

Determination of Some Acute Phase Proteins, Biochemical Parameters and Oxidative Stress in Sheep with Naturally Infected Sheeppox Virus

Kadir BOZUKLUHAN¹ Oğuz MERHAN² Halil İbrahim GÖKÇE³
Metin ÖĞÜN² Emine ATAKIŞI² Şemistan KIZILTEPE⁴ Gürbüz GÖKÇE⁴

^[1] This study was presented in "VIII. National Veterinary Biochemistry and Clinical Biochemistry Congress, September 22-24, 2016, Bursa/Turkey" as oral presentation

¹ University of Kafkas, Kars School of Higher Vocational Education, TR-36100 Kars - TURKEY

² University of Kafkas, Department of Biochemistry, Faculty of Veterinary Medicine, TR-36100 Kars - TURKEY

³ University of Mehmet Akif Ersoy, Department of Internal Medicine, Faculty of Veterinary Medicine, TR-15000 Burdur - TURKEY

⁴ University of Kafkas, Department of Internal Medicine, Faculty of Veterinary Medicine, TR-36100 Kars - TURKEY

Article Code: KVFD-2017-19167 Received: 05.12.2017 Accepted: 27.03.2018 Published Online: 27.03.2018

How to Cite This Article

Bozukluhan K, Merhan O, Gökçe Hİ, Öğün M, Atakışı E, Kiziltepe Ş, Gökçe G: Determination of some acute phase proteins, biochemical parameters and oxidative stress in sheep with naturally infected Sheeppox virus. *Kafkas Univ Vet Fak Derg*, 24 (3): 437-441, 2018. DOI: 10.9775/kvfd.2017.19167

Abstract

The aim of the present study was to determine oxidative stress and acute phase response in naturally infected sheep with poxvirus. For this purpose, 20 poxvirus infected and 10 clinically healthy sheep were used in the study. In these animals, alkaline phosphatase (ALP), aspartate aminotransferase (AST) activity, haptoglobin, ceruloplasmin, urea, creatinine, albumin, iron (Fe), malondialdehyde (MDA) and reduced glutathione (GSH) concentrations were determined. The present study determined that the concentrations of haptoglobin, ceruloplasmin, urea, creatinine, MDA and ALP, AST activity were significantly increased, and albumin, Fe and GSH concentrations were significantly decreased in the sheeppox virus infected group compared to the control group. Results of this study indicated that oxidative stress and acute phase response develop in sheep infected with poxvirus.

Keywords: Acute phase proteins, Biochemical parameters, Sheeppox

Poksvirüs İle Doğal Enfekte Koyunlarda, Bazı Akut Faz Proteinler, Biyokimyasal Parametreler ve Oksidatif Stresin Belirlenmesi

Öz

Çalışma poksvirüs ile doğal enfekte koyunlarda, oksidatif stres ve akut faz cevabı belirlemek amacıyla yapıldı. Çalışmada 20 adet çiçek hastalığı ile enfekte ve 10 adet klinik olarak sağlıklı koyun kullanıldı. Hayvanlarda alkalin fosfataz (ALP), aspartat aminotransferaz (AST) aktivitesi, haptoglobulin, seruloplazmin, üre, kreatinin, albümin, demir (Fe), malondialdehit (MDA) ve redükte glutatyon (GSH) konsantrasyonları belirlendi. Çalışmada, poksvirüs ile enfekte koyunlar ile kontrol grubundaki hayvanlar karşılaştırıldığında haptoglobulin, seruloplazmin, üre, kreatinin, MDA değerleri ile ALP ve AST aktivitelerinin yükseldiği, albümin, Fe ve GSH konsantrasyonunun ise kontrol grubuna göre düştüğü belirlendi. Bu çalışmada poksvirüs ile enfekte koyunlarda akut faz yanıtının oluştuğu ve oksidatif stresin geliştiği kanısına varıldı.

Anahtar sözcükler: Akut faz proteinler, Biyokimyasal parametreler, Koyun çiçeği

INTRODUCTION

Sheeppox is a serious, often fatal, disease characterized by widespread skin eruption and lung lesions in sheep and

goats. It is caused by a virus belong to poxviridae virus family and *Capripoxvirus* genus. The disease is common in southeastern Europe, Africa and Asia, including Turkey^[1-3]. Clinical signs vary from mild to severe, depending on



İletişim (Correspondence)



+90 474 2426807/5145



oguzmerhan@hotmail.com

host and viral factors. Fever and a variable degree of systemic disturbance develop. It is clinically manifested as conjunctivitis, rhinitis, and multiple non-vesicular swellings in skin and mucous membranes, respiratory distress, and death. Widespread skin lesions are seen on the muzzle, ears, and areas free of wool. In severe cases, lesions can also spread to the lungs, liver, kidneys and secondary pneumonia is commonly seen [1,3-6].

The acute phase response (APR) is a systemic reaction of the body to local or systemic disturbances in association with a wide variety of disorders, including infection, trauma, inflammatory diseases, various neoplasms and immunological disorder. Acute phase proteins (APP) are circulating blood proteins primarily synthesized by the liver in a few hours in response to inflammatory stimulus [7-10]. Acute phase proteins are defined as positive APP, which serum concentrations increase and negative APP, which serum concentrations decrease in response to APR [11,12]. Synthesis of APP is induced by proinflammatory cytokines such as interleukin-1 (IL-1), interleukin-6 (IL-6) and tumor necrosis factor- α (TNF- α), and they are released into the bloodstream as long as inflammatory stimulus persists. Acute phase proteins are considered as a part of the innate host defense system, and its quantification provides valuable clinical information in the early identification of inflammation or injury, diagnosis, monitoring health status and prognosis of various diseases. Furthermore, these proteins have been shown to be useful for differential diagnosis of clinical and subclinical diseases, bacterial and viral infections [9,10]. Acute phase proteins have been studied widely in human medicine, especially as biomarkers of inflammatory processes, various infections, to diagnose and monitor the outcome of the treatment of diseases. However, they have been relatively under-utilised in veterinary medicine. In veterinary medicine, APPs have been identified as useful diagnostic biomarkers of diseases in dogs, cats and cattle. However, the value of APPs as indicators of inflammatory conditions, monitoring animal health and detection of diseases in veterinary clinical practice, especially in farm animal medicine is not well-documented [11,12].

It is known that free radicals play an important role in the pathogenesis of various infectious diseases [13], and that poxvirus causes an increase in free radicals, affecting the membrane phospholipids and thus causing damage to cells and tissues [14]. In the organism, there is a balance between oxidants and antioxidants, and when the balance shifts in favour of oxidants, oxidative stress develops. Development of oxidative stress results in increased formation of free radicals and lipid peroxidation [15]. Malondialdehyde (MDA) is an important indicator of lipid peroxidation and is used as an indicator to determine the severity of tissue damage [16,17]. The system preventing damage caused by free radicals in the organism is called the antioxidant (Reduced Glutathione 'GSH') system [18].

It is well-known that the main lesions occur in the skin in poxvirus disease, and that many organ or system functions such as liver, kidney, respiratory and circulatory system are impaired as a result of systemic inflammatory response [4]. Acute phase proteins are synthesized from the liver as a consequence of APR in many viral, bacterial and parasitic diseases [9,10]. Therefore, the aim of the present study is to determine whether or not oxidative stress and APR develop in the naturally infected sheep with poxvirus.

MATERIAL and METHODS

Twenty, 1 to 2-year-old, naturally poxvirus-infected Mor-karaman sheep and 10 clinically healthy sheep were used. In poxvirus infected sheep, fever, loss of appetite, rhinitis, respiratory problems, and pox lesions under the tail, in the eyelids, axillae, mucous membranes and in between the back legs were observed. From these sheep, poxvirus was isolated and identified in the research laboratory (Erzurum Veterinary Control Institute-Turkey). The research work was carried out with the approval of the Institutional Ethics committee of Kafkas University, Faculty of Veterinary Medicine (KAU 2016/75). All sheep were routinely examined and the lesions, rectal temperatures, heart rates and respiratory rates were recorded. Blood samples were collected into the tubes with and without anticoagulant from the jugular veins. Serum ceruloplasmin concentrations were determined spectrophotometrically, developed by Colombo and Ricerich [19], while serum haptoglobin concentrations were determined by methods of Skinner et al. [20]. Whole blood GSH and serum MDA concentrations were measured using spectrophotometric methods as described by Beutler et al. [21], and Yoshioka et al. [22], respectively (UV-1201, Shimadzu, Japan). Serum alkaline phosphatase (ALP), aspartate aminotransferase (AST) activities, urea, creatinin, albumin, and Fe concentrations were determined using colorimetric commercial test kits (IBL, Turkey) according to the manufacturer's instructions (Epoch, Biotek, USA).

Statistical Analysis

SPSS for Windows 20.0 was used for the statistical analyses. The distributions of the data obtained from the groups were shown as normal distribution according to the Kolmogorov-Smirnov test. Therefore, Student's t-test was used for the comparison of the groups.

RESULTS

In infected animals, fever ($39.85 \pm 0.20^\circ\text{C}$), conjunctivitis, rhinitis, nasal discharge (serous and/or mucopurulent), lacrimation, depression, anorexia, swelling in the hairless areas (under the tail, eyelids, axillae etc.), coughing and coarsening of vesicular respiratory sounds, increase in heart rate ($98.0 \pm 3.5/\text{min}$), rapid and shallow respiration ($30.0 \pm 1.00/\text{min}$) were observed. There were no abnormal

Table 1. Acute phase proteins, biochemical and oxidative stress parameters in control and sheep with naturally infected Sheeppox virus. Data are presented as mean±standard error (X±SEM)

Parameters	Control	Infected	P
Haptoglobin (g/L)	0.138±0.008	0.647±0.020	P<0.01
Ceruloplasmin (mg/dL)	13.05±0.89	18.23±0.93	P<0.05
Albumin (g/dL)	3.26±0.15	2.45±0.16	P<0.01
ALP (U/L)	62.78±1.67	99.56±4.72	P<0.05
AST (U/L)	57.36±5.09	91.45±6.65	P<0.05
Urea (mmol/L)	6.87±0.33	8.96±0.43	P<0.01
Creatinin (µmol/L)	96.52±3.17	194.57±3.58	P<0.01
Fe (µg/dL)	116.08±8.26	72.12±3.26	P<0.01
MDA (nmol/mL)	1.89±0.17	2.87±0.24	P<0.01
GSH (mg/dL)	85.42±4.60	69.27±2.17	P<0.01

clinical findings in control group. In these animals, the mean rectal temperature, heart rate and respiratory rate were 38.9±0.03°C, 76.0±1.5/min. and 22.0±0.50/min. respectively. Poxvirus were also isolated and identified in the samples collected from infected sheep.

In the study, blood concentrations of haptoglobin (P<0.01), ceruloplasmin, ALP and AST activities (P<0.05), urea, creatinine and MDA (P<0.01) were significantly high in sheeppox virus infected-animals compared to those of control animals. While, blood levels of albumin, Fe and GSH (P<0.01) were significantly lower in infected group than that of control group (Table 1).

DISCUSSION

In agreement with literature [1,2,14], swelling in the hairless areas (under the tail, eyelids, axillae etc.), conjunctivitis, rhinitis, lacrimation, depression, nasal discharge (serous and/or mucopurulent) and anorexia etc. were found on clinical examination of sheep with naturally infected sheeppox virus.

The acute phase response is a very fast response, developing with increased and/or decreased concentrations of APP within a few hours, which remain altered as long as the inflammatory stimulus persists. For this reason, they can be used as early identification of inflammation or injury, diagnosis and prognosis of diseases. These proteins can also be used as indicators in the monitoring of animal health [9,10,12,23,24]. In this study, haptoglobin and ceruloplasmin concentration were increased and albumin concentration was decreased, indicating the development of APR in sheeppox infected sheep.

The main lesions occur in the skin in poxvirus disease, and that many organ or system functions such as liver and respiratory system are impaired as a result of systemic inflammatory response [4]. Acute phase proteins, a non-specific marker of tissue damage, are synthesized in the

liver as a result of the systemic inflammatory response developed in the organism. Haptoglobin, a marker of acute inflammation and tissue damage in ruminants, is a glycoprotein synthesized in the liver by pro-inflammatory cytokines [9,11,12,24]. Haptoglobin has been reported to be present in very low concentrations (<0.1 mg/mL) in healthy animals. But its concentrations increase and remain high in many bacterial, viral, parasitic and inflammatory diseases as long as the stimulus persists [7,25-28]. It is a nonspecific marker of tissue damage, and suggested to be used to determine prognosis and severity of the diseases. Its levels between 0.1-1 g/L, and >1 g/L suggested to indicate good and poor prognosis of the diseases, respectively [20,29]. In addition, haptoglobin concentrations of 0.2-0.4 g/L were defined as mild and 1-2 g/L as severe infection [20]. In a study in cattle with FMD, Merhan et al. [30] detected increased APP levels and they reported that APP concentrations might be a relationship between the severity of disease. In our study, serum haptoglobin levels were approximately 0.7 g/L in naturally infected sheep, and the disease prognosis was good, severity was moderate in according to reported studies [20,29]. In addition to these, all the infected sheep fully recovered and no cases of death were observed in the study, supporting to the good prognostic value of haptoglobin. Ceruloplasmin is considered as an indicator of the presence of infection [27,30,31]. It protects the cells against oxidative damage by reducing the effects of oxygen radicals, as well as increasing the phagocytic and antimicrobial efficiency of immune system cells [32]. Increased in haptoglobin and ceruloplasmin levels determined in the study were presumably due to increased activity in phagocytic cells and/or tissue damage and inflammation reported in poxvirus infection [4,14].

Albumin, a negative APP, is a indicator of liver function and, its blood concentrations decrease in case of anorexia, liver disorders and intestinal malabsorptions [27,33]. Furthermore, its concentration has been shown to decrease in tissue damage and inflammatory conditions, in association with

increased albumin catabolism [9,11]. In this study, albumin concentration was also decreased and it was thought that this might be caused by an increase in albumin catabolism due to liver cell damage, tissue damage and inflammation or anorexia, observed in poxvirus infected sheep.

Aminotransferases found in hepatocytes are considered to be an important indicator of hepatotoxicity in patients with elevated blood levels in liver injury. The relative activity of AST and ALP indicates the severity of liver damage [33,34]. Aspartate aminotransferase activity is increased in passive congestion, tension in proventriculus, muscle destruction and liver damage. Alkaline phosphatase activity is known to increase during the cholestasis, bone formation/repair, impaired hepatobiliary circulation and stress [4,33,34]. In addition, poxvirus has been reported to cause liver cell damage and cause changes in enzyme activities [4]. In the study, increased AST and ALP activities are most probably occurred due to liver cell damage and glucocorticoid activity and/or disease-induced stress [4,33]. Serum Fe concentration is reduced in malnutrition, APR, acute or chronic infections and chronic liver disease [33,35]. It was thought that, reduced Fe levels obtained in the study might be due to APR, liver disorders and anorexia, occurs in poxvirus infection [4,14]. Serum urea and creatinine are used to evaluate renal function [33]. However, its concentrations also increase during the infection, anorexia, high fever and increased protein catabolism [36]. In addition to these, systemic inflammation that develops in the infections, disturb kidney perfusion and impair its filtration capability, resulting with increase in blood urea and creatinine [37]. In this study, serum urea and creatinine levels were also increased and it was thought that this might be caused by an increase in protein catabolism or impaired renal function due to systemic inflammation.

Oxidative stress develops when the balance between oxidants and antioxidants shift in favour of oxidants [13,15,38]. Viral infections have been reported to cause damage to the cells and tissues via increased free radicals in host cells [13,38]. However, it is also stated that this increase is an important factor in body defence against viral infections and that pathogens are destroyed in this way [39]. Kirmizigul et al. [16] were reported that the increase in nitric oxide, MDA, total sialic acid, total oxidant capacities and total antioxidant capacity decreased in the study of natural infected sheep with poxvirus. In addition, Issi et al. [14] were reported that the increase in blood catalase activities, MDA and antioxidants (vit E, vit C and GSH) decreased in the study of natural infected sheep with poxvirus. In this study, MDA level was increased while GSH level was decreased, which was thought to be caused by free radicals formed by phagocytes, which are part of the stress and/or defence system in animals. In the present study, supported by the other reports [14,16], MDA levels were increased, while GSH levels were decreased, indicating the development of oxidative stress in sheep infected with poxvirus.

In conclusion, it is suggestive that oxidative stress and APR develop in sheep infected with poxvirus. Furthermore, both liver and kidney functions are also affected in the infected animals. We consider that the data obtained from this study might contribute to the understanding of the pathogenesis and diagnosis of the disease.

REFERENCES

- Beytut E:** Sheep pox virus induces proliferation of type II pneumocytes in the lungs. *J Comp Path*, 143, 132-141, 2010. DOI: 10.1016/j.jcpa.2010.01.014
- Plowright W, Macleod WG, Ferris RD:** The pathogenesis of sheep pox in the skin of sheep. *J Comp Path*, 146, 97-105, 2012. DOI: 10.1016/j.jcpa.2011.12.004
- Constable PD, Hinchcliff KW, Done SH, Grünberg W:** Sheeppox and goatpox. In, *Veterinary Medicine: A Textbook of the Diseases of Cattle, Horses, Sheep, Pigs, and Goats*. 11th ed., 1591-1593, Elsevier, China, 2017.
- Altinel C:** Investigation of some biochemical parameters in blood serum of sheep pox vaccinated and infected sheep. *Pendik Vet Microbiol J*, 24, 203-219, 1993.
- Bowden TR, Babiuk SL, Parkyn GR, Copps JS, Boyle DB:** Capripoxvirus tissue tropism and shedding: A quantitative study in experimentally infected sheep and goats. *Virology*, 371, 380-393, 2008. DOI: 10.1016/j.virol.2007.10.002
- Buller RM, Palumbo GJ:** Poxvirus pathogenesis. *Microbiol Rev*, 55, 80-122, 1991.
- Çenesiz M, Sağan Öztürk A, Dalğın D, Yarım GF, Çiftçi G, Özdemir R, Güzel M, Kazak F, Çenesiz S:** Investigation of acute phase reactants and antioxidant capacity in calves infected with *Cryptosporidium parvum*. *Kafkas Univ Vet Fak Derg*, 23, 481-485, 2017. DOI: 10.9775/kvfd.2016.17183
- Cray C, Zaias J, Altman NH:** Acute phase response in animals: A review. *Comp Med*, 59, 517-526, 2009.
- Petersen HH, Nielsen JP, Heegaard PMH:** Application of acute phase protein measurements in veterinary clinical chemistry. *Vet Res*, 35, 163-187, 2004. DOI: 10.1051/vetres:2004002
- Toussaint MJM, van Ederen AM, Gruys E:** Implication of clinical pathology in assessment of animal health and in animal production and meat inspection. *Comp Haematol Internat*, 5, 149-157, 1995. DOI: 10.1007/BF00368037
- Gruys E, Toussaint MJM, Niewold TA, Koopmans SJ:** Acute phase reaction and acute phase proteins. *J Zhejiang Univ Sci B*, 6, 1045-1056, 2005.
- Tothova C, Nagy O, Kovac G:** Acute phase proteins and their use in the diagnosis of diseases in ruminants: A review. *Vet Med*, 59, 163-180, 2014.
- Akaike T:** Role of free radicals in viral pathogenesis and mutation. *Rev Med Virol*, 11, 87-101, 2001. DOI: 10.1002/rmv.303
- Issi M, Gul Y, Yilmaz S:** Clinical, haematological and antioxidant status in naturally poxvirus infected sheep. *Revue Med Vet*, 159, 54-58, 2008.
- Birben E, Sahiner UM, Sackesen C, Erzurum S, Kalayci O:** Oxidative stress and antioxidant defense. *World Allergy Organ J*, 5, 9-19, 2012.
- Kirmizigul AH, Ogun M, Ozen H, Erkilic EE, Gokce E, Karaman M, Kukurt A:** Oxidative stress and total sialic acid levels in sheep naturally infected with pox virus. *Pak Vet J*, 36, 312-315, 2016.
- Nisbet C, Cenesiz S, Acici M, Umur S:** Determination of the serum malondialdehyde, ceruloplasmin, adenosine deaminase levels in cattle with cystic echinococcosis. *J Fac Vet Med Univ Erciyes*, 5, 1-4, 2008.
- Sezer K, Keskin M:** Role of the free oxygen radicals on the pathogenesis of the diseases. *FÜ Sağ Bil Vet Derg*, 28, 49-56, 2014.
- Colombo JP, Richterich R:** Zur bestimmung des caeruloplasmin im plasma [on the determination of ceruloplasmin in plasma]. *Schweiz Med Wochenschr*, 23, 715-720, 1964.
- Skinner JG, Brown RA, Roberts L:** Bovine haptoglobin response in

clinically defined field conditions. *Vet Rec*, 128, 147-149, 1991. DOI: 10.1136/vr.128.7.147

21. Beutler E, Duron O, Kelly BM: Improved method for the determination of blood glutathione. *J Lab Clin Med*, 61, 882-888, 1963.

22. Yoshioka T, Kawada K, Shimada T, Mori M: Lipid peroxidation in maternal and cord blood and protective mechanism against activated-oxygen toxicity in the blood. *Am J Obstet Gynecol*, 135, 372-376, 1979. DOI: 10.1016/0002-9378(79)90708-7

23. Murata H, Shimada N, Yoshioka M: Current research on acute phase proteins in veterinary diagnosis: An overview. *Vet J*, 168, 28-40, 2004. DOI: 10.1016/S1090-0233(03)00119-9

24. Pradeep M: Application of acute phase proteins as biomarkers in modern veterinary practice. *Ind J Vet Anim Sci Res*, 43, 1-13, 2014.

25. Merhan O, Bozukluhan K, Gokce HI: Acute phase proteins and biochemical and oxidative stress parameters in *Hypoderma spp.* infested cattle. *J Hellenic Vet Med Soc*, 68, 535-540, 2017. DOI:10.12681/jhvms.16049

26. Kuru M, Merhan O, Kaya S, Oral H, Kukurt A: The effect of short term progesterone-releasing intravaginal device treatment on acute inflammation markers for Holstein heifers. *Rev Med Vet*, 166 (11-12): 336-340, 2015.

27. Kaya S, Merhan O, Kacar C, Colak A, Bozukluhan K: Determination of ceruloplasmin, some other acute phase proteins, and biochemical parameters in cows with endometritis. *Veterinary World*, 9, 1056-1062, 2016. DOI: 10.14202/vetworld.2016.1056-1062

28. Bozukluhan K, Merhan O, Özcan A, Gökçe Hİ, Gökçe G: Investigation of the levels of serum haptoglobin, oxidative indicators and some biochemical parameters in calves naturally infected with *Toxocara vitulorum*. *Vet J Ankara Univ*, 64, 75-79, 2017. DOI: 10.1501/vetfak_0000002778

29. Eckersall PD, Conner JG: Bovine and canine acute phase proteins.

Vet Res Commun, 12, 169-178, 1988.

30. Merhan O, Bozukluhan K, Kiziltepe S, Gokce HI: Investigation of levels of haptoglobin, serum amyloid A, ceruloplasmin and albumin in cattle with foot-and-mouth disease. *Isr J Vet Med*, 72, 14-17, 2017.

31. Fagliari JJ, McClenahan D, Evanson OA, Weiss DJ: Changes in plasma protein concentrations in ponies with experimentally induced alimentary laminitis. *Am J Vet Res*, 59, 1234-1237, 1998.

32. Cerone SI, Sansinanea AS, Streitenberger SA, Garcia MC, Auza NJ: Cytochrome c oxidase, Cu, Zn-superoxide dismutase, and ceruloplasmin activities in copper-deficient bovines. *Biol Trace Elem Res*, 73, 269-278, 2000. DOI: 10.1385/BTER:73:3:269

33. Tennant BC, Center SA: Hepatic function. In, Kaneko JJ, Harvey JW, Bruss ML (Eds): *Clinical Biochemistry of Domestic Animals*. 6th ed., 379-412, Academic Press, New York, 2008.

34. Russell KE, Roussel AJ: Evaluation of the ruminant serum chemistry profile. *Vet Clin North Am: Food Anim Pract*, 23, 403-426, 2007. DOI: 10.1016/j.cvfa.2007.07.003

35. Gruys E, Obwolo MJ, Toussaint MJM: Diagnostic significance of the major acute phase proteins in veterinary clinical chemistry: A review. *Vet Bull*, 64, 1009-1018, 1994.

36. Gokce HI, Woldehiwet Z: The effects of *Ehrlichia (Cytoecetes) phagochytophila* on the clinical chemistry of sheep and goats. *J Vet Med*, 46, 93-103, 1999. DOI: 10.1111/j.0931-1793.1999.00210.x

37. Aral H: Laboratory in diagnosis and monitoring of systemic inflammatory response syndrome or sepsis. *Eur J Basic Med Sci*, 5, 10-20, 2015. DOI: 10.15197/sabad.2.5.02

38. Lykkesfeldt J, Svendsen O: Oxidants and antioxidants in disease: Oxidative stress in farm animals. *Vet J*, 173, 502-511, 2007. DOI: 10.1016/j.tvjl.2006.06.005

39. Akaike T, Maeda H: Nitric oxide and virus infection. *Immunology*, 101, 300-308, 2000. DOI: 10.1046/j.1365-2567.2000.00142.x