit (IDEXX), the second one is the conjugate (Life Technologies) diluted 1:2000 in 5% skimmed milk powder, and the third one is the conjugate (Life Technologies) diluted 1:2000 in 3% FBS (in 0.05% PBST). When the full procedure of commercial ELISA kit was followed, the OD of positive sera and negative sera were 1.5905±0.0355 and 0.2545±0.0125, respectively. Once the conjugate (Life Technologies) diluted 1:2000 in 5% skimmed milk powder, and the third one is the conjugate (Life Technologies) diluted 1:2000 in 3% FBS (in 0.05% PBST). When the full procedure of commercial ELISA kit was followed, the OD of positive sera and negative sera were 1.5905±0.0355 and 0.2545±0.0125, respectively. Once the conjugate (Life Technologies) diluted 1:2000 in 5% skimmed milk powder (in 0.05% PBST), the OD values were 0.5105±0.0125 for positive sera and 0.2455±0.0035 for negative sera. The most optimised results gained by 1:2000 dilution of the conjugate (Life Technologies) in 3% FBS (in 0.05% PBST), with OD values of 2.2125±0.0235 and 0.3155±0.0205 for positive and negative sera, respectively. ELISA reactions were stopped by adding 2M sulphuric acid, but not with 1% SDS.

### Table 1. The S/P% results of in-house indirect ELISA were calculated with formulation of indirect ELISA kit

<table>
<thead>
<tr>
<th>Antigens</th>
<th>Positive Sera, S/P% (mean±SD)</th>
<th>Negative Sera, S/P% (mean±SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>30% PEG</td>
<td>53.0±2.181</td>
<td>9.0±0.371</td>
</tr>
<tr>
<td>30% PEG and Triton X-100 inactivation</td>
<td>21.3±0.876</td>
<td>12.8±0.525</td>
</tr>
<tr>
<td>30% PEG and formaldehyde inactivation</td>
<td>69.9±2.877</td>
<td>-6.3±0.259</td>
</tr>
<tr>
<td>PEG kit</td>
<td>40.6±1.668</td>
<td>4.6±0.189</td>
</tr>
<tr>
<td>PEG kit and Triton X-100 inactivation</td>
<td>24.8±1.019</td>
<td>13.3±0.548</td>
</tr>
<tr>
<td>PEG kit and formaldehyde inactivation</td>
<td>60.4±2.483</td>
<td>1.0±0.042</td>
</tr>
</tbody>
</table>

The indirect ELISA kit results were calculated and interpreted as positive if S/P% ≥40%, as doubtful if 30% ≤ S/P% < 40%, and as negative if S/P% < 30%
To find the best antigen preparation system, 50% PEG and 30% PEG precipitation, ultracentrifugation, commercial PEG precipitation kit, dialysis, and antigen inactivation (Triton X-100 or formaldehyde) methods were implemented and compared to each other. Comparison of OD values of the different antigens indicated that the ELISA antigen gained by 30% PEG precipitation with formaldehyde inactivation method has the highest efficiency as having 0.961 and 0.149 mean OD values for positive and negative sera, respectively (Fig. 2-C). However, the result of commercial indirect ELISA kit has higher OD for positive sera (mean OD of 1.481). The S/P% values of different antigens-based in-house ELISA were given in Table 1.

Following optimisation of steps of in-house ELISA, results were compared with commercial indirect ELISA kit (IDEXX, Westbrook, Maine, USA) (Fig. 2-C). All positive and negative sera which were already determined with two commercial kits were positive and negative, respectively, with in-house ELISA; however, the OD values were higher in commercial indirect ELISA kit in comparison to OD values of in-house ELISA (Fig. 2-C). The sensitivity and specificity of in-house ELISA were determined as 100% when compared to commercial ELISA kits.

DISCUSSION

ELISA is one of the most commonly used serological techniques and the use of ELISA in serosurveys for viral diseases provides convenience to the scientists to assess the epidemiology, and rate of spread of the diseases. Since the first discovery of SBV in 2011, ELISA is widely used for SBV diagnosis and seroepidemiology. Commercial ELISA kits are available and in-house ELISAs are developed by some researchers. In present study, it is aimed to optimise indirect in-house ELISA for SBV antibody detection with assessing the results of different antigens and diluents which have not tested for SBV ELISA to date.

In the present study, different ELISA antigen preparation methods were implemented: precipitation with 50% PEG, precipitation with 30% PEG, commercial PEG precipitation kit, dialysis, and antigen inactivation with Triton X-100 or formaldehyde. Among these methods, the best ELISA antigen was prepared by 30% PEG purification following inactivation with formaldehyde in the present study. A previously used method for inactivation of SBV [17] with Triton X-100 has been resulted in low OD value when compared to formaldehyde inactivation method (Fig. 2-C). Besides the methods used in the present study, sucrose gradient, caesium chloride density gradient, ultrafiltration could be applied for virus purification. Sucrose gradient method is found to be better for foot-and-mouth disease virus [23]. Some combination of purification methods could increase viral yield, such as sucrose gradient following dialysis and ultrafiltration method found to have the best yield for norovirus [24]. In the present study some combination of methods were tested and the antigen gained by 30% PEG purification method in combination with formaldehyde inactivation increased OD results (Table 1). ELISA for detection of virus specific antibodies can be based on either whole virus [16,17] or recombinant protein of the virus [25,26]. In the present study, only whole SBV was implemented in indirect ELISA. Using the whole virus as antigen in ELISA may bring the risk of cross-reaction with other Orthobunyaviruses and this can be overcome by using pure SBV antigen.

ELISA antigen coating could be maintained by PBS and carbonate/bicarbonate buffer. It is known that using a coating buffer with pH of 1-2 units higher than the isoelectric point of the antigen could increase binding of the ELISA antigen [22]. Because isoelectric point of SBV still remains unknown, both PBS and carbonate/bicarbonate buffer were tested for in-house ELISA in the present study. Antigen coating step was successfully maintained in PBS in the present study, although other researchers used carbonate/bicarbonate buffer for coating of SBV antigen [16,17].

Foetal bovine serum, skimmed milk powder, normal goat serum, normal chicken serum, bovine serum albumin (BSA), and gelatine could be used as blocking agent in homemade ELISAs [27]. In this study blocking with 1:10 diluted FBS had the best results in comparison to 10% skimmed milk powder. Conjugate dilution buffer may differ among studies, e.g. PBS with 0.05% Tween 20, PBS with 0.05% Tween 80, 5% FBS, 1% BSA could be used for conjugate dilution [16,17,23,26]. Several diluents and concentrations were tried out for conjugate optimisation in the present study. Conjugate was diluted in 5% skimmed milk (in 0.05% PBST) and 3% FBS (in 0.05% PBST) with different concentrations. The most optimised result was gained by 1:2000 dilution of conjugate in 3% FBS (in 0.05% PBST).

In conclusion, an in-house ELISA for detection of SBV-specific antibodies was optimised with the antigen gained by 30% PEG purification following inactivation with formaldehyde. In further studies, different methods such as sucrose gradient, caesium chloride density gradient, and ultrafiltration can be tested for SBV ELISA antigen.

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

STATEMENTS AUTHORS CONTRIBUTIONS

AKA, EA designed the experiments, made serum samples, the preparation of ELISA antigen stages, and SDS-PAGE western blotting, optimization works and wrote the manuscript. AKA, made a substantial contribution to interpretation of data. All authors discussed the results and contributed to the final manuscript.
Different ELISA Antigens to Detect SBV Antibodies

REFERENCES


