Research Article

Molecular Analysis and Associated Risk Factors of *Theileria annulata* in Cattle from Various Zones of Balochistan, Pakistan

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How to cite this article?

Kebzai F, Ashraf K, Rehman MU, Akbar H, Avais M: Molecular analysis and associated risk factors of *Theileria annulata* in cattle from various zones of Balochistan, Pakistan. *Kafkas Univ Vet Fak Derg*, 30 (1): 15-21, 2024.

DOI: 10.9775/kvfd.2023.30151

Article ID: KVFD-2023-30151 Received: 28.06.2023 Accepted: 25.11.2023 Published Online: 12.12.2023

Abstract

Theileriosis is a protozoan parasite that is transmitted by ticks and infects a wide range of animals worldwide. This study aims to assess the molecular prevalence and related risk factors of theileriosis in Balochistan, Pakistan. Standard microscopy methods, polymerase chain reaction (PCR), 18S small subunit ribosomal RNA gene sequencing, and phylogenetic analysis were used. For this purpose, a total of 408 blood samples were collected from tick-infested cattle in Zhob, Loralai, and Quetta districts of Balochistan, Pakistan. Microscopy and subsequent PCR analysis confirmed the highest prevalence of Theileria annulata in Loralai district (11.76% and 12.75%), followed by Zhob district (11.27% and 12.25%), and Quetta district (8.34% and 9.56%), respectively. Moreover, the prevalence of *T. annulata* was higher in young cattle (85.82%), followed by female cattle (58.87%), and exotic crossbred cattle (33.33%) in the study area. However, various variable such as sex, area, and breeds of the cattle were not significantly correlated (P>0.05) with the presence of *T. annulata*, except for the age of animals (P<0.05). In addition, sequencing and phylogenetic analyses revealed that the isolated T. annulata was closely related to the isolates from Türkiye, Italy, Egypt, Iran, and Pakistan. Hence, these findings will contribute to the development of more effective control strategies for theileriosis in the cattle population of Balochistan, as well as in Pakistan on a broader scale.

Keywords: Balochistan, Cattle, Phylogeny, Prevalence, Risk factors, Theileria annulata

INTRODUCTION

Agriculture is the backbone of Pakistan's economy, with livestock contributing 14.04% to the country's overall national GDP^[1]. In Balochitsan province, 85% of population is directly or indirectly linked to livestock, which contributes over 45% to the province's GDP^[2]. The cattle population in Pakistan is around 51.5 million, playing a significant role in animal husbandry for milk, meat, and hides^[3]. Livestock is the main source of livelihood for the local people in Balochistan, especially cattle breeding, which serves as a key source of income^[4]. A recent study has revealed that Ixodid ticks have a substantial impact on infesting livestock and transmitting a wide range of pathogens in Pakistan. Lack of farmer awareness about the threat of theileriosis and the presence of tick vectors

on pasture and animals are key factors leading to severe economic losses in the livestock industry ^[5].

Ixodidae ticks can transmit parasitic, viral, bacterial, and rickettsial diseases to animals and humans, and affect the host in several ways, including reduced body weight, milk production, reproduction, and reduced quality of hides ^[6]. Similarly, tropical theileriosis in cattle is caused by a tick-borne haemoprotozoan parasite *Theileria annulata*, which is transmitted by ticks of the genus *Hyalomma*, and the Ixodidae ^[7]. Tropical theileriosis is frequently reported from North Africa, Europe, Middle East, India, and Central Asia, including Pakistan ^[8]. In Pakistan, *T. annulata* is transmitted by *Hyalomma anatolicum* to bovine animals ^[1]. While, *T. annulata* infection is characterized by high fever, anorexia, enlargement of

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superficial lymph nodes, anemia, hemoglobinuria, ocularnasal discharges, cessation of rumination, and diarrhea in cattle ^[9]. Henceforth, *T. annulata* not only affects the health of animals but also causes serious economic losses in terms of vaccination, treatment, reduction in milk yield and live weight of the animal, while it also increases the inter-calving interval, delay in the age of maturity and causes up to 70% mortality in infected animals ^[10].

In Balochistan, parasitic infections are the most prevalent among all other animal diseases ^[11]. This could be attributed to the poor healthcare system, open grazing practices, and inadequate animal management, resulting in significant economic losses ^[11]. In addition, there is scarce information on the molecular identification of parasitic species in the cattle population of Balochistan. Therefore, this study was designed to investigate and characterize the molecular prevalence of theileriosis and its associated risk factors in different zones of Balochistan, Pakistan. The aim is to develop comprehensive control and therapeutic strategies for theileriosis infection in cattle.

MATERIAL AND METHODS

Ethics Statement

This experimental design was duly approved by departmental ethics committee and directorate of advanced studies, University of Veterinary and Animal Sciences (UVAS), Lahore (ID: DAS/2053).

Study Area, Study Animals and Blood Sampling

A total of 408 blood samples were collected aseptically from tick-infested cattle in Balochistan between July 2021 and November 2022. The sampling was conducted randomly, irrespective to the sex, age, and breed of the cattle, from three different zones in Balochistan, Pakistan. Briefly, 136 blood samples were collected from the 1st zone, Suleiman mountainous region "Zhob", while 153 blood samples were collected from the 2nd zone, Northern highlands "Loralai",



and 119 blood samples were collected from the 3rd zone, Kachhi basin region "Quetta/Sibi" (Shoaib et al.^[12]). The sampling area is depicted in *Fig. 1*. The sample size was determined using the formula provided by Thrusfield;

 $N = [Z^2 p (1 - p)] / d^2$

 $N = (1.96)^{2*} 0.5(1-0.5) / (0.05)^2 = 384$

Where, N is the desired sample size, P is the predicted prevalence, and d is the 5% precision level.

As mentioned above, the sample size was determined to be 384. However, a total of 408 blood samples were collected in the study area to increase the level of accuracy. A standard performa was used to collect relevant details, such as, age, breed, area, and sex, pertaining to various risk factors ^[13]. Foe each cattle, five (5) mL of blood was drawn from the jugular vein and placed in vacutainers with EDTA. The blood samples were transported in a cold chain to the Parasitology Laboratory of UVAS, Lahore, and stored at -20°C till further analysis.

Microscopic Analysis

Microscopic analysis was conducted by preparing thin blood smears from the collected samples. The smears were preserved using absolute ethanol and stained with the field's stain A and the field's stain B, as previously described by ^[14]. Subsequently, the smears were observed under a compound microscope, Olympus CX21 (Olympus, Tokyo, Japan), at a magnification of 100x to detect the presence of *Theileria*.

DNA Extraction and Molecular Analysis

Genomic DNA was extracted from the blood samples using a DNA extraction kit (Thermo Fisher; Cat. No: FABGK001-2), following the manufacturer's instructions. The purity and concentration of extracted DNA were measured using a NanoDrop at 260/280 nm and stored at -20°C for further PCR analysis.

All of the extracted DNA samples were subsequently subjected to PCR, targeting the *T. annulata* cytochrome b gene fragment using specific primers, as previously described by Farooqi et al.^[8]. The forward primer used was 5'-ACTTTGGCCGTAATGTTAAAC-3', and the reverse primer was 5'- CTCTGGACCAACTGTTTGG-3', resulting in an amplification size of 450 base pairs.

For each PCR sample, a 20 μ L reaction mixture was prepared, consisting of 2 μ L of extracted DNA, 25 pmol of each primer, 200 mM of each dNTP, 5 μ L of 10X PCR buffer, and 1.5 mM MgCl2 (Promega, Madison, WI, USA), and 1.5 U of Taq DNA polymerase. The first phase of the PCR reaction cycle invovled an initial denaturation step at 94°C for 5 min, followed by thermal degradation for 40 cycles at 94°C for 35 sec, annealing at 51°C for 35



sec, and extension at 72°C for 35 sec. The final elongation was performed for 10 min at 72°C, followed by a hold step at 4°C. Each PCR run included a positive control (lab-isolated clinical isolate) and a negative control (sterile water). Positive bands were confirmed using gel electrophoresis and then trimmed for sequencing analysis.

Sequencing Analysis

The PCR bands testing positive for haemoparasites (constituting 50% of the bands positive for *T. annulata*-specific primers) were excised from a 1.5% agarose gel and purified using a gel extraction kit (WizPrepTM; Ref. W70150-300), following the manufacturer's instructions. The purified DNA samples were sent to CELEMICS Product & Services in South Korea for sequencing. A phylogenetic tree was constructed using Mega 11 software with a maximum likelihood algorithm and bootstrapping at 1000 replications.

The PCR bands testing positive for haemoparasites (constituting 50% of the bands positive for *T. annulata*-specific primers) were excised from a 1.5% agarose gel and purified using a gel extraction kit (WizPrepTM; Ref. W70150-300), following the manufacturer's instructions.

Statistical Analysis

Statistical analysis was conducted using SPSS version 20.00. The prevalence of theileriosis was compared to independent variables such as sex, age, area, and breed using the chi-square (χ^2) test. A P-value <0.05 was considered statistically significant, and a 95% confidence interval was established.

RESULTS

Molecular Epidemiology of Theileriosis in cattle

The results of this study showed an overall 34.56% (141/408) PCR-based prevalence of theileriosis in cattle from 3 different zones of Balochitsan, Pakistan i.e. Loralai having a prevalence of 12.75%, Zhob with 12.25% and Quetta with 9.56% (*Fig. 2*). However, microscopic examination indicated a prevalence of 31.37% (128/408),

 Table 1. Results of the microscopic identification of Theileria species in cattle from three different zones of Balochistan, Pakistan (n=408)

Sample No.	Districts	Positive n (%)	Negative n (%)	Total n (%)	
1	Quetta	34 (8.34)	85 (20.83)	119 (29.17)	
2	Zhob	46 (11.27)	90 (22.06)	136 (33.33)	
3	Loralai	48 (11.76)	105 (25.74)	153 (37.50)	
Total		128 (31.37)	280 (68.63)	408 (100)	

Table 2. PCR based identification of the Theileria species in cattle from 3different zones of Balochistan, Pakistan (n=408)						
Sample No.	Districts	Positive n (%)	0			
1	Quetta	39 (9.56)	80 (19.61)	119 (29.17)		
2	Zhob	50 (12.25)	86 (21.08)	136 (33.33)		
3	Loralai	52 (12.75)	101 (24.75)	153 (37.50)		
Total		141 (34.56)	267 (65.44)	408 (100)		

suggesting that the PCR method is more sensitive compared to microscopic identification (*Table 1, Table 2*). In addition, the presence of intraerythrocytic inclusion



Fig 3. Thin blood smear of cattle samples collected from 3 different livestock zones of Balochistan, Pakistan (n=408), showing typical *Theileria* piroplasms, intraerythrocytic bodies, under oil immersion lens after field staining

Parameters		Quetta, (n=39) % Positive	Zhob, (n=50) % Positive	Loralai, (n=52) % Positive	Total (n=408)		Chi-square Value	P-value
					%Positive %Negative (n=141) (n=267)			
Breed	Sahiwal	20.51	16.00	13.46	16.31	18.73	6.085	0.808
	Friesian	33.34	30.00	23.08	28.38	22.85		
	Red Sindhi	7.69	6.00	9.62	7.80	8.99		
	Cholistani	7.69	4.00	7.69	6.38	11.98		
	Cross Bred	20.51	36.00	40.38	33.33	26.22		
	Jersey	10.26	8.00	5.77	7.80	11.23		
Age	Young (1-2 year)	97.44	82	80.77	85.82	28.84	6.012	0.049*
	Adult (>2 year)	2.56	18	19.23	14.18	71.16		
Sex	Male	56.41	36	34.62	41.13	37.08	- 5.215	0.074
	Female	43.59	64	65.38	58.87	62.92		
Area	+Ve	9.56	12.25	12.75	34.56	-	0.402	0.704
	-Ve	19.61	21.08	24.75	- 65.44		0.483	0.786

bodies resembling *Theileria* was also confirmed under a light microscope using thin blood smears (*Fig. 3*).

As shown in Table 3, the prevalence of T. annulata was higher in young cattle (85.82%), followed by female cattle (58.87%), and exotic crossbred cattle (33.33%) in the studied area. However, various variable such as sex, area, and breeds of the cattle were not significantly correlated (P>0.05) with the presence of *T. annulata*, except for the age of the animals (P<0.05). Exotic cattle breeds, such as Friesian and crossbred, had a higher prevalence of theileriosis compared to local breeds like Sahiwal, Red Sindhi, or Cholistani. This may be attributed to the local breeds being more acclimatized to the conditions in the area. In addition, young animals (0-2 years) were found to be more susceptible to theileriosis compared to adult animals (P<0.05). Female cattle also exhibited a higher prevalence of theileriosis compared to males (P>0.05). These findings suggest that young animals are at a higher risk of theileriosis, compared to adult animals.

Sequencing Analysis of *Theileria* 18S rRNA Gene in Cattle

PCR positive samples were sequenced to analyze the 18S rRNA gene fragments of *Theileria*. The sequences were subjected to analysis using tools such as BLAST & CLUSTAL W alignments. Nucleotide sequences of the *Theileria* 18S rRNA gene were obtained from the NCBI GenBank for comparison. Phylogenetic analysis and BLAST queries revealed a 99% sequence similarity





to *T. annulata* 18S rRNA gene from various countries including Pakistan (MT318159), Iran (MN960099), Italy (MT341858), Türkiye (MK918607), and Egypt (MN227666). Notably, some of the sequences obtained in this study have been submitted to the NCBI GenBank (*Fig. 4*).

DISCUSSION

Tropical theileriosis is a well-known hindrance to the growth of the dairy industry worldwide and causes significant economic losses to the livestock industry ^[15,16]. However, there is a lack of sufficient and up-to-date molecular epidemiological studies on tick-borne

infections, particularly theileriosis, which hinders the development of preventive and control strategies ^[16]. In Balochistan, where a majority of the people rely on livestock for their livelihood, tick-borne diseases pose serious risks to the livestock industry ^[17]. Therefore, this study was designed to investigate the molecular prevalence and relevant risk factors of theileriosis in cattle from three different zones of Balochistan: Zhob, Loralai, and Quetta. Initial examination of thin blood smears from the sampled animals showed intra-erythrocytic bodies of theileriosis under a light microscope, consistent with previous findings Ekoka Mbassi et al.^[14], Adehan et al.^[18] and Chauhan et al.^[19], which also observed similar intra-erythrocytic bodies in the blood smears of the *Theileria* positive animals.

In this study, a PCR-based prevalence of theileriosis was observed in cattle from 3 different zones of Balochitsan, Pakistan i.e. Loralai (12.75%), Zhob (12.25%) and Quetta (9.56%), accounting for a total of 34.56%. These results were in line with a study conducted by Durrani et al.^[20], which reported a PCR based prevalence of Theileria in cattle from China to be 32.4%. Similarly, the prevalence of T. annulata in cattle from North-Western Pakistan was found to be 29.9% [21]. However, microscopic examination revealed a lower prevalence of theileriosis, specifically 31.37% (128/408), compared to PCR based prevalence. This findings aligns with previous reports by Chen et al.^[22], Zeb et al.^[23], and Zaman et al.^[24], suggesting that microscopic identification is less precise and sensitive compared to PCR analysis. Therefore, PCR is considered the preferred tool for identifying blood-born parasites ^[25-,27].

In the present study, age of the animals was statistically significant risk factor for the occurrence of theileriosis in cattle of Balochitsan (P<0.05). However, other risk factors such as sex, area, and breeds of animals were statistically non-significant (P>0.05). These results were consistent with the reports of Mohammed-Ahmed et al.^[28], Farhan et al.^[29], and Kawan ^[30], suggesting that the occurrence of theileriosis in different zones may be linked with various known or unknown variables. Furthermore, the prevalence of theileriosis was observed to be higher in crossbred and Friesian cattle compared to Sahiwal, Red Sindhi, and, Cholistani, breeds in Balochitsan. These findings were similar to the results reported by Ghafar et al.[31] and Rana et al.[32], which indicated a higher prevalence of T. annulata in crossbred and exotic breeds of cattle compared to local breeds. Collectively, these findings indicate that exotic breeds and their crossbreeds face an increased susceptibility to tick-borne diseases [33,34]. This vulnerability may be attributed to inherent resistance, grooming behavior, or other biological factors influencing the interaction between native breeds and ticks. Furthermore, it is crucial to explore environmental

conditions, climatic factors, and other ecological parameters that could contribute to a more stable or unstable enzootic state in various areas of Balochistan ^[33,34].

In addition, this study showed that female and young cattle (0-2 years) appeared to have a higher prevalence of Theileria than male and adult cattle in Balochistan. These findings were consistent with the results reported by Aslam et al.^[35] and Farooq et al.^[36], who also observed a higher prevalence of T. annulata in younger animals. However, Valente et al.^[37] found no significant correlation between sex and the presence of theileriosis in cattle. On the other hand, Marwaha et al.^[38] observed that the prevalence of theileriosis was higher in adults compared to young animals, possibly due to the higher immunity level in young animals caused by fetal hemoglobin in their circulatory system^[39]. Moreover, the increased occurrence of theileriosis in younger animals, as observed in this study and others, might be linked to the larger body surface area of adult cattle, their outdoor management, prolonged movement over long distances in search of feed and water. In addition, the hormonal issues in female animals could potentially contribute to the higher prevalence of haemoparasitic infections in female cattle^[40].

Finally, BLAST and phylogenetic analysis of the study isolates revealed a 99% sequence similarity with the *Theileria* isolates from Italy, Pakistan, Egypt, Iran, and Türkiye. The 18S ribosomal RNA gene sequence of studied isolates was aligned with similar reference sequences, showing significant variation in the genotype of *T. annulata* from 3 different zones of Balochistan, Pakistan. Similar studies on the sequencing and phylogenetic characterization of *T. annulata* have been conducted by Sisson et al.^[41], Gargano et al.^[42], and Solomon and Tanga ^[43], which are closely related to this study. Hence, it is important to highlight that the sequence similarities among globally discovered haemoparasites are primarily due to the export and import of animals ^[44].

The present study concludes that *T. annulata* is the main species causing theileriosis in the cattle populations of Balochistan, Pakistan. Several risk factors such as sex, area, and breeds of the animals, were not significantly associated with the disease dynamics, except for the age of the animals. Consequently, young animals are more susceptible to suffering from theileriosis. Overall, these results will be beneficial in designing more effective control strategies for theileriosis and provide a baseline for further research to eradicate or decrease haemoparasites in the future.

Availability of Data and Materials

The datasets generated during and analyzed during the current study are available from the corresponding author (K. Ashraf & M.U. Rehman) on reasonable request.

Acknowledgements

The authors are thankful to molecular parasitology laboratory, University of Veterinary and Animal Sciences, Lahore, for the provision of the laboratory during the study. The authors are also grateful to the staff of civil veterinary hospital at Quetta, Zhob, and Loralai for their assistance in collecting blood samples.

Funding Support

N/A

Conflicts of interest

All other authors declare no conflicts of interest

Ethical Statement

This experimental design was duly approved by departmental ethics committee and directorate of advanced studies, University of Veterinary and Animal Sciences (UVAS), Lahore (ID: DAS/2053).

Author Contributions

KA and MUR contributed to the conceptualization, design, funding and supervision of the study. FK conducted all experiments and wrote the first draft of the manuscript. HA and MA collected, analyzed and interpreted the data. All authors contributed to the critical revision of the manuscript and have read and approved the final version.

References

1. Tariq M: Opportunities for improving feed use efficiency for sustainable dairy production in Pakistan. *Proceedings*, 73:11, 2020. DOI: 10.3390/ IECA2020-08 826

2. Durrani H, Syed A, Khan A, Tareen A, Durrani NA, Khwajakhail BA: Understanding farmers' risk perception to drought vulnerability in Balochistan, Pakistan. *AIMS Agric Food*, 6 (1): 82-105, 2021. DOI: 10.3934/ agrfood.2021006

3. Asif M, Ben Said M, Vinueza RL, Leon R, Ahmad N, Parveen A, Khan A, Ejaz A, Ali M, Khan AU: Seasonal investigation of *Anaplasma marginale* infection in Pakistani cattle reveals hematological and biochemical changes, multiple associated risk factors and msp5 gene conservation. *Pathogens*, 11 (11):1261, 2022. DOI: 10.3390/pathogens1111261

4. Raziq A, Younas M, Rehman Z: Continuing education article prospects of livestock production in Balochistan. *Vet J*, 30 (3): 181-186, 2010.

5. Khan A, Muhammed AA, Nasreen N, Iqbal F, Cossio-Bayugar R, ali Sha SS, Alanazi AD, Zajac Z: Tick-borne haemoparasitic diseases in small ruminants in Pakistan: Current knowledge and future perspectives. *Saudi J Biol Sci*, 29 (4): 2014-2025, 2022. DOI: 10.1016/j.sjbs.2021.12.046

6. Hurtado OJB, Giraldo-Ríos C: Economic and health impact of the ticks in production animals. *Ticks Tick Borne Pathog*, 1-19, 2018. DOI: 10.5772/ intechopen.81167

7. Aydın N, Vatansever Z, Arslan MÖ: Molecular epidemiology of *Babesia* and *Theileria* Species in sheep in Kars region of Turkey. *Turkiye Parazitol Derg*, 46 (1): 20-27 2022. DOI: 10.4274/tpd.galenos.2021.09709

8. Farooqi S, Ijaz M, Saleem M, Rashid M, Ahmad S, Islam S, Aqib A, Khan A, Hussain K, Khan N: Prevalence and molecular diagnosis of *Theileria annulata* in bovine from three distincts zones of Khyber Pakhtunkhwa province, Pakistan. *J Animal Plant Sci*, 27 (6): 1836-1841, 2017.

9. Ziam H, Kernif T, Saidani K, Kelanemer R, Hammaz Z, Geysen D: Bovine piroplasmosis-anaplasmosis and clinical signs of tropical theileriosis in the plains of Djurdjura (north Algeria). *Vet Med Sci*, 6 (4): 720-729, 2020. DOI: 10.1002/vms3.305

10. Maharana BR, Tewari AK, Saravanan BC, Sudhakar NR: Important hemoprotozoan diseases of livestock: Challenges in current diagnostics and therapeutics: An update. *Vet World*, 9 (5): 487-495, 2016. DOI: 10.14202/

vetworld.2016.487-495

11. Khan MA, Khan M, Ahmad I, Anjum A, Durrani A, Hameed K, Kakar I, Wajid A, Ramazan M: Risk factors assessment and molecular characterization of *Theileria* in small ruminants of Balochistan. *J Anim Plant Sci*, 27, 1190-1196, 2017.

12. Thrusfield M: Veterinary Epidemiology. John Wiley & Sons, 2018.

13. Shoaib M, Rashid MI, Akbar H, Sheikh AA, Farooqi SH, Ayari-Akkari A, Yassin HM, Khan R: Molecular epidemiology and characterization of *Babesia bovis* in cattle of Khyber Pakhtunkhwa province, Pakistan. *Trop Anim Health Pro*, 54 (6): 1-8, 2022. DOI: 10.1007/s11250-022-03346-w

14. Ekoka Mbassi FA, Mombo-Ngoma G, Ndoumba WN, Yovo EK, Eberhardt KA, Ekoka Mbassi D, Adegnika AA, Agnandji ST, Bouyou-Akotet MK, Ramharter M, Zoleko-Manego R: Performance of field's stain compared with conventional giemsa stain for the rapid detection of blood Microfilariae in Gabon. *Am J Trop Med Hyg*, 107 (2): 383-387, 2022. DOI: 10.4269/ajtmh.22-0061

15. Zerek A, Erdem İ, Yaman M, Altuğ ME, Orkun Ö: Ixodid ticks (Ixodoidea: Ixodidae) infesting wild animals in Hatay, Türkiye. *Kafkas Univ Vet Fak Derg*, 29 (6): 641-647, 2023. DOI: 10.9775/kvfd.2023.30132

16. Ullah R, Shams S, Khan MA, Ayaz S, Akbar Nu, Din Qu, Khan A, Leon R, Zeb J: Epidemiology and molecular characterization of *Theileria annulata* in cattle from central Khyber Pakhtunkhwa, Pakistan. *PloS One*, 16 (9):e0249417, 2021. DOI: 10.1371/journal.pone.0249417

17. Garcia K, Weakley M, Do T, Mir S: Current and future molecular diagnostics of tick-borne diseases in cattle. *Vet Sci*, 9 (5): 241, 2022. DOI: 10.3390/vetsci9050241

18. Adehan SB, Adakal H, Gbinwoua D, Yokossi D, Zoungrana S, Toé P, Ouedraogo M, Gbaguidi AM, Adoligbé C, Fandohan AB, Hounmanou G, Glèlè Kakaï R, Farougou S, De Clercq EM: West African cattle farmers' perception of tick-borne diseases. *Ecohealth*, 15 (2): 437-449, 2018. DOI: 10.1007/s10393-018-1323-8

19. Chauhan H, Patel B, Bhagat A, Patel M, Patel S, Raval S, Panchasara H, Shrimali M, Patel A, Chandel B: Comparison of molecular and microscopic technique for detection of *Theileria annulata* from the field cases of cattle. *Vet World*, 8 (11): 1370-1374, 2015. DOI: 10.14202/ vetworld.2015.1370-1374

20. Durrani AZ, Mehmood N, Shakoori A: Comparison of three diagnostic methods for *Theileria annulata* in Sahiwal and Friesian cattle in Pakistan. *Pak J Zool*, 42, 467-472, 2010.

21. Yasein G, Ashraf K, Naveed U, Rashid MI, Shabbir MZ: First genetic evidence of *Trypanosoma theileri* in indigenous cattle in Southern Punjab province of Pakistan. *Pak Vet J*, 42 (3): 322-327, 2022. DOI: 10.29261/ pakvetj/2022.034

22. Chen Y, Chen YY, Liu G, Lyu C, Hu Y, An Q, Qiu HY, Zhao Q, Wang CR: Prevalence of *Theileria* in cattle in China: A systematic review and meta-analysis. *Microb Pathogenesis*, 162:105369, 2022. DOI: 10.1016/j. micpath.2021.105369

23. Zeb J, Shams S, Din IU, Ayaz S, Khan A, Nasreen N, Khan H, Khan MA, Senbill H: Molecular epidemiology and associated risk factors of *Anaplasma marginale* and *Theileria annulata* in cattle from North-western Pakistan. *Vet Parasitol*, 279:109044, 2020. DOI: 10.1016/j.vetpar.2020.109044

24. Zaman MA, Mehreen U, Qamar W, Qamar MF, Kashif M, Shahid Z, Abbas RZ: Brief account of bovine theileriosis prevalence in some South Asian countries. *Agrobiol Rec*, 2, 38-48, 2020. DOI: 10.47278/journal. abr/2020.010

25. Parveen A, Ashraf S, Aktas M, Ozubek S, Iqbal F: Molecular epidemiology of *Theileria annulata* infection of cattle in Layyah District, Pakistan. *Exp Applied Acarology*, 83, 461-473, 2021. DOI: 10.1007/s10493-021-00595-6

26. Anter RG: Molecular and microscopical identification of bovine *Theileria* species isolates in Sharkia Governorate, Egypt. *Egyptian Vet Med Soc Para J (EVMSPJ)*, 15 (1): 52-63, 2019. DOI: 10.21608/evmspj.2019.80818

27. Sudan V, Shanker D, Jaiswal A, Singh A, Pandey V: Standardization and validation of simple PCR, duplex PCR and RAPD in comparison to blood smear examination for diagnosing bovine tropical theileriosis. *Biologicals*, 46, 88-91, 2017. DOI: 10.1016/j.biologicals.2017.01.003

28. Mohammed-Ahmed G, Hassan S, El Hussein A, Salih D: Molecular, serological and parasitological survey of *Theileria annulata* in North Kordofan State, Sudan. *Vet Parasitol Reg Stud Reports*, 13, 24-29, 2018. DOI: 10.1016/j.vprsr.2018.03.006

29. Atif FA, Khan MS, Iqbal HJ, Arshad GM, Ashraf E, Ullah S: Prevalence of *Anaplasma marginale, Babesia bigemina* and *Theileria annulata* infections among cattle in Sargodha District, Pakistan. *African J Agri Res*, 7 (22): 3302-3307, 2012.

30. Kawan MH: Molecular surveillance and phylogenetic analysis of *Theileria annulata* in bovine at Baghdad city/Iraq. *Iraqi J Vet Med*, 43 (1): 93-101, 2019. DOI: 10.30539/iraqijvm.v43i1.479

31. Ghafar A, Gasser RB, Abbas T, Rehman A, Gauci CG, Jabbar A: Ticks and tick-borne diseases of bovines in a smallholder livestock context: The Pakistani example. *Adv Parasitol*, 114, 167-244, 2021. DOI: 10.1016/ bs.apar.2021.08.009

32. Rana HAA, Iftikhar M, Watto MA, Bilal MQ: Institutional role in coping livestock diseases on farm level in rural areas of Punjab, Pakistan. *Pak J Agri Sci*, 58 (4): 1411-1421, 2021. DOI: 10.21162/PAKJAS/21.1408

33. Selim A, Weir W, Khater H: Prevalence and risk factors associated with tropical theileriosis in Egyptian dairy cattle. *Vet World*, 15 (4): 919-924, 2022. DOI: 10.14202/vetworld.2022.919-924

34. Larcombe S, Kolte S, Ponnudurai G, Kurkure N, Magar S, Velusamy R, Rani N, Rubinibala B, Rekha B, Alagesan A: The impact of tick-borne pathogen infection in Indian bovines is determined by host type but not the genotype of *Theileria annulata*. *Infect Genet Evol*, 75:103972, 2019. DOI: 10.1016/j.meegid.2019.103972

35. Aslam F, Rehman MU, Saleem G, Ashraf K, Hafeez MA, Saqib M: Identification and molecular characterization of *Theileria annulata* with associated risk factors in naturally infected camels from selected districts in Punjab, Pakistan. *Pak Vet J*, 43 (1): 79-84, 2023. DOI: 10.29261/ pakvetj/2022.084

36. Farooq U, Tufani N, Malik H, Mir M: Clinical and morpho-molecular epidemiology of bovine theileriosis in Kashmir, India. *Ind J Ani Res*, 53 (3): 375-381, 2019. DOI: 10.18805/ijar.B-3512

37. Valente D, Dutra AP, Carolino N, Gomes J, Coelho AC, Espadinha P,

Pais J, Carolino I: Prevalence and risk factors associated with *Theileria* annulata infection in two bovine portuguese autochthonous breeds. *Pathogens*, 12 (5):669, 2023. DOI: 10.3390/pathogens12050669

38. Marwaha S, Brar B, Jain VK, Poonia R, Prasad M: Multiplex PCR for rapid differential diagnosis of co-prevalent species of *Theileria (Theileria annulata* and *Theileria orientalis)* in cattle. *Parasitol Res*, 122 (5): 1189-1197, 2023. DOI: 10.1007/s00436-023-07819-1

39. Onyiche TE, Suganuma K, Igarashi I, Yokoyama N, Xuan X, Thekisoe O: A review on equine piroplasmosis: Epidemiology, vector ecology, risk factors, host immunity, diagnosis and control. *Int J Environ Res Pub Health*, 16 (10):1736, 2019. DOI: 10.3390/ijerph16101736

40. Das D, Sarma K, Eregowda CG, Roychoudhury P, Rajesh JB, Behera P, Prasad H, Lalrinkima H, Aktar F, Bora N, Deka C, Thakur N, Tolenkhomba TC: Naturally occurring *Anaplasma marginale* infection in cattle: Molecular prevalence and associated risk factors, haemato-biochemical alterations, oxidant/antioxidant status and serum trace mineral levels. *Microb Pathog*, 167:105575, 2022. DOI: 10.1016/j.micpath.2022.105575

41. Sisson D, Beechler B, Jabbar A, Jolles A, Hufschmid J: Epidemiology of *Anaplasma marginale* and *Anaplasma centrale* infections in African buffalo (*Syncerus caffer*) from Kruger National Park, South Africa. *Int J Parasitol Parasites Wildl*, 21, 47-54, 2023. DOI: 10.1016/j.ijppaw.2023.04.005

42. Gargano V, Blanda V, Gambino D, La Russa F, Di Cataldo S, Gentile A, Schirò G, Torina A, Millán J, Vicari D: Serological survey and molecular characterization of Theileria annulata in Sicilian cattle. *Pathogens*, 10 (2): 101, 2021. DOI: 10.3390/pathogens10020101

43. Solomon A, Tanga BM: The first investigation of tick vectors and tickborne diseases in extensively managed cattle in Alle District, Southwestern Ethiopia. *Vet Med Int*, 2020: 8862289, 2020. DOI: 10.1155/2020/8862289

44. Zaman MA, Rafique A, Mehreen U, Mehnaz S, Atif FA, Abbas A, Hussain K, Raza MA, Altaf S, Siddique F, Omar M: Epidemiological investigation and development of loop mediated isothermal amplification for the diagnosis of ovine theileriosis. *Pak Vet J*, 42 (3): 370-375, 2022. DOI: 10.29261/pakvetj/2022.039