Moraxella ovis and Mycoplasma conjunctivae Isolation from an Ovine Infectious Keratoconjunctivitis Outbreak and Fortified Treatment Approaches

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Abstract

The aim of this study was to determine the etiology of naturally occurring infectious keratoconjunctivitis (IKC) outbreak observed in a Tuj sheep herd rearing semi-intensively and to try fortified treatment options in symptomatic sheeps. Conjunctival samples from 42 sheep with keratoconjunctivitis were collected aseptically using sterile cotton swabs. All swab were cultivated for bacterial isolation. To test treatment options symptomatic animals were randomly divided into 3 groups each were composed of adult ewes. The first group (n=13) was treated with the subconjunctival administration of penicillin G potassium (250.000 IU/animal), the second group (n=14) was treated with oxytetracycline and polymyxin B sulfate, and the third group (n=15) was treated with amikacin sulfate (diluted with isotonic NaCl to 25 mg per mL), ceftriaxone disodium and fluconazole (2 mg/mL) combination. As the results of the bacteriological examination, 36 (85.71%) of 42 conjunctival samples were found positive for Moraxella ovis. Also, Mycoplasma conjunctivae was identified in 3 (7.14%) conjunctival samples positive for M. ovis. After treatment, 84.61% (11/13) of the first group; 71.42% (10/14) of the second group and all animals in the third group were observed to be healed. This study demonstrated that M. ovis was the primary pathogen causing IKC in Tuj sheeps and M. conjunctivae was the second rank. The combination of amikacin sulfate, ceftriaxone disodium and fluconazole have been identified as the most effective treatment option of IKC in sheep.

Keywords: Keratoconjunctivitis, Sheep, Moraxella ovis, Mycoplasma conjunctivae

How to Cite This Article


Bir İnfeksiyöz Koyun Keratokonjonktivit Salgısından Moraxella ovis ve Mycoplasma conjunctivae İzolasyonu ve Güçlendirilmiş Tedavi Yaklaşımı

Öz

Bu çalışmada, semi-entansif olarak yetiştirilen bir Tuj koyun sürüsunde doğal olarak gözlenen bir infeksiyöz keratokonjonktivit (IKC) salgının etyolojisinin belirlenmesi ve semptom gösteren koyunlarla kuvvetlendirilmiş tedavi seçeneklerinin denenmesi amaçlanmıştır. Klinik keratokonjonktivit belirtisi olan 42 koyunun konjunktival örnekleri, steril sıvap kullanılarak aseptik koşularda toplanmıştır. Tüm örnekler bakteri izolasyonu için kültüre edildi. Tedavi seçeneklerini test etmek için erin koyunlardan oluşan semptomatik hayvanlar rastgele 3 gruba ayrıldı. Birinci grupta (n=13) penisilin G potasyum (250.000 IU/hayvan) subkonjunktival uygulanarak, ikinci grupta (n=14) oksitetrasiklin ve polymixin B sulfat ve, üçüncü grupta ise (n=15) amikasin sulfat, seftriakson disonid (0.5 g) ve flukonazol (2 mg/mL) kombinasyonu damlatma şeklinde uygulanarak tedavi denemeleri yapıldı. Bakteriyolojik muayene sonucunda, 42 konjunktival swab örneklerinin %85.71’inde Moraxella ovis ve %7.14’ünde Mycoplasma conjunctivae izole edildi. Tedavi sonrası, birinci grubun %84.61’si (11/13); ikinci grubun %71.42’si (10/14) ve üçüncü grubun ise tümünde iyileşme görüldü. Bu çalışma, M. ovis’in Tuj koyunlarında IKC’ye neden olan primer patojen, M. conjunctivae’nin ise sekonder etken olduğunu göstermiştir. Amikasin sulfat, seftriakson disoynid ve flukonazol kombinasyonu, koyunlarda IKC’nin en etkili tedavi seçeneği olarak tanımlanmıştır.

Anahtar sözcükler: Keratoconjunctivitis, Koyun, Moraxella ovis, Mycoplasma conjunctivae

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INTRODUCTION

Infectious keratoconjunctivitis (IKC), also known as pink eye disease of cattle, sheep and goats, is characterized by blepharospasm, conjunctivitis, lacrimation, and varying degrees of corneal opacity and ulceration [1]. In sheep and goats, naturally occurring conjunctivitis or keratoconjunctivitis can be associated with Chlamydia pecorum, Chlamydophila psittaci, Mycoplasma spp. (notably Mycoplasma conjunctivae), Moraxella ovis, Colesiota conjunctivae, Listeria monocytogenes, Acholeplasma oculi, Staphylococcus aureus, Corynebacterium conjunctivae, Staphylococcus oculi and Thelazia spp [2,3]. M. ovis has been implicated in epizootics of IKC in domestic sheep and goats [4]. Dagnall [5] reported that M. ovis could be isolated from both healthy sheep and those with IKC, but isolation occurred at a higher rate in sick animals. M. conjunctivae has been associated with most of the IKC outbreaks in small domestic ruminants [4] and wild caprinae worldwide [6] and is considered the primary pathogen of this infection. However, M. conjunctivae is commonly detected in the eyes of asymptomatic sheep and is eventually endemic in sheep herds throughout Europe [4]. IKC is a highly contagious disease and it is influenced by host and environmental factors. Predisposing factors such as age, breed, immune response and eye pigmentation, season (fly season), mechanical irritation (dust, grass, weeds, etc.), and concurrent presence of the disease-involved pathogenic bacteria in the environment influences the prevalence of this disease [7]. There is usually no mortality reported associated with IKC however, the morbidity rate can reach as high as 80% [1]. Loss of production are compounded by the cost of keratoconjunctivitis for producers in terms of incurring additional labour and treatment costs [8]. If the control and treat of the disease are not taken, it may spread in the flock and blindness may result and blind animals on range may subsequently die [9].

The effective antibiotic selection is important for the treatment of keratoconjunctivitis cases in animals. The fortified antibiotic treatments are reported to be extremely effective and reliable in the treatment of acute bacterial keratitis in human medicine [10,12]. Fortified preparations of ophthalmic antibiotics are made from antibiotics found in the market. These fortified eye drops have two main advantages, which are an increase in antibiotic concentration in the corneal stroma and a wide range option of availability. Given the potential for synergistic effects on corneal penetration, strong antibiotic combinations continue to be used in treatment for severe bacterial keratitis [13]. Fluconazole is a drug that is soluble in water and is suitable for topical application of epithelial defect. This drug with corneal penetration is effective against Candida sp. and Aspergillus sp in both human and animals [14-17]. Polymyxins have similar effect on fungi and can also be used against bacterial keratitis [18]. The combination of cephalosporins (ceftriaxone, cephaloz, ceftazidime-50 mg/mL) and aminoglycosides (tobramycin, gentamycin or amikacin) gives good results when used topically against polybacterial keratitis [19]. Penicillin and tetracycline susceptibility was reported to control M. ovis infection, especially to avoid exacerbation of lesions primarily caused by M. conjunctivae [20]. Additionally, animals with M. conjunctivae require antibiotic combination of oxytetracycline and polymyxin B, but not to penicillin [21].

In this study, it was aimed to determine the etiology of naturally occurring infectious keratoconjunctivitis and to try different treatment approaches in a Tuj sheep herd semi-intensively farming in Veterinary Faculty Farm of the Kafkas University in Kars, Turkey.

MATERIAL and METHODS

Ethical Approval

The experiment was carried out with the approval of Kafkas University Local Ethical Committee for Animal Experiments (KAÜ-HADYEK/2018-016).

Case Definition

The study was carried out on a naturally occurring infectious keratoconjunctivitis outbreak observed in a Tuj sheep herd rearing semi-intensively in the farm of the Faculty of Veterinary Medicine, Kafkas University, in July 2017. Out of 150 animals, 42 (28%) adult (>3 years) sheep were affected by the disease and thus subjected to the study. While all affected sheep had clinical complaints such as lacrimation and conjunctivitis, some had ulceration and corneal opacity in generally one eye in addition. But no subclassification was carried out on the severity of clinical presentation of animals. The fluorescent staining test was used in the detection of corneal ulcer together with clinical examination, and defects were controlled under the daylight.

Sampling

The conjunctival samples of each of 42 sheep with clinical complaints were collected aseptically using two sterile cotton swabs, and the samples was transferred within 1 h for process at the Microbiology Laboratory of Veterinary Faculty, Kafkas University, Turkey.

Bacterial Isolation

For the pre-enrichment progress of Mycoplasma sp., one of conjunctival swab samples was transferred in a 3 mL Mycoplasma broth (Oxoid, England) enriched with 10% fresh yeast extract (Oxoid, England), 20% heat-inactivated horse serum (Sigma, USA), 0.025% thallium acetate (Fluka, Belgium), and 100 IU of penicillin G (IE, Istanbul) and incubated at 37°C in a humidified atmosphere with 5% CO2 for 3 days. One hundred μL of pre-enriched content was then sub-cultured on Mycoplasma agar plates (Oxoid,
England) enriched with 10% fresh yeast extract, 20% heat-inactivated horse serum, 0.025% thallium acetate, and 100 U of penicillin G, and incubated again at 37°C with 5% CO₂ in a moist environment. These agar plates were incubated for up to 21 days and monitored every second day for signs of Mycoplasma growth [22]. The second samples were also inoculated on 7% sheep blood agar (Oxoid, England) plates and MacConkey agar (Oxoid, England) plates in order to determine the presence of other bacterial agents and incubated at 37°C, aerobically for two days. Identification of Mycoplasma isolates were carried out by conventional bacteriological methods such as catabolism of glucose, hydrolysis of arginine, phosphatase activity, tetrazolium reduction, serum digestion, digitonin sensitivity, film and spot formation [23]. The other causative agents were identified by using tests as catalase, oxidase, indole, methyl red, voges proskauer, citrate, nitrate reduction which were carried out as described earlier [24].

Clinical Trials

Affected animals were randomly distributed into three treatment groups. The subconjunctival penicillin G potassium (Ibrahim Ethem®) was administered three times (every other day) at the daily dose of 250,000 IU/animal to animals in the first group (n=13). The ocular ointment (Terramycin, Pfizer®) containing oxytetracycline and polymyxin B sulfate were applied to the animals in the second group (n=14) once per day in the lentil size area on the inside of the eyelid and treatment was continued for 1 week. Amikacin sulphate (Amikosit, Zentiva®) (diluted with isotonic NaCl to 25 mg per mL), ceftriaxone sodium (Unacefin 0.5 g, Avis®) and fluconazole (2 mg/mL) (Triflukan, Pfizer®) [19,25-27], for fungal infection probability was administered in combination with 2 drops three times per day for 1 weeks to animals in third group.

RESULTS

Culture Results

As the results of bacteriological examination, M. ovis was isolated and identified in 36 (85.71%) of 42 conjunctival samples. In addition, M. conjunctivae was identified in 3 (7.14%) conjunctival samples of ewe together with M. ovis. Six conjunctival swab samples were found bacterial culture negative. The other bacteria could not isolate from samples. All the treatment groups were included two culture negative animals, as well. The results of the cultivated samples and fortified treatment options were showed in Table 1.

Clinical Trial Results

The treatment groups were randomly adjusted as three groups. All the groups were included animals suffered from M. ovis and M. conjunctivae, except the first group which is infected only with M. ovis. After treatment, it was detected that 84.61% (11/13) of the animals in first group; 71.42% (10/14) of the animals in the second group and 100% (15/15) of the animals in the third group were observed to be healed (Fig. 1).

Table 1. The summary of the cultivated samples and fortified treatment options

<table>
<thead>
<tr>
<th>Total Number of Samples</th>
<th>Culture Results</th>
<th>Treatment Options (Healing Rate)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Positive Groups</td>
<td>Penicillin G</td>
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<tr>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>36 (36 M. ovis and 2 M. conjunctivae)</td>
<td>1&lt;sup&gt;st&lt;/sup&gt; group n: 13</td>
</tr>
<tr>
<td></td>
<td></td>
<td>2&lt;sup&gt;nd&lt;/sup&gt; group n: 14</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3&lt;sup&gt;rd&lt;/sup&gt; group n: 15</td>
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NA: Not applicable

Fig 1. While affected sheeps were exhibited corneal opacity, remarkable healing was observed after treatment
DISCUSSION

The etiology of infectious keratoconjunctivitis involves many predisposing factors such as age, race, daylight, dust irritations which facilitate the colonization of the pond are important in the formation of the disease [27]. Bacteria such as C. pecorum, Mycoplasma spp. (notably M. conjunctivae), Moraxella spp., C. psittaci and M. ovis are often blamed as causative agents of IKC in animals [2]. Moraxella bovis is reported to be the most common agent of infectious bovine keratoconjunctivitis in the world [8,20]. In contrast, the main microorganisms isolated from IKC of sheep lesions are M. conjunctivae, C. psittaci and M. ovis [3,28]. Karthik et al. [28] reported 2 Moraxella spp. were isolated from the eye swabs of 5 animals with keratoconjunctivitis in a sheep herd consist of 100 animals in India. Also, they found to be 99% compatible with M. bovis by PCR-based sequence analysis. The researchers showed that the isolates were susceptible to all antimicrobials as gamoxyxclav, enrofloxacin, ciprofloxacin, cefixime, gentamicin, tetracycline and norfloxacin. Åkerstedt and Hofshagen [3] conducted a study on Norwegian Sheep in which they took conjunctival swab from 43 sick and 42 healthy animals in 15 farms with ovine keratoconjunctivitis and 50 conjunctival swabs from sheep in 17 farms without disease and cultivated for the presence of Mycoplasma and other bacterial species. They reported that M. (Branhamella) ovis was isolated in 28% of the samples taken from the affected herds and 10% of the samples from healthy animals. In addition, M. conjunctivae was isolated in 16 (37%) animals with keratoconjunctivitis and 3 (7%) non-clinical animals from the farms where keratoconjunctivitis was observed, whereas M. conjunctivae was isolated in only 4 (8%) samples from farms without evidence of keratoconjunctivitis. L. monocytogenes (1%), S. aureus (5%), Corynebacterium spp. (2%) and E. coli (4%) were isolated in herd. The researchers has reported that it is the first time that M. conjunctivae was isolated from a conjunctivitis case in Norway and the other predisposing agents can be isolated such as M. (Branhamella) ovis and L. monocytogenes. They claimed that different agents may play a role in the formation of keratoconjunctivitis in the sheep, and that some are seconder invaders.

Shahzad et al. [20] have reported an IKC infection in a few Lohi lambs showing clinical symptoms as conjunctivitis, keratitis, severe lacrimation and varying degree of blindness in Livestock Experiment Station (LES), Bahadurnagar, Okara, Pakistan. Thirty six conjunctival swabs sampled from the animals were evaluated by cultural and molecular methods. Sixteen (44.44%) out of 36 samples showed turbidity in PPLO broth. Twelve (75%) out of 16 broth samples showed colony growth on PPLO agar and this isolates identified as M. conjunctivae. Twenty one (59%) out of 36 conjunctival direct swab samples were found positive for M. conjunctivae by polymerase chain reaction (PCR). Topical application of 0.5% sterile solution of gentamycin (100 mg/mL) (Gentafar 10%, FARVET, Netherlands) proved suitable for the treatment of IKC in Lohi lambs as all clinical signs of IKC disappeared after 5 days of treatment with this antibiotic. Taghavi Razavizadeh and Razmyar [31] have reported an IKC infection in sheep herds consisting of 300 ewes with ocular lesions in Mashhad, Iran. About 10% of the flock was affected by the disease and 6 animals with severe clinical symptoms were sampled for cultural process. Following inoculation of the ocular swab samples in PPLO broth and agar, turbidity and growth of colonies were observed in them, respectively. The causative agent of the outbreak has identified to be M. conjunctivae. This was also the first documented report of isolation and molecular characterization of M. conjunctivae in Iran. The affected sheep were succesfully treated with oxytetracycline 20%, IM (20 mg/kg) twice, with interval of 72 h, and flunixin meglumin, IM (2.2 mg/kg) three times, with interval of 24 h. In addition to the listed drugs, tylosin 20%, IM (17 mg/kg), was administrated three times for sheep with pulmonary involvement. Motha et al. [32] have performed direct PCR for examination of M. conjunctivae in samples taken form 5 sheep herds with infectious keratoconjunktivitis. In addition, the researchers have run ELISA for screening antibody produced against M. agalactiae and M. conjunctivae and CFT for antibody of M. capricolum, and Chlamydiophila spp. M. conjunctivae was detected specifically from conjunctival swab samples from 3 herds. Antibodies only against to M. conjunctivae antibodies were also detected in blood sera of animals (60/75) from all farms. M. (Branhemella) ovis was isolated from the remaining 2 herds. The researchers have reported that M. conjunctivae is the primary pathogen of IKC in sheep and that this is the first time that the agent has been reported in New Zealand. Fernandez-Aguilar et al. [4] have reported a study of IKC on sheep and goat population in Pakistan with quantitative PCR (qPCR) to investigate the presence of M. conjunctivae and Chlamydiaceae. The infection rate for M. conjunctivae has been reported as 19.3% for sheep and 9.5% for goat. In India, Vaid et al. [33] reported an outbreak of IKC in nomadic sheep herds and 3 M. ovis were isolated out of 6 cases of IKC in sheep.

In Turkey, Erdeğer et al. [34] reported that M. bovis was isolated from 41 (19.7%) of 208 conjunctival swab that were obtained from 168 cattle with IBK suspects. Samsar et al. [35] reported that M. bovis was isolated from the all of 51 (100%) symptomatic cattle with IBK. M. bovis was isolated from 26 (17.9%) of 145 cattle with IBK suspected cattle in Erzurum and central villages [36].

In this study, conjunctival swab samples from 42 (28%) animals with an epidemic keratoconjunctivitis in Taj sheep herd were collected and cultured for bacteriological examination. M. ovis was isolated from 36 (85.71%) of the samples, while M. conjunctivae (with M. ovis) was detected from 3 (7.14%). In this study, M. ovis was isolated from IKC infection similar to the results obtained in other
Mycoplasma and Moraxella profile were individually reported both from applications on existing cases. Nevertheless, susceptibility of isolated microorganisms and are lack of the clinical of antimicrobials are limited with susceptibility in vitro [11,13]. Fortified drugs are made from anti-biotics (parenteral or lyophilized preparations) on the market. The first advantage of these drugs is the increased antibiotic concentration in the corneal stroma when applied and the second advantage is being a wide selection chance [13]. Given the potential for synergistic and combined effects of corneal penetration and antibiotic association, fortified antibiotic drops for severe keratitis remain standard therapy [13]. Animal studies with fortified achatelnits applications especially in ocular infections are widely reported in cattle [37-39] but ovine cases are quite rare [32,40]. Kibar et al. [38] have reported mean time for healing of cornal ulcers and amelioration of clinical signs was significantly less for calves that received enrofloxacin or penicillin + streptomycin than for the untreated controls. Gokce et al. [37] reported that 30 M. bovis isolates were obtained from IBK outbreaks of calves and found that animals treated with florfenicol recovered more rapidly than the animals treated with oxytetracycline. However, studies addressing the effectiveness of clinical application of antimicrobials on animals infected with Moraxella spp. and Mycoplasma spp. are scarce in the literatures [20,28]. Most of antimicrobials are limited with in vitro susceptibility of isolated microorganisms and are lack of the clinical applications on existing cases. Nevertheless, susceptibility profile were individually reported both Moraxella spp. and Mycoplasma spp., as Moraxella species show susceptibility to ampicillin, cefitouf, enrofloxacin, florfenicol and gentamicin [20,42] and Mycoplasma species to tetracyclines, kanamycin, danofloxacin, tylosin and linco-spectin [39].

As the results of the treatment options in this study, it was found that the healing rates were higher in the first and third group, in which penicillin G and amikacin/ceftriaxone/ fluconazole combination were administered, respectively. When considering the cultivated agents (completely M. ovis, except 2 culture negative ewes) of the first group animals the penicillin effectiveness is expectable on Moraxella spp. However, the penicillin resistance is likely as reported before for M. ovis [20,28] and was encountered in the first group (11/13) of this study. Combined antibiotic therapy is widely practised in polyfactorial infectious disease and in that in order to limit the spread of antimicrobial resistance. The trilateral combination antimicrobials in the third group of this study yielded a maximum (15/15) healing of animals infected with M. ovis and M. conjunctivae. This can support a concomitant synergistic interaction between cephalosporins and aminoglycosides [43]. An anti-fungal (fluconazole) supplementation into the treatment may have contributed to the healing since the remaining two animals found as bacterial culture negative and a fungal infections are included in the probability. Oxytetracycline is usually the first choice for antimicrobial treatment of IKC [44]. Chapman et al. [40] has gained the greatest clinical improvement when sheep with clinical signs of IKC were treated with long-acting oxytetracycline with intramuscularly. In the present study, oxytetracycline and polymyxin B combination was administered in the second treatment group animals infected M. ovis and M. conjunctivae and 71.42% (10/14) of healing was obtained. Resistance of Moraxella spp. to oxytetracycline is scarcely reported [39], but the indiscriminate use of oxytetracycline (most typically is terramycin) in Kars province over the years can be related with the bacterial agent resistant to this drug. Furthermore, polymyxin B as a cationic decapeptide cell membrane destroyer of Gram-negative bacteria may be inadequate in treatment of Gram positive M. ovis.

As the result, determining the etiology of infectious keratoconjunctivitis in animals with appropriate microbiological methods and developing an ideal fortified treatment approach to implementation will reduce the size of estimated damages of disease and contribute to animal welfare.

REFERENCES


Moraxella ovis and Mycoplasma conjunctivae ...